Grayanane Diterpenoids from Pieris formosa¹

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Abstract – Three grayanane diterpenoids (1-3) were isolated from *Pieris formosa*. 1 was identified as a new natural product and 2 and 3 as known grayanoside C and grayanotoxin XVIII on the basis of spectral analysis.

Key words - Pieris formosa; Ericaceae; diterpenoids; pierisformosin A.

Introduction

The grayanoids are a relatively unknown class of diterpenoids, which possess a 5/7/6/5 (trans or cis/cis/cis) ring system, formed pro- bably by rearrangement of kaurane skeleton in biogenetic pathway. Its existence was found especially in the genera Pieris, Rhododendron, Lyonia and Leucothoe of Erica- ceae family. Up to now more than 50 grayan- oids have been reported in literatures, am- ong which the majority exhibited remark- able toxicities, and significant antifeedant and insecticidal activities (Wang et al., 1997). In order to search the natural products which have potential to be developed as insecticide, pesticide and herbicide, we have initiated the chemical studies on grayanane diterpenoids from Chinese Ericaceae plants.

Pieris formosa (Wall) D. Don (Ericaceae) is an evergreen shrub or tree, growing mainly in hilly and valley regions of south and southwest China. It is a well known poisonous plant, which Chinese name "Mei-Li-Ma-Zui-Mu" means the nice wood that horse would get drunk after eating them. The monograph

concerned describes that the poultry would fall into coma after taking its leaves or stems accidentally. The symptoms including dyspnea, motion imbalance, spreading the four limbs would appear if mice were administrated with the chloroform extracts. As folk practice, the juice of the fresh leaves can be used as insecticide and as lotion for treatment of tinea and scabies in clinic (Chen et al., 1987). Previously some steroids have been isolated from this plant (Puri et al., 1982), but no any gravanoids were reported. In the course of searching bioactive natural products, we found that ethyl acetate and n-BuOH fractions of the plant extracts were effective in brine shrimp tests, which encouraged us to do its chemical investigation. This paper describes the isolation and structural elucidation of three grayanane diterpenoids (1-3) from the ethyl acetate fraction of the plant. Compound 1 was identified as a new natural product, named pierisformosin A and 2 and 3 as known grayanoside C and grayanotoxin XVIII (Sakakibara et al., 1979, 1980), respectively, which were first discovered in the plant.

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¹Part 1 in the series: "Chemical Stadies on Ericaceae Plants".

Experimental

General – $[\alpha]_D$: JASCO, DIP-181, polarimeter. IR: Perkin-Elmer 599B spectrometer. 1H and ^{13}C NMR spectra: Bruker AM-400. Chemical shifts are reported in ppm with solvent signal as int. standards. MS: MAT-95.

Plant material – The leaves of the plant were collected from Kaihua county of Zhejiang Province in November, 1996 and identified by Prof. Bing-Yang Ding of Hangzhou University. A voucher specimen was deposited in the Herbarium of Shanghai Institute of Materia Medica.

Extraction and isolation - The leaves of P. formosa (20 kg) were airdried, ground and extracted with 95% ethanol under reflux. After removal of the solvent by evaporation, the residue was adjusted to about 15% ethanol solution and stored in refrigerator overnight to precipitate chlorophyll. The supernatant was extracted with CHCl₃, EtOAc and n-BuOH, respectively. The EtOAc extract was evaporated to give a red mass (200 g), which was applied to a silica gel column, eluting with EtOAc containing increasing amounts of MeOH. Repeated column chromatography of the fraction, eluting with CHCl₃: MeOH (15:1) led to yield 1 (12 mg) and 2 (10 mg). And 3 (19 mg) was obtained from a fraction of the column chromatography and purified by RP-8 column chromatography, eluting with MeOH:H₂O (6:4).

1, pierisformosin A: amorphous powder. $[\alpha]_{0}^{15}$ 18.33° (MeOH, c 0.36); IR: ν_{max}^{KBr} 3419, 1637, 1456, 1036 cm⁻¹; ¹H NMR (C₅D₅N), see Table 2. ¹³C NMR (C₅D₅N), see Table 1. EIMS m/z: 318(75, [M-H₂O]+), 282(32), 271(35), 257(38), 239(28), 229(22), 211(25), 159(31), 147(32), 145(40), 135(45), 119(43), 109(100), 91(62), 69(56), 55(54).

2, grayanoside C: viscous syrup, $[\alpha]_{\rm D}^{15}$ 0.66° (MeOH, c 0.35); IR: $v_{\rm max}^{\rm KBr}$ 3100-3500, 1635, 1448, 1080, 916, 883 cm⁻¹; FABMS m/z: 522 [M+Na+H]⁺, 537 [M+K]⁺; ¹H NMR (C_5D_5N): δ 1.31, 1.48, 1.91 (each 3H, s), δ 3.17 (1H, t, J=9.0 Hz), δ 3.28 (1H, dd, J=13.8, 10.3), δ 5.14 (1H, d, J=7.8 Hz), δ 5.26, 5.28 (2H, each s). ¹³C NMR (C_5D_5N), see Table 1.

3, grayanotoxin XVIII: oil. $[\alpha]_{15}^{15}$ -4.23° (MeOH, c 1.50); ¹H NMR (C₅D₅N), see Table 2. ¹³C NMR (C₅D₅N), see Table 1. EIMS m/z: 318 (28, [M-H₂O]⁺, 300(41), 282(31), 267(15), 275(18), 257(35), 239(41), 229(21), 211(30), 185(32), 159(37), 135(53), 119(86), 109(86), 93(73), 69(100), 55(80).

Results and Discussion

Compound 1, $[\alpha]_D^{15}$ 18.33° (MeOH, c 0.36), had a molecular formula $C_{20}H_{32}O_4$ from its EIMS (m/z 318, [M-H₂O]⁺) and the ¹H and ¹³C NMR spectra. The IR spectrum showed

Table 1. 13C NMR data of 1-3

C	1	2	3	С	1	2	3
1	58.7d	59.3d	44.0d	14	35.5t	35.8t	36.1t
2	36.7t	35.4t	39.0t	15	57.2t	57.5t	62.1t
3	83.5d	92.2d	80.9d	16	78.5s	78.7s	79.2s
4	50.7s	51.7s	50.2s	17	24.7q	24.9q	25.0q
5	86.7s	86.3s	83.2s	18	22.7q	23.5q	23.9q
6	71.0d	71.4d	70.4d	19	20.9q	21.1q	18.9q
7	48.7t	49.0t	46.3t	20	110.5t	110.4t	112.0t
8	45.6s	45.9s	44.3s	glu-1'		105.2d	
9	55.6d	55.5d	52.1d	2'		75.9d	
10	153.9s	154.2s	152.6s	3'		78.8d	
11	26.5t	26.8t	23.7t	4'	71.9d		
12	26.7t	26.9t	25.5t	5'		78.8d	
13	49.5d	49.8d	47.5d	6'		63.0t	

Table 2.	. 'H NMR	data of	1 and	l 3
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Н	1	3	Н	1	3
1	3.01 dd(9.8, 3.4)	3.15t(9.4)	12	1.38 m	1.57-1.80 m
2	1.99 m	2.64 m	13	2.02 m	2.27 m
	2.19 m	2.75 m	14	1.83 d(11.0)	1.85 d(11.1)
3	3.85 d(4.0)	3.96 dd(6,6, 2.5)	Į	2.09 dd(11.0, 4.0)	2.37 dd(11.1, 4.5)
6	3.80 dd(10.0, 2.0)	4.20 dd(9.4, 2.3)	15	1.63 d(15.0) 1.77 d(15.0)	2.07 d(14.2), 2.17 d(14.2)
7	1.67 m	1.92 dd(14.0, 2.0)	17	1.3 ls	1.55 s
	2.99 m	2.57 dd(14.0, 9.4)	18	1.0 ls	1.11 s
9	2.01 m	2.82 m	19	1.51 s	$1.57 \mathrm{\ s}$
11	1.43 m, 1.55 m	1.57-1.80 m	20	5.04 s, 5.09 s	5.17 s, 5.19 s

absorption band of hydroxyl group (3419 cm⁻¹) and double bond (1637 cm⁻¹). The ¹H and ¹³C NMR spectra (Table 1 and 2) indicated the presence of three singlet methyls $(\delta_{\rm H} \ 1.01, \ 1.31, \ 1.51; \ \delta_{\rm C} \ 24.7, \ 22.7, \ 20.9), \ {\rm two}$ oxygenated methines ($\delta_{\rm H}$ 3.84, 3.80; $\delta_{\rm c}$ 83.5, 78.5), two oxygenated quaternary carbons $(\delta_{\rm C} 86.7, 78.5)$ and one terminal double bond $(\delta_{\rm H} 5.04, 5.09; \delta_{\rm C} 153.9, 110.5)$. The ¹H-¹H COSY revealed the existence of the following fragments: H_aC=C-CH-CH₂-CH(OH)-, H_bC=C-CH-CH₂-CH₂-CH- and -CH(OH)-CH₂-, each of which was connected to quaternary carbon atoms at one or both ends. The all above data confirmed to structural requirement of grayanane diterpenoids. Further investigation exhibited that the ¹³C NMR data of 1 were in good agreement with that of the aglycone of grayanoside C (2), obtained by its enzymatic hydrolysis (Sakakibara et al., 1980). Considering other spectral evidences. 1 was determined as the aglycone of 2. Due to complicated stereochemistry, the structure of 2 and its conformation was finally determined by X-ray crystallography of its derivative (Sakaki- bara et al., 1980). However, after our literature investigation, 1 was determined as a new natural product and named as pierisformosin A.

In our NMR study, three methyls and four quaternary carbons were differentiated by NOESY and HMBC experiments. The NOE's between one methyl (δ 1.01) and both H-3 and H-6 were observed, which indicated the signal at δ 1.01 should be assigned as C_{18} methyl. The signal at δ 1.51 was assigned as C_{19} methyl from correlation with the signal at δ 1.01 in NOESY spectrum. The HMBC spectrum revealed cross-peaks between the signals at δ_c 50.7/H-18, 19; δ_c 86.7/H-1, 2, 7, 18, 19; δ 45.6/H-6, 7, 9, 11 and δ 78.5/H-14, 17, which indicated C₄, C₅, C₈ and C₁₆ respectively. From above analyses together with ¹H-¹H COSY results, ¹H and ¹³C NMR data of 1 were assigned unambiguously as shown in Table 1 and 2.

Comparing general *trans*-fused ring A and B, 1 and 2 were found to be a rare exception among the grayanoids, possessing the *cis*-conjunctive ring A/B system with 1β-H. It was confirmed by the fact that H-1 and H-9 showed correlation in the NOESY spectrum. The conformation of 1 was elucidated as shown in Fig. 1 by previous study on X-ray crystallography of its derivative (Sakakibara,

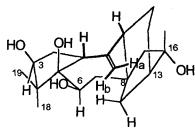


Fig. 1. The conformation of 1.

Vol. 4, No. 2, 1998

et al., 1980). According to this conformation, it could be rationalized that H-20a/H-1 and H-20b/H-9 appeared allylic coupling combined with W-type long-range coupling in ¹H
¹H COSY due to taking the same plane. It also could be explained that only C₁₈ methyl had correlation with both H-3 and H-6 in NOESY spectrum.

The second compound isolated from the plant had a molecular formula $C_{26}H_{42}O_9$ by its FABMS and ¹H and ¹³C NMR data. It showed positive reaction for Molisch test. On acidic hydrolysis, it afforded glucose as sugar constituent, detected by co-TLC. Further spectral study revealed that all ¹H and ¹³C NMR data of this compound were identical with that of grayanoside C. Thus it was determined as known grayanoside C, **2**.

Compound 3 had the molecular formula $C_{20}H_{32}O_4$ by EIMS and NMR spectra. The ¹H and ¹³C NMR spectra of 3 were very similar with that of 1, except little differences around C-1. The further investigation suggested that 3 should be grayanotoxin XVIII, the C_1 epimer of 1 with α -orientation of H-1, which was isolated from *Leucothoe grayana* (Sakakibara *et al.*, 1979). The ¹H and ¹³C

NMR data of **3** could be assigned by ¹H-¹H COSY and comparison with **1** and **2**. The compounds **2** and **3** were known grayanane diterpenoids, but first isolated from *Pieris formosa*. The biological tests of compounds **1**-**3** are in progress.

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(Accepted March 9, 1998)