Oleanane Triterpenoids from Gordonia ceylanica

H. M. T. B. Herath* and P. S. Athukoralage

Institute of Fundamental Studies, Hantana Road, Kandy, Sri Lanka

Abstract – Chemical investigation of hot hexane extract of the stem bark of *Gordonia ceylanica* afforded a new triterpenoid, 3β -acetoxy-28-hydroxyolean-12-ene (1) and three other oleanane triterpenoids, 3β -hydroxyolean-12-ene (2), 3β -acetoxyolean-12-ene (3), and 3β -acetoxyolean-12-en-11-one (4) which are new to the species. Structure of compound 1 was suggested by ¹H NMR, ¹³C NMR and MS spectral data and confirmed by converting to previously reported compound, erythrodiol diacetate (5). Structures of 2, 3 and 4 were established by comparison of the spectral data with the previously reported compounds. Further the acid hydrolysate of 4 was identical with 3β -hydroxyolean-12-en-11-one (6).

Key words – Gordonia ceylanica, Theaceae, triterpenoids, oleanans.

Gordonia ceylanica belongs to the family Theaceae which consist approximately 520 species under 25 genera in the tropical and warm temperate areas of the world. The twelve Theaceae species distributed in Sri Lanka are categorize under five different genera. Among them all species belong to genus Gordonia, G. ceylanica, G. dassanayakei, G. speciosa, and G. elliptica are endemic to the country (Dassanayake, 1996). "Mihiriya" is the local name for all Gordonia species and the color of the flower is used as adjective to differentiate the certain species. However though all the Gordonia species are being used in traditional medicine in Sri Lanka, no previous chemical investigation is reported on any of these endemic species. Therefore the chemical and biological investigation of Sri Lankan Gordonia species are of our great interest.

Experimental

Plant material

The stem bark of *G. ceylanica* was collected from Nuwara Eliya district in the Central Province of Sri Lanka and the voucher specimen was authenticated by comparison with the herbarium specimen, 8291, *Jayasuriya* (17.10.92) at the National Herbarium, Royal Botanic Gardens, Peradeniya Sri Lanka.

Extraction

Air dried and ground stem bark of *G. ceylanica* (1.0 kg) was successively extracted with hot hexane, dichloromethane and methanol. The concentrated hot hexane extract (4.82 g) was fractionated by medium pressure column packed with silica gel using gradient elution with hexane, dichloromethane and methanol. These fractions were further purified by small scale medium pressure, flash and thin layer chromatography to give following compounds.

3β-Acetoxy-28-hydroxyolean-12-ene (1) – Colorless needles, mp 144-146°C; ¹H NMR (300 MHz, CDCl₃) δ: 5.19 (1H, dd, J=3.4 & 3.2 Hz, H-12), 4.50 (1H, dd, J=6.0 & 8.0 Hz, H-3), 3.55 (1H, d, J=10.8 Hz, H-28), 3.21 (1H, d, J=10.8 Hz, H-28) 2.05 (3H, s, OCOCH₃),

^{*}Author for correspondence.

Natural Product Sciences

1.16, 0.95, 0.94 (each 3H, s, 3xCH₃), 0.92-0.82 (12H, br.s, 4xCH₃); ¹³C NMR (75 MHz, CDCl₃) table-1; MS (70 eV), m/z (rel. int.%): 484 (M⁺, 0.5), 234 (8), 203 (100), 189 (9), 105 (15), 81 (8).

3β-Hydroxyolean-12-ene (2) – Colorless needles, mp 194-195°C {(Lit. Faujan et al., 1995) mp 195-196°C}; ¹H NMR (300 MHz, CDCl₃) δ: 5.18 (1H, dd, *J*=3.6 & 3.6 Hz, H-12), 3.21 (1H, dd, *J*=10.2 & 5.7 Hz, H-3), 1.13 (3H, s, H-27), 0.99, 0.96, 0.93 (each 3H, s, 3xCH₃), 0.87 (6H, br.s., 2xCH₃), 0.83, 0.79 (each 3H, s, 2xCH₃); ¹³C NMR (75 MHz, CDCl₃) table-1; MS (70 eV), m/z (rel. int.%): 426 (M⁺, 10), 218 (100), 203 (35), 135 (25), 95 (45).

3β-Acetoxyolean-12-ene (3) – Colorless needles, mp 236-237°C {Lit. Shiojima et al., 1996 (mp 238-240°)}; 1 H NMR (300 MHz, CDCl₃) δ: 5.18 (1H, dd, J=3.2 & 3.6 Hz, H-12), 4.50 (1H, dd, J=7.2 & 8.4 Hz, H-3), 2.05 (3H, s, OCOCH₃), 1.13 (3H, s, H-27), 0.96 (6H, s, H-25, H-26), 0.87 (12H, br.s., H-23, H-24, H-29 & H-30), 0.83 (3H, s, H-28); 13 C NMR (75 MHz, CDCl₃) table-1; MS (70 eV), m/z (rel. int.%): 468 (M⁺, 3), 218 (100), 203 (42), 189 (17), 135 (19), 95 (45), 81 (33), 69 (52), 55 (30).

3β-Acetoxyolean-12-en-11-one (4) – Colorless needles, mp 272-273°C {Lit. Bandaranayake, 1980 (mp 269-272°)}; ¹H NMR (300 MHz, CDCl₃) & 5.58 (1H, s, H-12), 4.51 (1H, dd, *J*=11.5 & 5.1 Hz, H-3), 2.79 (1H, ddd, *J*=13.5, 3.6 & 3.6 Hz, H-18), 2.35 (1H, s, H-9), 2.04 (3H, s, OCOCH₃), 1.56, 1.35, 1.15, 1.13, 0.91, 0.88, 0.87, 0.85 (each 3H, s, 8xCH₃); ¹³C NMR (75 MHz, CDCl₃) table-1; MS (70 eV) m/z (rel. int%): 482 (M⁺, 5), 273 (38), 232 (59), 217 (12), 175 (33), 135 (100), 121 (18), 95 (30), 81 (20), 55 (25), 43 (55).

Erythrodiol diacetate (5) – Colorless needles, mp 186-187°C {Lit. Shiojima et al., 1995 (mp 189-190°)}; 1 H NMR (200 MHz, CDCl₃) & 5.20 (1H, dd, J=3.5 & 3.5 Hz, H-12), 4.50 (1H, dd, J=7.0 & 9.5 Hz, H-3), 4.06 (1H, d, J=11.2 Hz, H-28), 3.70 (1H, d, J=11.4 Hz, H-28), 2.04 (6H, s, 2x OCOCH₃), 1.16 (3H, s,

Table 1. 13C NMR spectral data of compounds 1-6

Carbon No.	1	2	3	4	5	6
1	38.3	38.5	38.3	38.8	38.3	39.1
2	23.6	27.2	23.6	23.4	23.5	27.3
3	80.8	79.0	80.9	80.6	80.9	78.7
4	37.7	38.7	37.8	38.1	37.8	38.5
5	55.2	55.1	55.3	55.1	55.2	54.9
6	18.3	18.3	18.3	18.7	18.2	18.7
7	32.5	32.6	32.6	32.7	32.5	32.7
8	39.8	39.7	39.8	43.4	39.8	43.4
9	47.6	47.6	47.6	61.5	47.5	61.7
10	36.9	36.9	36.9	37.0	36.8	37.0
11	23.6	23.5	23.7	200.0	23.5	200.3
12	122.3	121.7	121.7	128.1	122.8	128.1
13	144.5	145.2	145.2	170.5	143.6	170.6
14	41.7	41.7	41.7	45.4	41.6	45.4
15	25.6	26.1	28.4	26.4	25.5	26.4
16	22.0	26.9	26.2	26.5	22.2	26.4
17	36.9	32.5	32.5	32.4	35.7	32.3
18	42.3	47.2	47.3	47.6	42.5	47.6
19	46.8	46.8	46.8	45.2	46.2	45.1
20	31.0	31.1	31.0	31.0	30.9	31.0
21	34.1	34.7	34.8	34.4	34.0	34.4
22	31.0	37.1	37.2	36.5	31.4	36.5
23	29.7	28.1	28.1	28.0	29.7	28.1
24	15.5	15.5	16.7	16.7	15.5	16.3
25	15.5	15.6	15.6	16.4	15.5	15.5
26	16.7	16.8	16.8	17.4	16.7	17.5
27	25.9	26.0	26.0	23.5	25.9	23.4
28	69.7	28.4	27.0	28.7	70.7	28.7
29	33.3	33.3	33.3	33.0	33.1	33.0
30	23.6	23.6	23.6	23.6	23.5	23.9
$o\underline{c}o$	170.9	-	170.9	170.9	171.3	-
					171.0	
$COCH_3$	21.3	-	21.3	21.3	21.3	-
					20.9	

CH₃), 0.96 (6H, br.s., 2xCH₃), 0.90-0.83 (12H, br.s., 4xCH₃), ¹³C NMR (50 MHz, CDCl₃) table-1; m/z (rel. int.%): 526 (M⁺, 6), 276 (11), 203 (100), 189 (14), 105 (21), 81 (7).

3β-hydroxyolean-12-en-11-one (6) – Colorless needles, mp 227-229°C {Lit. Bandaranayake, 1980 (mp 229-232°)}; ¹H NMR (200 MHz, CDCl₃) δ: 5.58 (1H, s, H-12), 3.28 (1H, m, H-3), 2.79 (1H, ddd, *J*=13.4, 3.4 & 3.4 Hz, H-18), 2.35 (1H, s, H-9), 1.14 (6H, s 2xCH₃), 1.36, 0.99, 0.91, 0.88, 0.85 and 0.79 (each 3H, s, 6xCH₃); ¹³C NMR (50 MHz, CDCl₃) table-1; MS (70 eV) m/z (rel. int.%): 440 (M⁺, 4), 273 (52), 232 (76), 175 (23), 135 (100), 121 (14).

Vol. 4, No. 4, 1998

Results and Discussion

1b, m/z = 203

The molecular formula C₃₂H₅₂O₃, suggested by the molecular peak at m/z=484 in the mass spectrum of compound 1 was in concurrence with the thirty two corresponding signals appeared in the ¹³C NMR spectrum. The presence of single olefinic proton (H-12) and the proton under the vicinity of the acetate group at C-3 were indicated by double doublets at δ 5.19 and 4.50 respectively while two protons attached to C-28 were represented by the two doublets at δ 3.55 and 3.21 in the ¹H NMR spectrum of 1. Methyl protons of the acetate group of the molecule were appeared as 3H singlet at δ 2.05 while the other seven methyl groups were appeared in the region of δ 1.16 to 0.86 in the ¹H NMR spectrum of compound 1. ¹³C NMR spectrum of 1 also showed the presence of two olefinic carbons, C-12 and C-13 at δ 122.3 and 144.5 respectively. Further the C-3 carbon flanking the acetate group and C-28 containing hydroxyl group were appeared at δ 80.8 and 69.7 respectively in the ¹³C NMR spectrum of 1. The peak at m/z 234 in the MS spectrum was assigned for the fragment ion 1a which gives as a result of retro Diels-Alder

fragmentation of compound 1. The base peak at m/z 203 in the mass spectrum was analogous to the resulted ion 1b by cleavage of CH₂OH moiety from 1a. These mass spectroscopic evidence are also in consistent with other spectroscopic evidence suggested that the CH2OH group in the molecule is flanking to the C-17 position. Further the acetylation of 1 to give erythrodiol diacetate (5) (Xue at al., 1988 & Shiojima et al., 1995) were strongly supported to confirm the structure proposed for compound 1. The structures assigned for compound 2 and 3 were identical with the previously reported compounds, β-amyrin (Faujan et al., 1995) and β-amyrin acetate (Shiojima et al., 1996 & Agrawal et al., 1992) respectively and confirmed by co-TLC with authentic samples. The presence of the carbonyl group at C-11 in the proposed structure for compound 4 was indicated by the signal at δ 200.0 in the ¹³C NMR and further supported by the two sharp singlets at δ 2.35 and 5.58 in the ¹H NMR which appeared due to H-9 and H-12 respectively. Further the spectral data of compound 4 were identical with the reported data of the previously isolated compound 3-βacetoxyolean-12-en-11-one (Bandaranayake, 1980) and also the spectral data of the acid hydrolysed product of compound 4 were identical with the previously reported values of 3β-hydroxyolean-12-en-11-one (6) (Bandaranayake, 1980).

Acknowledgements

Authors wish to thank Mr. Aruna Weerasooriya of the Department of Botany, University of Peradeniya, Sri Lanka for collection and the identification of the plant materials, Mr. Wimal Padmasiri, Department of Chemistry, University of Colombo, Sri Lanka and Dr. Joanne Jamie at the Department of Chemistry, University of Wollongong, NSW, Australia for the NMR

256 Natural Product Sciences

and MS spectral data and Mr R.G.U. Jayalal, Institute of Fundamental Studies, Kandy, Sri Lanka for the technical assistance.

References

- Dassanayake, M. D. and Clayton, W. D., A revised Handbook to the Flora of Ceylon Vol. 10, page 392, Oxford 1BH Publishing Co. Ltd., New Delhi, 1996.
- Faujan, B. H. A., Ahmad, S. S., Zuridah, C. A. and Sahri, M. H., β-Amyrin from the bark of Malaysian merbau Intsia Palembanica. ACGC Chemical Research Communications. 3, 35-37 (1995).
- Shiojima, K., Suzuki, H., Kodera, N., Ageta, H., Chang, H. C. and Chen, Y. P., Composite Constituents: Thirty nine triterpenoids including two novel compounds from *Ixeris chinensis*. Chem. Pharm. Bull. 44, 509-514 (1996).

- Bandaranayake, W. M., Terpenoids of Canarium zeylanicum. Phytochemistry 19, 255-257 (1980).
- Shiojima, K., Suzuki, H. and Ageta, H., Composite Constituents: Three New Triterpene Triols Isolated from Fresh Roots of Picris hieracioides subsp. Japonica. Chem. Pharm. Bull., 43, 1640-1642 (1995).
- Xue, H. Z., Lu, Z. Z., Konno, C., Soejarto, D. D., Cordell, G. A., Fong, H. H. S. and Hodgson, W., 3β-(3,4-Dihydroxycinnamoyl)-erithodiol and 3β-(4hydroxycinnamoyl)-erithodiol from Larrea tridentata. Phytochemistry 27, 233-235 (1988).
- Agrawal, P. K. and Jain, D. C., ¹⁸C NMR Spectroscopy of Oleanane Triterpenoids. *Progress in Nuclear Magnetic Resonance Spectroscopy* **24**, 1-90 (1992).

(Accepted September 17, 1998)