

[報 文]

Effects of Hydroxybrazilin on Glutathione Depletion Induced by BrCCl₃ and Menadione in Cultured Rat Hepatocytes

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ABSTRACT

In this study we investigated the effect of hydroxybrazilin on glutathione depletion induced by BrCCl₃ and menadione in cultured hepatocytes to understand the cellular mechanisms of hepatoprotective effect of hydroxybrazilin. Hydroxybrazilin alone had no effect on total glutathione level and the ratio of reduced glutathione/total glutathione (GSH/(GSSG+GSH)). BrCCl₃ dramatically decreased total glutathione level and hydroxybrazilin significantly prevented glutathione depletion by BrCCl₃. The ratio of GSH/(GSSG+GSH) was also decreased by BrCCl₃ and recovered by hydroxybrazilin treatment. Menadione decreased total glutathione level and the ratio of GSH/(GSSG+GSH) but hydroxybrazilin showed no significant effects on menadione-induced glutathione depletion. These data suggest that hydroxybrazilin might prevent the hepatotoxicity induced by chemical-derived radicals but not the toxicity linked with oxidative stress.

Key words : Hydroxybrazilin, Bromotrichloromethane, Menadione, Cultured hepatocyte, glutathione

INTRODUCTION

Glutathione is the most important and widely distributed nonprotein thiol in living systems, which plays a major role in many redox and detoxification reactions in the liver¹⁾; reduced form of glutathione (GSH) traps the electrophilic metabolites of some toxic chemicals and prevents their binding to hepatic proteins and enzymes^{2,4)} and the conjugation

of the chemical with glutathione usually results in the formation of a nontoxic, water-soluble metabolites that is easily excreted. Nearly all of the glutathione is present as reduced form (GSH) and the amount of oxidized form of glutathione disulfide (GSSG), is very much lower. Redox status of these thiols is maintained by intracellular glutathione reductase and NADPH.⁵⁾

In recent years, the redox transition of glu-

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tathione has been reported to affect or modulate a variety of cellular process, including enzyme activity⁶⁾ and the transport of small solutes and ions,⁷⁾ especially calcium.⁸⁾ And disturbances in intracellular glutathione homeostasis induced by hepatotoxins were associated with changes in cell surface structure and preceded cell death.⁹⁾ It has also been reported that lowering of the intracellular glutathione level by the addition of diethyl maleate promoted the ADP · Fe³⁺-induced cellular lipid peroxidation (malondialdehyde production), and the formation of conjugated dienes.¹⁰⁾

According to our previous studies,¹¹⁻¹³⁾ some γ -pyranoidal natural dyes such as brazilin and hydroxybrazilin have hepatoprotective effects. They lowered sALT and sAST levels in the CCl₄-treated mice and reduced lipidperoxidation in ethanol- and CCl₄-treated rats. In the present study, we studied the effects of hydroxybrazilin on the BrCCl₃- and menadione-induced GSH depletion in cultured rat hepatocytes to investigate the cellular hepatoprotective mechanisms of by hydroxybrazilin.

MATERIALS AND METHODS

1. Materials

SD male rats (200~250g) were obtained from Laboratory Animal Center, Seoul National University, housed 2~4 per cage, and allowed foods and water *ad libitum*. BrCCl₃ was obtained from Wako Chemical Co., Japan. Hydroxybrazilin was obtained from Merck Co., Germany. Other reagents were obtained from Sigma Chemical Co., USA.

2. Monolayer culture of hepatocytes

Hepatocytes were isolated from SD male rats by a collagenase perfusion technique as described by Berry and Friend.¹⁴⁾ The isolated hepatocytes were inoculated onto collagen coated dishes in CO₂ incubator and then, mo-

no-layer was formed within 4 hours. This monolayer hepatocytes were cultured for 24 hours in William's E medium supplemented with 5% FBS.

3. Determination of intracellular glutathione

The medium was replaced with serum free William's E medium containing hydroxybrazilin (1~100 μ M). BrCCl₃ (0.2 μ M/ml) or menadione (100 μ M) was added directly to the culture medium and incubated for 30 or 60 minutes, respectively. At the end of incubation, the cells were washed with ice cold phosphate buffered saline. The washed hepatocytes were collected with 0.5% picric acid (1 ml) into microcentrifuge tube. After centrifugation, the supernatant was withdrawn for the determination of total glutathione and oxidized form of glutathione. To determine the oxidized glutathione, reduced glutathione was derivatized with 2-vinylpyridine. The amount of glutathione was determined by DTNB (5, 5'-dithiobis-2-nitrobenzoic acid)-GSSG (oxidized glutathione) reductase recycling method.¹⁵⁾ Reduced form of glutathione (GSH) was calculated from the difference between the total and the oxidized glutathione.

All statistical comparisons were performed using a Student t-test. Differences were considered statistically significant at $p < 0.05$.

RESULTS AND DISCUSSION

Hydroxybrazilin, a γ -pyranoidal natural dye, is known to have various biological activities including hepatoprotective effects. In order to clarify cellular hepatoprotective mechanisms of hydroxybrazilin, we introduced the monolayer cultured hepatocyte system, and investigated the effects on the glutathione depletion induced by hepatotoxins such as BrCCl₃ and menadione. For the application of hydroxybrazilin to in vitro hepatocyte system, the

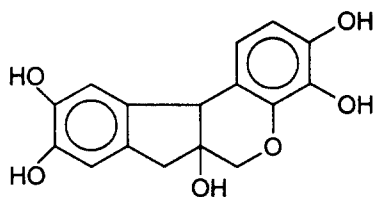


Fig. 1. Structure of hydroxybrazilin.

Table 1. Effects of hydroxybrazilin on glutathione level and GSH/(GSH+GSSG) ratio in normal hepatocytes.

Groups	GSH+GSSG (nmol glutathione / 3×10^6 cells)	GSH	GSH/(GSH+GSSG) (% value)
Normal	101 \pm 5	89 \pm 5	88.1%
10 ⁻⁶ M Hrx	107 \pm 3	94 \pm 1	87.9%
10 ⁻⁴ M Hrx	105 \pm 4	92 \pm 4	87.6%

Hrx : hydroxybrazilin

Values are the mean \pm SE of three separate experiments

suitable concentration of hydroxybrazilin which does not affect the cellular glutathione level should be found. As shown in table 1, 100 μ M and 10 μ M of hydroxybrazilin had no effect on total glutathione level and GSH/(GSH+GSSG) ratio. hydroxybrazilin was not cytotoxic to monolayer cultured hepatocytes at these or lower concentrations.

Carbontetrachloride (CCl₄) has been widely used for the investigation of chemical-induced hepatotoxic mechanism. But CCl₄ was not evaluated as a suitable hepatotoxin for the in vitro study, especially in cultured rat hepatocytes, in which the level of cytochrome P-450 is markedly lowerd. In the present study, BrCCl₃ was used as a hepatotoxin instead of CCl₄ because BrCCl₃ metabolized to \cdot CCl₃ radical more easily than CCl₄.⁽¹⁶⁾ Trichloromethane radical (\cdot CCl₃) attacks membrane lipids, cellular proteins and DNA. Subsequently it causes lipid peroxidation and cellular damage leading to cell death. BrCCl₃ treatment on the monolayer cultured hepatocytes showed depletion of total glutathione level (Table 2).

Table 2. Effects of hydroxybrazilin and vitamine E on glutathione level and GSH/(GSH+GSSG) ratio in BrCCl₃-treated hepatocytes.

Groups	GSH+GSSG (nmol glutathione / 3×10^6 cells)	GSH	GSH/(GSH +GSSG) (% value)
Normal	129 \pm 6	123 \pm 7	95.3%
BrCCl ₃	15 \pm 3*	5 \pm 1*	33.3%
BrCCl ₃ +10 ⁻⁶ M Hrx	13 \pm 4	6 \pm 4	46.2%
BrCCl ₃ +10 ⁻³ M Hrx	15 \pm 8	9 \pm 7	60.0%
BrCCl ₃ +10 ⁻⁴ M Hrx	61 \pm 10*	48 \pm 6*	78.7%
BrCCl ₃ +10 ⁻⁴ M Vit. E	39 \pm 8*	27 \pm 6*	69.2%

BrCCl₃: 1.5 \times 10⁻⁴M

Hrx : hydroxybrazilin, Vit E : Vitamine E

Values are the mean \pm SE of three separate experiments

* : Significantly different from normal control group

= : Significantly different from BrCCl₃-treated group

Table 3. Effects of hydroxybrazilin on glutathione levels and GSH/(GSH+GSSG) ratios in BrCCl₃- and menadione-treated hepatocytes.

Groups	GSH+GSSG (nmol glutathione / 3×10^6 cells)	GSH	GSH/(GSH +GSSG) (% value)
Normal	100 \pm 12	90 \pm 11	90.0%
BrCCl ₃	16 \pm 3*	5 \pm 1*	31.3%
BrCCl ₃ +10 ⁻⁴ M Hrx	57 \pm 4*	44 \pm 8*	77.2%
Menadione	57 \pm 5*	30 \pm 2*	52.6%
Menadione+10 ⁻⁴ M Hrx	45 \pm 3	21 \pm 3	46.7%

BrCCl₃: 1.5 \times 10⁻⁴M, Menadione 10⁻⁴M

Hrx : hydroxybrazilin

Values are the mean \pm SE of three separate experiments

* : Significantly different from normal control group

= : Significantly different from BrCCl₃-treated group

The major loss of glutathione has been thought to be the result of the decrease of the reduced form of glutathione during BrCCl₃ metabolism. The highest concentration of hydroxybrazilin (100 μ M) recovered the glutathione loss in the monolayer cultured hepatocytes. And this concentration of hydroxybrazilin was more effective than that of vitamine E. Vitamine E, a well known antioxidant, scavenges free radicals and the hydroxyl group on aromatic ring of vitamine E forms chroma-

noxy or phenoxy radicals.¹⁷⁾ Similarly, it can be proposed that the hydroxyl group on aromatic ring of hydroxybrazilin may donate hydrogen to $\cdot\text{CCl}_3$ radical, thus detoxifying the BrCCl_3 toxicity.

With this concentration, we introduced another radical generating system to compare the effect of hydroxybrazilin on intracellular glutathione loss. Incubation of cultured hepatocytes with menadione (200 μM) for 1 hour resulted in a significant decrease in intracellular reduced form of glutathione but reduction in the total equivalents of glutathione was not comparable to that of BrCCl_3 treatment (Table 3). In isolated hepatocytes, a marked decrease of intracellular glutathione (GSH) level has been observed during the metabolism of menadione.¹⁸⁾ This is known to be related to the formation of $\cdot\text{O}_2^-$ and H_2O_2 caused by redox cycling of the quinone, in particular to the metabolism of H_2O_2 by the selenoprotein glutathione peroxidase, an enzyme which removes peroxides at the expense of GSH.¹⁹⁾ And these processes cause a oxidative stress in the cell. GSH can directly form a conjugate with menadione which is subsequently excreted from the cell⁸⁾ but the menadione-conjugate is not thought to give a major contribution to the glutathione loss.¹⁹⁾ As shown in table 3, 100 μM of hydroxybrazilin had no effect on menadione-intoxicated hepatocytes. Thus, it was thought that hydroxybrazilin could not trap the free radicals formed from the metabolism of menadione. These data suggest that hydroxybrazilin might prevent the hepatotoxicity induced by chemical-derived radicals but not the toxicity linked with oxidative stress.

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