

# The Novel Synthetic Substance MR-387C[(2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-prolyl-L-leucine] as an Aminopeptidase M Inhibitor

Myung-Chul Chung, Hyo-Kon Chun, Ho-Jae Lee and Yung-Hee Kho\*

Microbial Chemistry Research Group, Genetic Engineering Research Institute  
Korea Institute of Science and Technology, Taejon 305-600, Korea

(Received August 1, 1994)

**Abstract:** In the course of screening for new aminopeptidase M inhibitors which were expected to be analgesic, immunopotentiating, or anti-metastatic agents, the novel synthetic substance MR-387C[(2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-prolyl-L-leucine] (M.W. 504 daltons) was obtained. It was competitive with the substrate and had an  $IC_{50}$  value of 0.04  $\mu\text{g/ml}$  ( $7.9 \times 10^{-8}$  M) and an inhibition constant ( $K_i$ ) of  $3.8 \times 10^{-8}$  M. This novel MR-387C was compared with various known inhibitors of aminopeptidase M. It inhibited the enzyme more strongly than any other microorganism-originated inhibitor, except probestin.

**Key words:** Aminopeptidase M, inhibitor, synthetic peptide.

Cell-surface metallopeptidases have specific functions that differ according to their cellular locations, including potential roles in the control of growth and differentiation in both hematopoietic and epithelial cell systems (Kenny *et al.*, 1989). Among these metallopeptidases, aminopeptidase M (EC 3.4.11.2, leucine aminopeptidase, microsomal, AP-M) plays an important role in the inactivation process of bioactive peptides, especially enkephalins, which have an analgesic function in cerebral membranes (Gros *et al.*, 1985). Recently, Saiki *et al.* (1993) reported that this enzyme may be partly involved in the activation mechanism for type IV collagenolysis to achieve tumor cell invasion. Menrad *et al.* (1993) reported that bestatin and amastatin, competitive inhibitors of AP-M, inhibit invasion of this enzyme-expressing metastatic melanoma cell line through the reconstituted basement membrane Matrigel in a dose-dependent manner. Thus, it is expected that AP-M inhibitors will act as anti-invasive agents for tumor cells, as well as acting as analgesic agents.

In the course of extensive screening for new AP-M inhibitors, the actinomycete *Streptomyces* sp. SL-387 (KCTC 0102BP) produced MR-387A[(2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl-valyl-prolyl-hydroxyproline] and MR-387B[(2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl-valyl-prolyl-proline] as novel AP-M inhibitors (Chung *et al.*, 1994). On the basis of both MR-387A and B structure, the novel synthetic peptide-like substan-

ce MR-387C[(2S,3R)-3-amino-2-hydroxy-4-phenylbutanoyl-L-valyl-L-prolyl-L-leucine] was obtained. This MR-387C was obtained by chance in searching molecules similar to MR-387A with the same molecular weight. MR-387C inhibited AP-M more strongly than both MR-387A and B. The physico-chemical properties and biological activities of MR-387C are reported herein.

## Materials and Methods

### General methods

Mass spectra were recorded on a Kratos Concept-1S spectrometer. NMR spectra were recorded on a Varian UNITY 300 (300MHz) spectrometer. UV spectra were recorded on a Shimadzu UV-260 spectrophotometer, and IR spectra on a Laser Precision Analytical IFX-65S spectrophotometer. Amino acid analysis was performed on a Pharmacia LKB 4151  $\alpha$  amino acid autoanalyzer. HPLC analysis used a Tosoh TSK 6011 pump with a TSK 6041 detector system.

### Assay for AP-M and inhibitory activities

The inhibitory activity of MR-387C against AP-M was determined by the method of Umezawa *et al.* (1985). Porcine kidney AP-M was purchased from Sigma Chemical Co. (USA). A substrate solution was freshly prepared by dissolving 100  $\mu\text{l}$  of L-leucine-p-nitroanilide stock solution (12.5 mg/ml in DMSO) in 10 ml of 0.1 mM Tris-HCl buffer (pH 7.0). The reaction mixture (total 200  $\mu\text{l}$ ) in a 96 well microplate contained 160  $\mu\text{l}$  of substrate solution, 20  $\mu\text{l}$  of water or aqueous solu-

\*To whom correspondence should be addressed.  
Tel.: (042) 860-4350, Fax: (042) 860-4595

**Table 1.** Physico-chemical properties of the synthetic substance

Appearance	Amorphous colorless powder
FAB-MS (m/z)	505 (M+H) <sup>+</sup>
Color reaction	Ninhydrin
R <sub>f</sub> value*	0.52
UV λ <sub>max</sub> in MeOH (ε)	218(1,580), 252(156), 258(240), 264(235), 268(192), 280(sh.96)
IR (cm <sup>-1</sup> , KBr)	3200, 2960, 1620, 1520, 1440, 1400, 1260, 1100, 1030, 800

\*On silica gel TLC plate (Merck Art No. 5715) with BuOH-MeOH-H<sub>2</sub>O (4:1:2).

tion containing the test compound, and 20 μl of AP-M solution (final 1 mU). The microplate was incubated at 37°C. After 30 min the absorbance at 405 nm was measured using a microplate reader (Bio-Rad model 3550, USA). Percent inhibition was calculated by the formula (A-B)/A×100, where A is the measured value of the enzymatic reaction in the system without an inhibitor, and B is the value with an inhibitor. The IC<sub>50</sub> value is the concentration of inhibitor which produced a 50% inhibition of enzyme activity.

### Synthesis of MR-387C

MR-387C was automatically synthesized by the solid phase method (Stewart and Young, 1969) using an Applied Biosystems (Model 431A) peptide synthesizer on HMP-resin(4-hydroxy-methylphenoxymethyl copoly-styrene-1% divinylbenzene resin). The t-BOC-(2S,3R)-3-amino-2-hydroxy-4-phenylbutanoic acid(t-BOC-AHPA) used in peptide synthesis was purchased from Sigma Peptides and Amino acids (USA). Purification of the inhibitor resulting from a cocktail of synthesis reactions was carried out by HPLC on a Phenomenex ODS column (5C<sub>18</sub>HS, φ10.6×250 mm, 2 ml/min, 30% MeCN-0.1% TFA). Active fractions were combined and concentrated under reduced pressure in a small volume of water, then lyophilized under a freeze dryer to give an amorphous white powder. The R<sub>f</sub> value on silica gel TLC developed with a solvent system BuOH-MeOH-H<sub>2</sub>O (4:1:2) was 0.52.

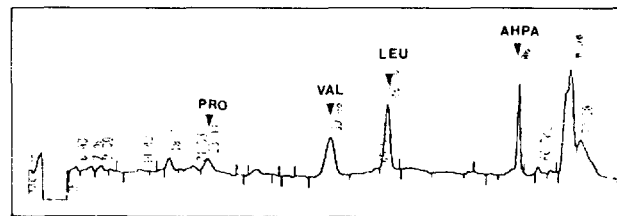
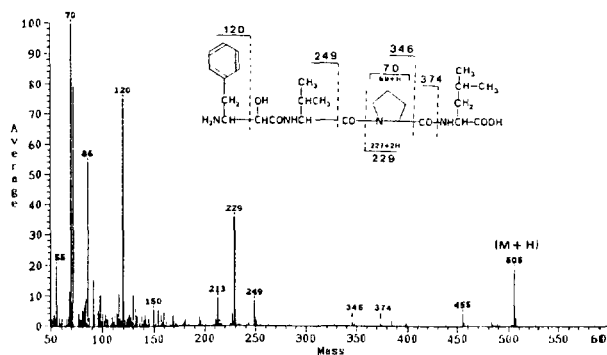
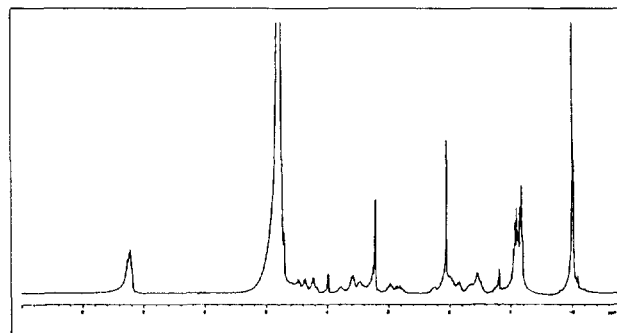
### Hydrolysis of MR-387C

An HCl solution (6 N, 200 μl) was added to a solution of MR-387C (500 μg) and heated at 105°C for 24 h in a sealed tube. The solution was evaporated to dryness. The evaporation was carried out several times following addition of H<sub>2</sub>O to remove HCl.

## Results

### Physico-chemical properties of MR-387C

The physico-chemical properties of MR-387C are su-

**Fig. 1.** Amino acid analysis of the synthetic substance.**Fig. 2.** FAB-MS spectrum of the synthetic substance. (Fast atom; Ar, Kinetic energy; 7 KeV).**Fig. 3.** <sup>1</sup>H-NMR spectrum of the synthetic substance at 300 MHz in CD<sub>3</sub>OD.

mmarized in Table 1. The substance is soluble in water, methanol, and dimethylsulfoxide, but insoluble in acetone, ethylacetate, chloroform, and hexane.

### Identification of MR-387C

Identification of the structure was carried out by amino acid analysis, FAB-MS, and <sup>1</sup>H-NMR spectroscopy. Amino acid analysis of the acid hydrolysate of the compound revealed a molar ratio of AHPA:Val:Pro:Leu of 1:1:1:1 (Fig. 1). The molecular weight of the compound was determined 505 (M+H) by the FAB-MS spectrum (Fig. 2). The assignment of protons by <sup>1</sup>H-NMR (Fig. 3) and <sup>1</sup>H-<sup>1</sup>H COSY data of the compound is as follows: chemical shift (ppm) from TMS in CD<sub>3</sub>OD at 300 MHz, AHPA(2-CH, 3.98d, J=2.4; 3-CH, 3.57m; 4-CH<sub>2</sub>, 2.97dd, 2.83dd, J=8.1, 13.8; Ph-o,m,p, 7.28m); Val(α-CH, 4.21m; β-CH, 2.10m; CH<sub>3</sub>,

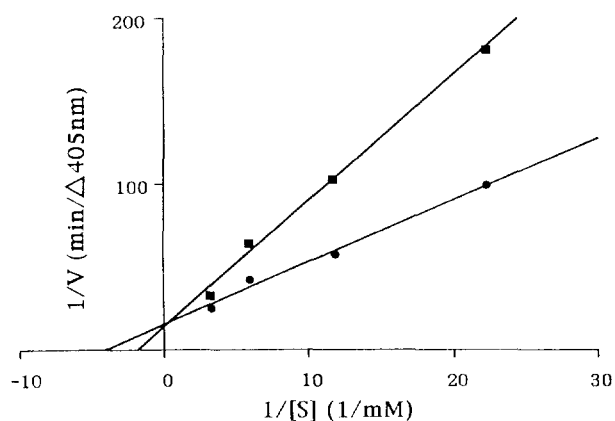
**Table 2.** Inhibitory activities of various inhibitors against aminopeptidase M

Inhibitors	IC <sub>50</sub> (μg/ml)	K <sub>i</sub>	Reference
Actinonin	0.40	1.7 × 10 <sup>-7</sup>	Umezawa <i>et al.</i> , 1985
Bestatin	6.20	4.0 × 10 <sup>-6</sup>	Umezawa <i>et al.</i> , 1976
Leuhistin	0.20	2.3 × 10 <sup>-7</sup>	Aoyagi <i>et al.</i> , 1991
Amastatin	0.58	1.6 × 10 <sup>-6</sup>	Aoyagi <i>et al.</i> , 1978
Probestin	0.03	1.9 × 10 <sup>-8</sup>	Aoyagi <i>et al.</i> , 1990
Synthetic	0.04	3.8 × 10 <sup>-8</sup>	this experiment

## Discussion

Strong inhibition of AP-M requires a 2(S)-hydroxyl group in the N-terminal amino acid derivatives of its inhibitors. The 2(S)-hydroxyl group is responsible for chelation of zinc atoms at the active site of the enzyme (Nishizawa *et al.*, 1977). In order to inhibit leucine aminopeptidase, hydrophobic amino acids such as L-leucine and L-valine as the second residue from the amino end, are better than hydrophilic amino acids (Tobe *et al.*, 1982). In addition, the size of the inhibitor is important in inhibitory activity to AP-M. Rich *et al.* (1984) reported that amastatin[(2S,3R)-3-amino-2-hydroxy-5-methylhexanoyl (AHMHA)-L-valyl-L-valyl-L-aspartic acid] (Aoyagi *et al.*, 1978), a tetrapeptide, is a 100-fold stronger inhibitor of AP-M than bestatin[(2S,3R)-AHPA-L-leucine] (Umezawa *et al.*, 1976) which is a dipeptide. They suggested that AP-M binds tri- and tetrapeptide inhibitors more strongly than dipeptide inhibitors, and that the number of amino acids in the inhibitor affects the binding tightness between the inhibitor and AP-M. The tetrapeptide MR-387C was, therefore, designed having both a 2(S)-hydroxy group in the N-terminal amino acid, and L-valine as the second residue from the amino end, on the basis of the structures of MR-387A and B isolated from the culture filtrate of *Streptomyces* sp. SL-387. The tetrapeptide MR-387C[(2S,3R)-AHPA-Val-Pro-Leu] strongly inhibited AP-M, which supports the idea that increasing the peptide chain length of the inhibitor produces more a potent inhibitor as a consequence of a slower binding process. In addition, the C-terminal amino acid of the tetrapeptide may be an important factor in the activity of inhibitors against AP-M. Probestin[(2S,3R)-AHPA-Leu-Pro-Pro] (Aoyagi *et al.*, 1990) and amastatin have proline and aspartic acid in the C-terminus, respectively. However, probestin inhibits AP-M more strongly than does amastatin (the inhibition constant, K<sub>i</sub> is 1.9 × 10<sup>-8</sup> M in probestin and 1.6 × 10<sup>-6</sup> M in amastatin). MR-387C with leucine in the C-terminus also inhibits AP-M more strongly than does amastatin.

Amastatin, bestatin, actinonin (Umezawa *et al.*, 1985), probestin, and leuhistin (Aoyagi *et al.*, 1991) are specific inhibitors isolated from culture broths of microorganisms. In the course of searching for AP-M inhibitors from soil microorganisms, the new AP-M inhibitors MR-387A[(2S,3R)-AHPA-Val-Pro-Hyp] and B[(2S,3R)-AHPA-Val-Pro-Pro], from the culture broth of *Streptomyces* sp. SL-387 (KCTC0102BP)(Chung *et al.*, 1994) were discovered. On the basis of the MR-387 structure, the novel MR-387C, (2S,3R)-AHPA-Val-Pro-Leu was designed. This specific inhibitor is expected to contribute greatly of studies on the various peptide

**Fig. 4.** Lineweaver-Burk plot of inhibition of AP-M by synthetic substance. ●●: [I]=0 M, ■■: [I]=49.8 nM.

0.85d,  $J=6.6$ ; CH<sub>3</sub>, 0.82d,  $J=6.6$ ); Pro(α-CH, 0.45m; β-CH<sub>2</sub>, 1.86m, 2.22m; γ-CH<sub>2</sub>, ca. 1.93m, 2.02m; δ-CH<sub>2</sub>, ca. 3.44m, 3.75m); and Leu(α-CH, 4.37m; β-CH<sub>2</sub>, 1.52m, 1.58m; γ-CH, 1.67m; CH<sub>3</sub>, 0.88d,  $J=6.5$ ; CH<sub>3</sub>, 0.92d,  $J=6.5$ ).

The amino acid sequence of MR-387C was confirmed by the FAB-MS spectrum (Fig. 2). In the spectrum of the compound the parent peak ( $m/z$  505, M+H) can be recognized. A mass difference of 130 between  $m/z$  504 and  $m/z$  374 corresponds to a loss of leucine from the C-terminus. A peak at  $m/z$  229 was regarded as being derived from elimination of Pro-Leu. A peak at  $m/z$  120 suggested that the N-terminal amino acid is AHPA (Suda *et al.*, 1976). From these results the amino acid sequence of the compound was confirmed to be AHPA-Val-Pro-Leu.

### Biological Properties of MR-387C

The inhibitory activities of this synthetic compound and various AP-M inhibitors are shown in Table 2. The synthetic substance is a competitive inhibitor (Fig. 4). The K<sub>i</sub> and IC<sub>50</sub> values of the compound are 3.8 × 10<sup>-8</sup> M and 0.04 μg/ml (7.9 × 10<sup>-8</sup> M), respectively. The inhibitory activity was stronger than any other inhibitor of microbial origin, except probestin.

processing and disease processes of AP-M.

### References

- Aoyagi, T., Tobe, H., Kojima, F., Hamada, H., Takeuchi, T. and Umezawa, H. (1978) *J. Antibiot.* **31**, 636.
- Aoyagi, T., Yoshida, S., Matsuda, N., Ikeda, T., Hamada, M. and Takeuchi, T. (1991) *J. Antibiot.* **44**, 537.
- Aoyagi, T., Yoshida, S., Nakamura, Y., Shigihara, Y., Hamada, M. and Takeuchi, T. (1990) *J. Antibiot.* **43**, 143.
- Chung, M.-C., Chun, H.-K., Lee, H.-J. and Kho, Y.-H. (1994) *Kor. J. Appl. Microbiol. Biotechnol.*, in preparation.
- Gros, C., Giros, B. and Schwartz, J.-C. (1985) *Biochemistry* **24**, 2179.
- Kenny, A. J., O'Hare, M. J. and Gusterson, B. A. (1989) *Lancet* **2**, 785.
- Menrad, A., Speicher, D., Wacker, J. and Herlyn, M. (1993) *Cancer Res.* **53**, 1450.
- Nishizawa, R., Saino, T., Takita, T., Suda, H., Aoyagi, T. and Umezawa, H. (1977) *J. Med. Chem.* **20**, 510.
- Rich, D. H., Moon, B. J. and Harbeson, S. (1984) *J. Med. Chem.* **27**, 417.
- Tobe, H., Morishima, H., Aoyagi, T., Umezawa, H., Ishiki, K., Nakamura, K., Yoshioka, T., Shimauchi, Y. and Inui, T. (1982) *Agric. Biol. Chem.* **46**, 1865.
- Saiki, I., Fujii, H., Yoneda, J., Abe, F., Nakajima, M., Tsuruo, T. and Azuma, I. (1993) *Int. J. Cancer* **54**, 137.
- Stewart, J. M. and Young, J. D. (1969) *Solid Phase Peptide Synthesis*, W. H. Freeman and Co., San Francisco.
- Suda, H., Takita, T., Aoyagi, T. and Umezawa, H. (1976) *J. Antibiot.* **29**, 100.
- Umezawa, H., Aoyagi, T., Suda, H., Hamada, M. and Takeuchi, T. (1976) *J. Antibiot.* **29**, 97.
- Umezawa, H., Aoyagi, T., Tanaka, T., Suda, H., Okuyama, A., Naganawa, H., Hamada, M. and Takeuchi, T. (1985) *J. Antibiot.* **38**, 1629.