Isolation and Structure Elucidation of a New Glycolipid from the Soft Coral *Lobophytum microlobulatum*Collected from Havellock Island of Andaman and Nicobar Group of Islands

Ammanamanchi S. R. Anjaneyulu*, Nidasanametla S. Kameswara Rao, H. S. Garg¹

Department of Organic Chemistry, Andhra University, Visakhapatnam-530 003, India. 'Central Drug Research Institute, Lucknow-226 001, India.

Abstract – A new glycolipid, 2-hydroxy-3 (octadecyloxy)-propyl-α-D-arabinopyranoside (**1**) has been isolated from *Lobophytum microlobulatum* and its structure has been elucidated by physical and spectral (UV, IR, ¹H, ¹³C NMR, FABMS) data.

Key words - Glycolipid, Lobophytum microlobulatum.

Introduction

We have recently undertaken the chemical examination of the soft coral *Lobophytum microlobulatum* and reported the isolation of four new and five known lobane diterpenoids along with three polyhydroxysteroids (Anjaneyulu, 1996). The isolation of a new glycolipid has also been briefly reported and its structure elucidation is now presented here.

Results and Discussion

More polar fractions (EtOAc: MeOH 19:1) from the column chromatography of the ethyl acetate extract of the soft coral left a residue which on crystallization from chloroform-methanol gave colorless needles, 140 mg, m.p. 125-27°, $[\alpha]_D^{25}$ +15.0° (c 1.4, MeOH). Its molecular formula was fixed as $C_{26}H_{52}O_7$ by elemental analysis and FABMS (+ve mode) m/z 499 (M*+Na). It is sparingly soluble in common organic solvents indicating

The compound on treatment with 3N HCl in methanol on steam bath for 6 hrs. gave an aglycone which was found to be identical in every respect with batyl alcohol (3). The aqueous solution remained after removal of the aglycone was evaporated to dryness in vacuum to obtain the sugar unit which was identified as D-arabinose by its specific rotation $[\alpha]_D^{25}$ -110.0 ° and co-paper chromatography with an authentic sample. Thus the compound was regarded as arabinopyrano-

its polyhydroxy nature and gave positive Molish test for glycosides. Its IR spectrum showed strong broad bands for hydroxyls (3400 cm⁻¹) and strong bands for ether linkage (1160 cm⁻¹). No conjugation was noticed in its UV spectrum. On acetylation with acetic anhydride and pyridine it gave a tetraacetyl derivative (1a) (C₃₄H₆₀O₁₁) as a colorless oil, indicating the presence of four acylable hydroxyls. The acetate showed acetyl groups in its IR spectrum (1735, 1260 cm⁻¹) but no further hydroxylic absorption. The three remaining oxygens of the molecule might be present as ether linkages in the absence of any carbonyl functionality.

^{*}Author for correspondence.

side of batyl alcohol. While batyl alcohol has been found to be more or less an ubiquitous constituent of soft corals, it has not been reported as glycoside so far except for an isolated reference where it was reported as a hexapyranoside from an unidentified species of *Sinularia* (Long, 1988). Neither identification of the hexapyranose nor its spectral details were reported. The present glycolipid has been considered to be new and its spectral details therefore presented.

Its ¹H and ¹³C NMR spectral data could be obtained in pyridine-d₅ at 400 and 22.5 MHz respectively (Table I). Its ¹³C NMR spectrum showed all the 26 carbons whose substitution pattern could be obtained by the DEPT spectrum. Its ¹³C showed four oxymethylene carbons, one of which being the methylene carbon of the sugar unit (δ 64.1) and four oxymethine carbons (δ 70.9, 70.1, 69.8, and 73.2) of which the first three belong to the arabinose unit. The anomeric carbon noticed at 103.3 (d) indicated its α-glyco-

sidic linkage (Agrawal, 1985). The α -glycosidic linkage was also evident from the cou-

Table I. ¹H and ¹³C NMR Spectral Data of

Compound 1								
¹ H NMR (400 MHz)			¹³ C NMR* (22.5 MHz)					
$(pyridine-d_5)$			$(pyridine-d_5)$					
Assignment	Chemical		Carbon No.	Chemical				
Ü	shift	t		shitf				
1'-H	5.45	(d,4)	1'	103.3(d)				
2'-H	4.69	(dd, 4, 8)	2'	70.9(d)				
3'-H	4.56	(dd, 4, 8)	3'	70.1(d)				
4'-H	4,40	(br s)	4^{\prime}	69.8(d)				
5'-H	4.22	(dd,4,8)	5'	64.1(t)				
5'-H	3.98	(dd, 4, 8)	1	71.6(t)				
1-H	4.30	(d,12)	2	73.2(d)				
2-H	4.08	(dd, 2, 12)	3	70.6(t)				
$3-H_2$	4.44	(p)	1"	64.2(t)				
1 "- H_2	3.85	(m)	2"	32.0(t)				
2 "- H_2	3.49	(t,5)	3"	30.1(t)				
Aliphatic chain	1.51	(t,5)	4"-17""	29.9(t)				
Terminal methyl	1.25	(br s)		29.7(t)				
	0.85	(t,7)		29.5(t)				
				26.4(t)				
				22.8(t)				
				14.2(q)				
			18"					

^{*13}C assignments are confirmed by DEPT experiments.

Table II. Comparative ¹H and ¹³C NMR Spectral Data of Compounds 1 and 2

¹H NMR				¹³ C NMR			
Assignment	Sugar part in Compound 1 (py-d ₅ , 400 MHz)	Sugar part in Compound 2 (CDCl ₃ , 360 MHz)	Carbon No.	Sugar part in Compound 1 (py-d ₅ , 22.5 MHz)	Sugar part in Compound 2 (CDCl ₃ , 50 MHz)		
1'-H	5.45(d, 4)	5.12 (d, 2.7)	1'	103.3(d)	104.1		
2'-H	4.69 (dd, 4, 8)	4.14 (br d)	2'	70.9(d)	69.7		
3'-H	4.56 (dd, 4.8)	4.13 (br d)	3'	70.1(d)	69.6		
4'-H	4.40 (br s)	4.07 (br s)	4'	69.8(d)	69.5		
5'-H	4.22 (dd, 4, 8)	4.33 (br d, 12.3)	5'	64.1(d)	69.3		
5'-H	3.98 (dd, 4, 8)	3.83 (br d, 12.3)					
	Aglycone part in 1	Batyl alcohol (py-d ₅ , 90 MHz)		Aglycone part in 1	Batyl alcohol (py-d ₅ , 22.5 MHz)		
1-H	4.30 (d, 12)	4.10 (2H, m)	1	71.6(t)	71.9(t)		
1-H	4.08 (dd, 2, 12)	-	2	73.2(d)	73.5(d)		
2-H	4.44 (q)	4.30 (q)	3	70.6(t)	71.7(t)		
$3-H_2$	3.85 (m)	3.82 (m)	1"	64.2(t)	64.6(t)		
1 "- \mathbf{H}_2	3.49 (t, 5)	3.55 (t, 5)	2"	32.0(t)	32.1(t)		
2 "- H_2	1.51 (t, 5)	1.55 (t, 5)	3"	30.1(t)	30.2(t)		
Aliphatic chain	1.25 (br s)	1.25 (br s)	4"-17"	29.9(t)	29.9(t)		
Terminal methyl	0.85 (t, 7)	0.90 (t, 7)		29.7(t)	29.8(t)		
•				29.5(t)	29.6(t)		
				26.4(t)	26.5(t)		
				22.8(t)	22.9(t)		
			18"	14.2(g)	14.2(q)		

104 Natural Product Sciences

pling constant of the anomeric proton (δ 5.45. d, J=4 Hz). Since the secondary alcoholic group of batyl alcohol was found to exist free, the end primary alcoholic group must have been involved in the glycosidic linkage. The ¹H and ¹³C NMR spectral data of the glycoside could be compared with the 1H and ¹³C NMR data of batyl alcohol, all taken in pyridine-d₅ (Table II). For a comparision of the spectral data of the sugar part with the corresponding protons in the glycoside, a neat spectrum of arabinose could not be obtained in d₅-pyridine. However, the ¹H and ¹³C NMR spectral data of a phenolic arabinoside, seco-pseudoperosine A (2) taken in CDCl₃ was reported in literature (Look, 1987) and the same values have beenz utilized in the table for the comparison. The comparative study of 1H and 13C NMR (Table II) spectral data of compound (1) with those reported for its aglycone and sugar parts strongly supported the proposed structure 2hydroxy-3(octadecyloxy)-propyl-α-arabonopyranoside for it.

Experimental

General experimental conditions, collection & extraction of the organism, and isolation procedure of compound (1) were given in detail in reference 2 (Anjaneyulu, 1006).

2-hydroxy-3(octadecyloxy)-propyl- α -arabonopyranoside (1) – colorless needles, 140 mg, m.p. 125-27, $[\alpha]_D^{25}+15.0^{\circ}$ (c 1.4, MeOH). Found: C, 65.31, H, 11.22; $C_{26}H_{52}O_7$ requires: C, 65.55, H, 10.92%. R_f :0.43 (Hexane: EtOAc 19:1); IR (KBr): 3400, 2900, 2850, 1140, 1080, 1040, 1000 cm⁻¹. FABMS (+ve mode): m/z 499 (M*+Na), 350, 334, 301, 287.

Acetylation of compound (1) – To the compound (20 mg) in pyridine (2 ml) was added acetic anhydride (2 ml) and the mixture was kept aside at room temperature for 10 hours. After usual workup, it gave tetraacetyl derivative as a colourless oil, 10 mg. Found: C, 62.50, H, 9.99; C₃₄H₆₀O₁₁ requires:

C, 63.35, H, 10.92%; IR (CHCl₃): 2900, 2850, 1735, 1260, 1140, 1080 and 1040 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ: 5.34 (1H, br s), 5. 31 (1H, br d, J=4Hz), 5.17 (1H, br d, J=4Hz), 5.14 (1H, br d, J=4Hz), 5.11 (1H, br d, J=4Hz), 3.96 (1H, d, J=10 Hz), 3.82 (1H, dd, J=4, 10 Hz), 3.65 (2H, m), 3.43 (2H, m), 3.55 (2H, m), 2.15 (3H, s), 2.09 (6H, s), 2.01 (3H, s), 1.6 (2H, m), 1.25 (30H, br s) and 0.89 (3H, t, J=7 Hz).

Acknowledgements

Financial assistance from the Department of Ocean Development, New Delhi is gratefully acknowledged. The authors thank Dr. (Mrs.) V. Jayasree, Scientist, NIO, Goa for identifying the soft coral. They also thank the Director CDRI, Lucknow and the Co-ordinator, UGC COSIST & DRS Programme, School of Chemistry, Andhra University, Visakhaptnam for providing spectral data.

References

Agrawal, P. K., Jain, D. C., Gupta, R. K., and Thakur, R. S., Carbon-13 NMR specroscopy of Vol. 2, No. 2, 1996

steroidal sapogenins and steroidal saponins. *Phytochemistry* **24**, 2479-2496 (1985).

- Anjaneyulu, A. S. R. and Kameswara Rao, N. S., Four new lobane diterpenoids from the soft coral Lobophytum microlobulatum of the Havellock Island of the Andaman and Nicobar Islands. Indian J. Chem. 35B, 1294-1303 (1996).
- Long, K., Lin, Y. and Lian, J., Chemical constituents of Chinese soft corals (18), Lochmodoside, a new
- glycoside from Sinularia lochmodes Kolonko. Zhongshan Daxue Xuebao, Ziran Kexueban. 3, 68-72 (1988) Chem. Abstr., 111, 4542M.
- Look, S.A. and Fenical, W., The seco-pseudopterosins, new anti-inflammatory diterpene glycosides from a carrinean gorgonian octocoral of the genus *Pseudogorgia*. *Tetrahedron* **43**, 3363 (1987).

(Accepted October 11, 1996)