

# Regulation of the Transport of Vitellogenin by Heterotrimeric G-Proteins during Oogenesis of a Polychaete, *Pseudopotamilla ocellata*

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**Key Words:**

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Coelomic fluid protein (CP), a vitellogenin contained in the coelomic fluid of polychaetes, is transported by receptor-mediated endocytosis that is controlled by GTP-binding proteins. Transport of  $^{125}\text{I}$ -CP was markedly inhibited by  $\text{AlF}_4$  and toxins such as cholera toxin and pertussis toxin. These effects appear to be mediated by cAMP, since  $^{125}\text{I}$ -CP transport was also greatly inhibited by dibutyryl cAMP. The results strongly suggest that heterotrimeric G-protein is involved in the regulation of  $^{125}\text{I}$ -CP transport through the activation of adenylyl cyclase. Immunoblotting tests with antibodies against  $\text{Gs}_\alpha$  and  $\text{Gi}_\alpha$  subunits showed a  $\text{Gs}_\alpha$  subunit of 45 kDa in the membrane of oocytes of intermediate and large size classes and a  $\text{Gi}_\alpha$  subunit of 41 kDa only in the oocytes of the intermediate size class.

Transport of vitellogenin into the oocytes of the polychaete, *Pseudopotamilla ocellata*, is carried out in specific stages by receptor-mediated endocytosis (Lee and Kim, 1993; Lee et al., 1997). GTP-binding proteins were found to be involved not only in the vesicular protein transport (Helms, 1995) and in the endocytosis of proteins (Brunskill et al., 1996) but also in the transport of vitellogenin during oogenesis of *Xenopus* and the polychaete (Schmalzing, 1994; Nam et al., 1996; Mukhopadhyay et al., 1997). Heterotrimeric G-proteins have been known to accelerate maturation of oocytes in sea urchin and starfish (Chiba et al., 1993; Jaffe et al., 1993) and to be involved in controlling fertilization in *Xenopus* (Downs et al., 1992; Tang, 1992) and early developmental stages (Allworth et al., 1991).

Toxins such as cholera toxin (CTX) and pertussis toxin (PTX) and aluminum fluoride ( $\text{AlF}_4$ ) have been used to explore functional aspects of G-proteins, since these toxins and  $\text{AlF}_4$  show different effects on G-proteins. In general, PTX acts on inhibitory G-protein ( $\text{Gi}$ ), whereas CTX acts on the  $\alpha$ -subunit of stimulatory G-protein ( $\text{Gs}$ ), affecting the sensitivity of G-protein in GTP (Allworth et al., 1991). For instance, the association of Gs protein with endosome fusion was found using CTX, and the association of G-protein with oocyte maturation in starfish was shown using PTX (Shilling, 1989; Colombo et al., 1994). The effects of toxins have been known to be dependent on the level of cAMP in some physiological phenomena such as exocytosis in sea urchin eggs (Turner et al., 1987),

endosome internalization in rat liver cells (Janicot et al., 1991) and oocyte maturation in *Xenopus* (Gallo et al., 1995). Aluminum fluoride has been known to activate heterotrimeric G-protein (Katada et al., 1984; Blackmore et al., 1985; Kanaho et al., 1985; Strnad and Wong, 1985; Gilman, 1987).

In the present paper we studied the effects of cholera toxin and pertussis toxin on the receptor-mediated transport of vitellogenin specifically into the oocytes of the intermediate size class so that we could characterize the G-protein involved in the transport and explore an association of the regulation of vitellogenin transport with the elevation of cAMP.

## Materials and Methods

### Materials

Oocytes of *Pseudopotamilla ocellata* Moore were collected on the east coast of Korea and kept in sea water at 10°C. Separation of oocytes into three size classes of small (30-80  $\mu\text{m}$  in diameter), intermediate (80-140  $\mu\text{m}$  in diameter) and large (180-200  $\mu\text{m}$  in diameter) was as described by Nam et al. (1996).

### Iodination of coelomic fluid proteins

Coelomic fluid was collected by rinsing the dissecting area of the female worms when dissected for the purpose of collection of oocytes. The proteins were extracted and iodinated as described by Lee and Kim (1993).

### Extraction and electrophoresis of membrane proteins

Membrane proteins were extracted from the membranes

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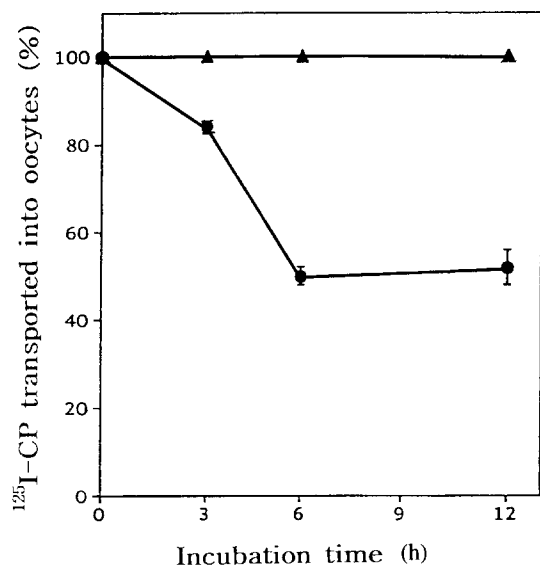


Fig. 1. Effects of AIF<sub>4</sub> on <sup>125</sup>I-CP transport into the oocytes of the intermediate size class. The oocytes were incubated with (●) or without (▲) AIF<sub>4</sub>. AIF<sub>4</sub> was prepared with 20 mM NaF and 10 μM AlCl<sub>3</sub>.

of the oocytes of the intermediate size class. Isolation of membranes, extraction of membrane proteins and gel electrophoresis were carried out as described by Lee (1988).

#### Immunoblotting

Membrane proteins of the oocytes were electrophoresed on 10% SDS-PAGE, transferred to nitrocellulose (NC) membrane from the gel and blocked with 5% non-fat dry milk at 4°C for 24 hours. NC membrane containing membrane proteins was washed three times with washing buffer (TBS + 0.1% Tween 20), each time for 10 minutes, and primarily incubated with rabbit polyclonal IgG in binding buffer (20 mM Tris-HCl, pH 8.0, 10 mM MgCl<sub>2</sub>, 2 mM dithiothreitol, 0.1% Triton X-100, 0.5% gelatin) and secondarily with peroxidase-labeled antibody. After incubation, the NC membrane was immersed in peroxidase-ECL solution (Amersham) for one minute and then exposed to X-ray film (XR O-mat, Kodak) for 3-5 minutes (Serafini, 1991).

#### Results

##### Effect of Aluminum fluoride on the transport of vitellogenin

Aluminum fluoride, a selective inhibitor of heterotrimeric G-protein, was introduced into the medium in which the oocytes of the intermediate size class were incubated with a labeled vitellogenin, <sup>125</sup>I-labeled coelomic fluid proteins (<sup>125</sup>I-CP), which were previously characterized by gel electrophoresis (Lee et al., 1997). Oocytes were incubated with 20 mM NaF and 10 μM AlCl<sub>3</sub> for 3

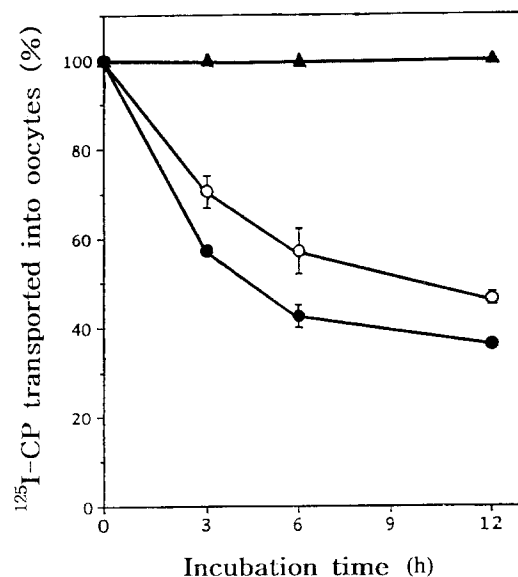


Fig. 2. Effects of cholera toxin on <sup>125</sup>I-CP transport into the oocytes of the intermediate size class. The oocytes were incubated with cholera toxin at the concentration of 2 μg/ml (○) and 4 μg/ml (●) or without (▲) cholera toxin.

hours at 12°C with gentle rotatory shaking, and then <sup>125</sup>I-CP of 4054 CPM/μl was introduced into the medium. The effect of AIF<sub>4</sub> on the transport of <sup>125</sup>I-CP into oocytes was remarkable: the transport was greatly reduced down to 50% of the control by the end of 6 hours of incubation without further reduction during subsequent incubation up to 12 hours (Fig. 1).

##### Effects of CTX and PTX on the transport of vitellogenin

CTX and PTX, both known to be bacterial toxins controlling different functions of G-proteins, were used to demonstrate the kind of G-protein associated with the transport of vitellogenin. The toxins were activated by incubating in 100 mM dithiothreitol at 37°C for 10 minutes.

The effect of CTX was assayed at two different concentrations of 2 μg/ml and 4 μg/ml. Fig. 2 shows that transport of <sup>125</sup>I-CP into the oocytes of intermediate size class drastically decreased at both concentrations with greater inhibition effect at the higher concentration. During 3 hours of incubation the transport decreased almost linearly with 43% reduction; it reached a plateau of 64% reduction by 12 hours.

The effect of PTX was similar to that of CTX but the reduction effect was not as great as that of CTX. The transport decreased with 40% reduction at 4 μg/ml for 12 hours (Fig. 3).

##### Effect of dbcAMP on the transport of vitellogenin

The effect of dibutyryl cAMP (dbcAMP) on the transport was also assayed in order to see whether the toxin

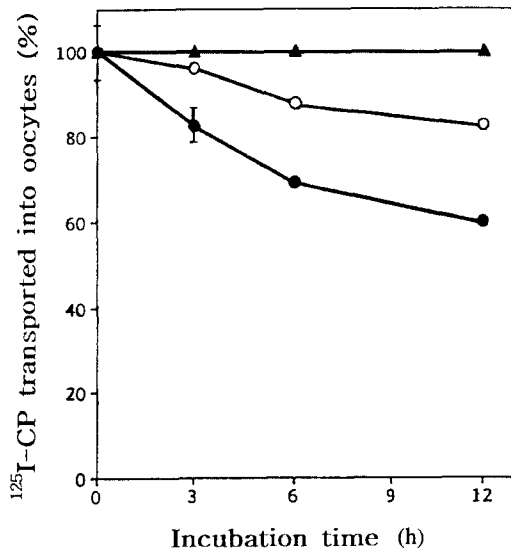


Fig. 3. Effects of pertussis toxin on <sup>125</sup>I-CP transport into the oocytes of the intermediate size class. The oocytes were incubated with pertussis toxin at the concentration of 2 μg/ml (○) and 4 μg/ml (●) or without (▲) pertussis toxin.

effects may result simply from the elevation of cAMP by activation of adenylate cyclase affected by G-protein. As shown in Fig. 4, transport of <sup>125</sup>I-CP was not affected at 50 mM dbcAMP but was greatly inhibited at 100 mM dbcAMP with more than 50% reduction.

Identification of G-proteins

From the studies on the effects of the toxins and AIF<sub>4</sub>

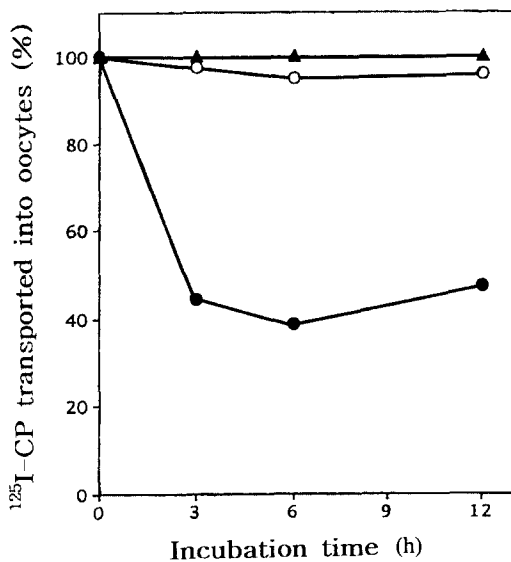


Fig. 4. Effects of dbcAMP on <sup>125</sup>I-CP transport into the oocytes of intermediate size class. The oocytes were incubated with dbcAMP at the concentration of 50 mM (○) and 100 mM (●) or without (▲) dbcAMP.

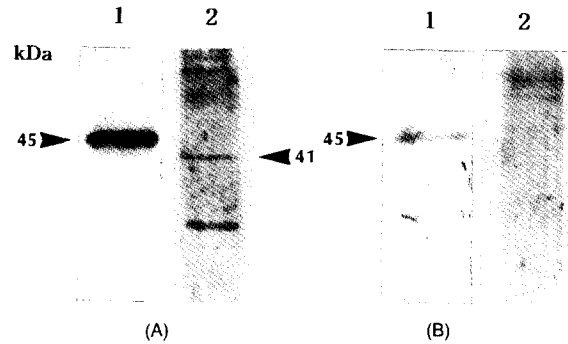


Fig. 5. Immunoblots of membrane proteins of the intermediate (A) and the large size classes (B) oocyte with antibodies raised against α subunits of G-protein. Blotting results of immunoblotting with antibodies against Gs<sub>α</sub> and Gi<sub>α</sub> are displayed in lane 1 and lane 2, respectively. The amount of protein used for gel electrophoresis was 200 μg for the intermediate and 150 μg for the large size class. Gs<sub>α</sub> of 45 kDa was detected in both size classes but Gi<sub>α</sub> of 41 kDa was detected only in the oocytes of the intermediate size class.

on the transport of <sup>125</sup>I-CP, the G-protein involved in the vitellogenin transport into the oocytes was expected to be heterotrimeric G-protein. In order to characterize G-proteins mediating vitellogenin transport, the membrane proteins of the intermediate and large oocytes were electrophoresed after treatment with β-mercaptoethanol and heat and immunoblotted with rabbit polyclonal IgG against Gs<sub>α</sub> and Gi<sub>α</sub>, which bind specifically to the C-terminal of the G subunit. A protein of 45 kDa, reacting positively to the antibody against the Gs<sub>α</sub> subunit, and three proteins of 60 kDa, 41 kDa and 29 kDa, reacting positively to the antibody against the Gi<sub>α</sub> subunit, were detected from the membrane proteins of the intermediate size class (Fig. 5). However, from the large size class only the proteins of 45 kDa and 60 kDa were detected and not the 41 kDa protein, which is assumed to be the Gi<sub>α</sub> subunit.

Discussion

The present study on the effects of AIF<sub>4</sub> and toxins on the transport of <sup>125</sup>I-CP strongly suggest that heterotrimeric G-protein is involved in the receptor-mediated endocytosis of vitellogenin specifically into the oocytes of intermediate size, in which transport is the most active compared to the small and large size classes (Lee and Kim, 1993). AIF<sub>4</sub>, a selective inhibitor of heterotrimeric G-protein (Chabre, 1990; Kahn, 1991; Helms, 1995), inhibits budding of clathrin-coated vesicles during endocytosis, in which multiple GTP-binding proteins, including heterotrimeric G-proteins, are involved (Carter et al., 1993). Endocytosis was also inhibited in the oocytes of *Xenopus* when incubated with AIF<sub>4</sub> (Mukhopadhyay et al., 1997). The inhibition by AIF<sub>4</sub> appears to be closely associated with a serious reduction in the transport of <sup>125</sup>I-CP into the oocytes of *Pseudopotamilla* by CTX in the present study. These results could be due to activation of adenylate cyclase by altering the conformation of the Gs<sub>α</sub> subunit of

heterotrimeric G-protein by treating the oocyte membrane with CTX (Janicot et al., 1991). Such interpretation is strongly supported by an observation that the transport of  $^{125}\text{I}$ -CP was greatly reduced by dbcAMP, leading to a speculation that the reduction in the transport of vitellogenin may arise as a result of increased concentration of cAMP. The CTX effect appears to be closely associated with the function of  $\text{Gs}_\alpha$ , since oocyte maturation was inhibited when the oocytes were treated with CTX or G-proteins of the oocyte membrane were bound with the antibody against  $\text{Gs}_\alpha$  subunit (Maller et al., 1979; Godeau et al., 1980; Gallo, 1995). This inhibition also resulted from the elevation of cAMP by the activation of Gs subunit with CTX (Schultz et al., 1983; Cork et al., 1990).

A significant association between the concentration of cAMP and the transport activity of vitellogenin appears in the immunoblotting studies, in which the  $\text{Gs}_\alpha$  subunit of 45 kDa was detected in intermediate and large size classes, whereas  $\text{Gi}_\alpha$  of 41 kDa was found only in the intermediate size class. Two proteins of 60 kDa and 29 kDa which reacted positively to the antibody against the  $\text{Gi}_\alpha$  subunit were disregarded, as the molecular weight of the  $\text{Gi}_\alpha$  subunit generally 38-42 kDa. In the oocytes of *Xenopus*, the  $\text{Gs}_\alpha$  and  $\text{Gi}_\alpha$  subunits were confirmed to be 43 kDa and 39 kDa, respectively (Gallo et al., 1995). Presence of the  $\text{Gi}_\alpha$  subunit of 38 kDa was also reported in the oocytes and cell-stage embryos of mouse (Jones and Schultz, 1990). In the oocytes of Chinese hamsters,  $\text{Gi}_{2,3}$  subunits of 42 kDa and 45 kDa were detected by immunoblotting tests (Xu, 1996). Detection of the  $\text{Gi}_\alpha$  subunit specifically in the oocytes of the intermediate size class is very significant in connection with the transport of vitellogenin. Activation of the  $\text{Gi}_\alpha$  subunit may result in a decrease of cAMP concentration, leading to an enhancement of vitellogenin transport, though the mechanism for this is not clear. However, a previous result that GTP enhanced the transport of vitellogenin, strongly suggested that the transport can be regulated by GTP-binding proteins other than heterotrimeric G-protein (Nam et al., 1996). This result was demonstrated in *Xenopus* oocytes, in which endocytosis is increased by injection of Rab 5 into the oocytes of a mutant with decreased GTPase activity (Mukhopadhyay et al., 1997). It may, therefore, be tentatively concluded that multiple GTP-binding proteins, including small G-proteins as well as heterotrimeric G-proteins, are all involved in the regulation of the vitellogenin transport in the oocytes of *Pseudopotamilla ocellata* through the activation of adenylyl cyclase.

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#### References

Allworth AE, Hilderbrant JD, and Ziomek CA (1990) Differential

- regulation of G-protein subunit expression in mouse oocytes, eggs and early embryos. *Dev Biol* 142: 129-137.
- Blackmore PF, Bocckino SB, Waynick LE, and Exton JH (1985) Role of a guanine nucleotide-binding regulatory protein in the hydrolysis of hepatocyte phosphatidylinositol 4,5-bisphosphate by calcium-mobilizing hormones and the control of cell calcium. Studies utilizing aluminum fluoride. *J Biol Chem* 260: 14477-14483.
- Brunskill NJ, Cockcroft N, Nahorski S, and Walls J (1996) Albumin endocytosis is regulated by heterotrimeric GTP-binding protein  $\text{Gi}_\alpha$ -3 in opossum kidney cells. *Am J Physiol* 271: F356-F364.
- Carter LL, Redelmeier TE, Woollenweber LA, and Schmid SL (1993) Multiple GTP-binding proteins participate in clathrin-coated vesicle-mediated endocytosis. *J Cell Biol* 120: 37-45.
- Chabre M (1990) Aluminum fluoride and beryll fluoride complexes: new phosphate analogs in enzymology. *Trends Biochem Sci* 15: 6-10.
- Chiba K, Kontani K, Taednuma H, Katada T, and Hoshi M (1993) Induction of starfish oocyte maturation by the  $\beta\gamma$  subunit of starfish G-protein and possible existence of the subsequent effector in cytoplasm. *Mol Biol Cell* 4: 1027-1034.
- Colombo MI, Mayorga LS, Nishimoto I, Ross EM, and Stahl PD (1994) Gs regulation of endosome fusion suggests a role for signal transduction pathway in endocytosis. *J Biol Sci* 269: 14919-14923.
- Cork RT, Tayler M, Varnold RL, Smith LD, and Robinson KR (1990) Microinjected GTP $\gamma$ S inhibits progesterone-induced maturation of *Xenopus* oocytes. *Dev Biol* 141: 447-450.
- Downs SM, Buccione R, and Eppig JJ (1992) Modulation of meiotic arrest in mouse oocytes by guanyl nucleotides and modifiers of G-proteins. *J Exp Zool* 262: 391-404.
- Gallo CJ, Hand AR, Jones TLZ, and Jaffe LA (1995) Stimulation of *Xenopus* oocytes maturation by inhibition of the G-protein  $\alpha$  subunit, a component of the plasma membrane and yolk platelet membranes. *J Cell Biol* 130: 275-284.
- Gilman AG (1987) G proteins: transducers of receptor-generated signals. *Ann Rev Biochem* 56: 615-649.
- Godeau F, Boquet P, Schorderet-Slatkine S, Schorderet M, and Baulieu E-E (1980) Studies of microbial toxins in *Xenopus laevis* oocytes. *Exp Cell Res* 129: 133-137.
- Helms JB (1995) Role of heterotrimeric GTP binding proteins in vesicular protein transport: indications for both classical and alternative G-protein cycles. *FEBS Lett* 369: 84-88.
- Jaffe LA, Gallo CJ, Lee RH, Ho YK, and Jones TLZ (1993) Oocyte maturation in starfish is mediated by the  $\beta\gamma$  subunit complex of a G-protein. *J Cell Biol* 121: 775-783.
- Janicot M, Fougue F, and Desbugois B (1991) Action of rat liver adenylyl cyclase by cholera toxin requires toxin internalization and processing in endosomes. *J Biol Chem* 266: 12858-12865.
- Jones J and Schultz RM (1990) Pertussis toxin catalyzed ADP-ribosylation of a G-protein in mouse oocytes, egg and preimplantation embryos: Developmental changes and possible functional roles. *Dev Biol* 139: 250-262.
- Kahn RA (1991) Fluoride is not an activator of the smaller (20-25 kDa) GTP-binding proteins. *J Biol Chem* 266: 15595-15597.
- Kanaho Y, Moss J, and Vaughan M (1985) Mechanism of inhibition of transducin GTPase activity by fluoride and aluminum. *J Biol Chem* 260: 11493-11497.
- Katada T, Northup JK, Bokoch GM, Ui M, and Gilman AG (1984) The inhibitory guanine nucleotide-binding regulatory component of adenylyl cyclase. Subunit dissociation and guanine nucleotide-dependent hormonal inhibition. *J Biol Chem* 259: 3578-3585.
- Lee YR (1988) Changes in the protein components of vitelline envelope during oogenesis of a tubicolous polychaete, *Schizobranhia insignis*. *Dev Cell Different* 25: 23-26.
- Lee YR and Kim YN (1993) Selective transport of coelomic fluid protein into oocytes of a tubicolous polychaete, *Pseudopotamilla ocellata*. *Invert Reprod Dev* 24: 119-126.

- Lee BK, Nam HJ, and Lee YR (1997) Receptor-mediated transport of vitellogenin during oogenesis of a polychaete, *Pseudopotamilla ocellata*. *Korean J Biol Sci* 1: 341-344.
- Maller JL, Butcher FR, and Krebs EG (1979) Early effect of progesterone on levels of cyclic adenosine 3',5'-monophosphate in *Xenopus* oocytes. *J Biol Chem* 254: 579-582.
- Mukhopadhyay A, Barbieri AM, Funato K, Roberts R, and Stahl PD (1997) Sequential actions of Rab5 and Rab7 regulate endocytosis in the *Xenopus* oocyte. *J Cell Biol* 136: 1227-1237.
- Nam HJ, Kang WS, and Lee YR (1996) Involvement of GTP-binding proteins in stage-specific receptor-mediated endocytosis of coelomic fluid proteins into oocytes of *Pseudopotamilla ocellata*. *Korean J Zool* 39: 292-298.
- Schmalzing G, Richter HD, Hansen A, Schwarz W, Just I, and Aktories K (1994) Involvement of the GTP-binding protein Rho in constitutive endocytosis in *Xenopus laevis* oocytes. *J Cell Biol* 130: 1319-1332.
- Schultz RM, Montgomery RR, Ward-Bailey PF, and Eppig JJ (1983) Regulation of oocyte maturation in the mouse: possible roles of intercellular communication, cAMP, and testosterone. *Dev Biol* 95: 294-304.
- Serafini T, Orci L, Amherdt M, Brunner M, Kahn RA, and Rothman JE (1991) ADP-ribosylation factor is a subunit of the coat of Golgi-derived cop-coated vesicle: a novel role for a GTP-binding protein. *Cell* 67: 239-253.
- Shilling F, Chiba KC, Hoshi M, Kishimoto T, and Jaffe LA (1989) Pertussis toxin inhibits 1-methyladenine-induced maturation in starfish oocytes. *Dev Biol* 133: 605-608.
- Strand CF and Wong K (1985) Calcium mobilization in fluoride activated human neutrophils. *Biochem Biophys Res Commun* 133: 161-167.
- Tang WJ and Gilman AG (1992) Adenylyl Cyclases. *Cell* 70: 869-872.
- Turner PR, Jaffe LA and Primakoff P (1987) A cholera toxin sensitive G-protein stimulates exocytosis in sea urchin eggs. *Dev Biol* 120: 577-583.
- Xu Y and Barbieri JT (1996) Pertussis toxin-catalyzed ADP-ribosylation of G<sub>i2</sub> and G<sub>i3</sub> in CHO cells is modulated by inhibitors of intracellular trafficking. *Infect Immun* 64: 593-599.

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