# Further Screening for Antioxidant Activity of Vegetable Plants and Its Active Principles from Zanthoxylum schinifolium

Sook-Im Mun<sup>†</sup>, Hong-Soo Ryu\*, Hee-Jung Lee\* and Jae-Sue Choi\*

Dept. of Food and Nutrition, Dong-Ju Women's Junior College, Pusan 604–080, Korea \*Dept. of Nutrition and Food Science, National Fisheries University of Pusan, Pusan 608–737, Korea

#### Abstract

The antioxidant activity of methanol extracts of thirty plants was tested using the method of 1,1-diphenyl-2-picryl hydrazyl (DPPH) reactivity. Four methanol extracts from Zingiber officinale, Piper nigrum, Zanthoxylum schinifolium and Capsicum annuum were found to be the most effective on DPPH radical scavenging activity. The next effective ones were Perilla frutescens, Sedum sarmentosum, Raphnus sativas, Arctium lappa, Beta vulgaris, Brassica oleracea var. acephala, Brassica juncea in order, and the others did not show a considerable activity. The methanol extract obtained from the seed coats of Zanthoxylum schinifolium was fractionated with several solvents. The interphase materials exhibited the strongest antioxidant activity and was further purified by silica gel and Sephadex LH-20 column chromatography. Two active principles were isolated and identified as quercetin-3-O- $\alpha$ -L-rhamnopyranoside (quercitrin) and quercetin 3-O- $\alpha$ -D-galactopyranoside (hyperoside) by ultraviolet (UV), proton nuclear magnetic resonance ("I-NMR) and carbon nuclear magnetic resonance ("C-NMR). Its antioxidative activity was a little higher than that of L-ascorbic acid.

Key words: antioxidant activity, plant extracts, hyperoside, quercitrin, Zanthoxylum schinifolium

## INTRODUCTION

Synthetic antioxidants such as butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT) has been used to prevent lipid peroxidation, but their applications as food additives are confined to a narrow range because of undesirable effects on the human body1-6). However phenolic antioxidants such as BH-A, BHT, TBHQ, propyl gallate are still used partly as food antioxidants because of their excellent effects and low cost. Therefore, the development of alternative natural antioxidants has been strongly desired, and the natural antioxidants such as  $\alpha$ -tocopherol and Lascorbic acid have attained a high degree of consumer acceptance because of their safety. The antioxidant activities of  $\alpha$ -tocopherol and L-ascorbic acid are, however, lower than those of synthetic antioxidants such as BHA and BHT. Hence, there is a pressing need to find safe, economic antioxidants with high antioxidant activity to replace these synthetic chemicals

In the previous paper<sup>20</sup>, we studied the antioxidative activity of the plants and marine algae and found (+)-catechin along with flavonoids as active principles from *Prunus davidiana*. In this report, we further extends the screening results of thirty vegetables and isolation of the antioxidant principles from the methanol extract of *Zanthoxylum schinifolium* seed coats used as a spice in Korea.

#### MATERIALS AND METHODS

## Instruments

The mps were taken on a Thomas Hoover 6406–H apparatus and are uncorrected. The infra red (IR) spectra were determined in KBr tablets on a Varian Tech-

with the natural sources. Especially, the antioxidant compounds present in edible plants have recently been considered as reasonable food additives. To date, although a large number of reports have appeared in the literature concerning antioxigenic activity of foods<sup>7–15)</sup>, a little data exist regarding antioxigenic principles isolated from foods<sup>16–20)</sup>.

<sup>&</sup>lt;sup>†</sup>To whom all correspondence should be addressed

tron Model 635 spectrophotometer and the ultra violet (UV) spectra were run with CE 599 Universal automatic scanning spectrophotometer. The  $^{1}$ H-NMR (300 MHz) and  $^{13}$ C-NMR (75.5 MHz) recorded with a Brucker-AM 300 spectrometer in DMSO-d<sub>6</sub> containing TMS as an internal standard and chemical shifts are given as  $\delta$ (ppm). Optical rotations were measured on Rudolph Autopol III automatic polarimeter.

#### Materials

Korean pepper, Zanthoxylum schinifolium was gathered from Mugori, Sacheon-Gun, Kyung Sang Nam Do on August 1993. All other samples were purchased at a local market in Pusan at the same time.

1,1-diphenyl-2-picryl hydrazyl (DPPH), 1-ascorbic acid, BHA and BHT were reagent grade, purchased from Sigma. All other reagents were of the highest grade commercially available.

# Measurement of antioxidant or radical scavenging activity<sup>21)</sup>

An 4ml of methanol solution of test extracts at various concentrations (2.5~120g/ml) was added to a solution of DPPH (1.5 × 10<sup>-4</sup>M) in MeOH (1ml), and the reaction mixture was shaken vigorously. After storage at room temperature for 30 minutes in air, the remaining DPPH was determined by spectrophotometry at 520nm. The radical scavenging activity (%) of each sample was expressed by the ratio of lowering of the absorption of DPPH, relative to the absorption of DP-PH solution in the absence of test sample (control). The mean values were obtained from duplicate experiments.

#### **Extraction and fractionation**

Commercially dried seed coats of Zanthoxylum schinifolium (680g) were extracted with hot MeOH under reflux as shown in Scheme 1. The MeOH extracts was partitioned with hexane, CHCl<sub>3</sub>, EtOAc, Bu-OH, and H<sub>2</sub>O successively.

# Isolation of active principles

The interphase materials was subjected to chromatography using SiO<sub>2</sub> (EtOAc: MeOH=gradient) and Sephadex LH-20 (MeOH) columns to yield compou-

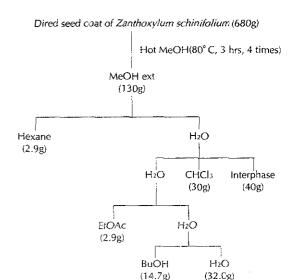
nds 1 and 2 in the order of elution.

## Compound 1(quercitrin)

mp 178–180° C,  $[\alpha]^{25}$ –108 (c=0.1, MeOH); UV  $\lambda$  MeOH, nm (log  $\varepsilon$ ); 260 (4.15), 305 (sh, 3.80), 355 (4.02);  $\lambda$  MeOH+NaOMe; 275 (4, 24), 332 (3.86), d 400 (4.11);  $\lambda$  MeOH+AlCl3 278 (4, 24), 305 (sh, 3.71), 440 (4.16);  $\lambda$  MeOH+AlCl3+HCl 275 (4.16), 305 (sh, 3.75), 360 (3.88), 405 (3.91);  $\lambda$  MeOH+NaOAc 275 (4.17), 325 (sh, 3.88), 370 (3.95);  $\lambda$  MeOH+NaOAc +H3BO3, 265 (4.24), 300 (sh,3.77), 370 (4.06);  $^{1}$ H-NMR (DMSO-d<sub>5</sub>, TMS) $\delta$ ; 12.60 (1H, brs, C5-OH), 7.15 (1H, d, J=2.0, H-2'), 7.11 (1H, dd, J=2.0 and 8.5, H-6;467'), 6.72 (1H, d, J=8.5, H-5'), 6.23 (1H, d, J=2.0, H-8), 6.05 (1H, d, J=2.0, H-6), 5.11 (1H, s, anomeric), 0.80 (3H, d, J=6.0, Me of rhamnose);  $^{13}$ NMR (DM SO-d<sub>6</sub>, TMS); see Table 4

# Compound 2(hyperoside)

mp 253–4° C,  $[\alpha]_{0}^{25}$ –70 (c=0.1, MeOH), IR KBr (cm  $^{-1}$ ); 3200 (OH), 1650 (C=O), 1600, 1540, 1500 (C=C), 1075, 1050 (C-O), 1015, 990, 928, 880, 855, 818, 785; UV  $\lambda$  MeOH nm (log  $\varepsilon$ ); 256 (4.10), 268 (sh, 4.00), 295 (sh, 3.69), 360 (4.02);  $\lambda$  MeOH + NaOMe 272 (4.15), 330 (3.73), 410 (4.11);  $\lambda$  MeOH + AlCl<sub>3</sub> 272 (4.16), 302 (3.64), 333 (3.50), 431 (4.14);  $\lambda$  MeOH + AlCl<sub>3</sub> + HCl 268 (4.14), 300 (3.76), 360 (3.94),



Scheme 1. Extraction and fractionation of Z. shinifolium.

398 (3.96) ;  $\lambda$  MeOH + NaOAc 272 (4.10), 325 (3.81), 372 (3.95) ;  $\lambda$  MeOH + NaOAc 272 (4.10), 325 (3.81), 372 (3.95) ;  $\lambda$  MeOH + NaOAc + H<sub>3</sub>BO<sub>3</sub> 260 (4.16), 296 (3.60), 380 (4.06) ; 'H-NMR (DMSO-d<sub>6</sub>, TMS) $\delta$ ; 12.60 (1H, brs, OH), 10.77 (1H, brs, OH), 9.80 (1H, brs, OH), 9.03 (1H, brs, OH), 7.66 (1H, dd, J=2.0 and 8.5, H-6'), 7.53 (1H, d, J=2.0, H-2'), 6.84 (1H, d, J=8.5, H-5'), 6.40 (1H, d, J=2.0, H-8), 6.19 (1H, d, J= 2.0, H-6), 5.36 (1H, d, J=7.0, anomeric) ; <sup>31</sup>C-NMR(DN-SO-d<sub>6</sub>, TMS) $\delta$ ; see Table 4

## Acid hydrolysis of 1 and 2

Ten mg of samples was refluxed with 5% H<sub>2</sub>SO<sub>4</sub> (30ml) for 5 hr. After cooling, the reaction mixture

was filtered. The aglycone was crystallized from Me-OH to afford quercetin as yellow needles, mp 312–5. It was confirmed by direct comparisons with an authentic samples (TLC and mmp). The filtrate was neutralized with BaCO<sub>3</sub>, filtered and concentrated. L-rhamnose from 1 and D-galactose from 2 was identified by TLC (precoated cellulose, pyridine: EtO Ac: HO-Ac: H<sub>2</sub>O=36: 36: 7: 21, Rf 0.65 and 0.40)

## **RESULTS AND DISCUSSION**

# Antioxidant activity of vegetable plants

To find the antioxidant materials from plants , the free radical scavenging activity was evaluated by the

Table 1. Free radical scavenging effects of several edible plants on DPPH

English name	Scientific name	Part	50% reduc. (mg)	
Burdock	Arctium lappa	seed	160.2	
Chard	Beta vulgaris seed		61.6	
Kale	Brassica oleracea var. acephala	seed	186.2	
Korean çabbage	Brassica campestris var. pekinensis	seed	210.2	
lettuce	Lactuca sativa	seed	323.0	
Black pepper	Piper nigrum	seed	52.8	
Korean peper	Zanthoxylum shinifolium	seed coat	59.0	
Redish rod	Raphnus sativas	seed	148.7	
Leaf mustard	Brassica juncea	seed	194.0	
Mallow	Malva verticillata	seed	480<	
Spinach	Spinacia oleracea	seed	267.2	
Sedum	Sedum sarmentosum	leaf	148.0	
Crown daisy	Chrysanthenum coronarium	leaf	347.9	
Amaranth	Amaranthus margostanus	leaf	480<	
Wild dropwort	Oenanthe stolonifera	leaf	338.0	
	Isodon japonicus	leaf	246.2	
Green perilla	Perilla frutescens	leaf	264.3	
	Perilla frutescens	seed	145.0	
Parsley	Petroselium sativum	leaf	480<	
Onion	Allium cepa	rhizome	480<	
Garlic .	Allium sativum	semen	480<	
Leek	Allium tuberosum	leaf & stem	480<	
Large Geen onion	Allium fistulosum	leaf & stem	480<	
Ginger root	Zingiber officinale	rhizome	49.6	
Butterbur	Petasites japonicus	leaf	206.8	
	Saururus chinensis	leaf	460.0	
Jujube	Zizypus jujuba	seed	480<	
Red peper	Capsicum annuum	seed	84.6	
	Youngia sonchifolia	leaf & stem	220.0	
Dandelion	Taraxacum platycarpum	l <del>e</del> af	302.6	
Doraji (Root of Chiness bellflower)	Platycodon grandiflorum	root	480<	
	внт		9.5	
	L-ascorbic acid		8.1	

<sup>\*</sup>Amount required for 50% reduction of DPPH after 30min

scavenging effect of DPPH radical. The control intensity (absence of sample extracts) was taken as 100%, and the percentage intensity was calculated. The concentration of each fraction for 50% free radical inhibition is shown in Table 1.

As shown in Table 1, some plant extracts such as *Perilla frutescens, Sedum sarmentosum, Raphnus sativas, Arctium lappa, Beta vulgaris,* and *Brassica juncea* exhibited somewhat high scavenging effects on DPPH. The most effective ones were *Zingiber officinale, Piper nigrum, Zanthoxylum schinifolium,* and *Capsicum annuum.* These results suggest that these spices contained a certain antioxidant (s).

# Antioxidant activity and active principles of Zanthoxylum schinifolium

The present study was also carried out to investigate the active principles in MeOH extract of Zanthoxylum schinifolium, whose seed coat showed marked antioxidant activity. The MeOH extract of Zanthoxylum schinifolium seed coat was partitioned by hexane, CHCl<sub>3</sub>, EtOAc, BuOH, interphase materials and water successively. And then, these solvent-soluble fractions were measured free radical scavenging effect on DPPH. As the result of them, interphase materials showed the strongest antioxidant activity (Table 2). This

Table 2. Effects of several fraction of the methanol extract from Zanthoxylum shinifolium on DPPH

Fractions	50% reduc." (mg)	
Hexane	375.2	
Interphase materials	9.1	
CHCl <sub>3</sub>	140.8	
<b>E</b> tOAc	10.2	
BuOH	34.0	
H <sub>2</sub> O	91.6	
L-ascorbic acid	14.7	

<sup>&</sup>lt;sup>a</sup> Amount required for 50% reduction of after 30min.

Table 3. Effects of isolated compounds from Z. shinifolium on DPPH

	Compounds	50% reduc. <sup>a)</sup> (mg)		
	Quercitrin	6.50		
	Hyperoside	8.45		
4	L-ascorbic acid	14.7		

Amount required for 50% reduction of DPPH after 30min.

fraction was further purified to obtain active compounds 1 and 2 by repeated silica gel and gel filtration column chromatography. The two compounds 1 and 2 was identified as quercitrin and hyperoside, respectively. Table 3 summarized the radical scavenging results of isolated compounds on DPPH. Their antioxidant effects were a little higher than those of ascorbic acid.

Antioxidant activities of various flavonoids are well known. As to flavonoids, the relationship between the position of the hydroxyl groups and the antioxidant activity has been discussed<sup>22</sup>. Quercetin is an effective flavonols in the same ways as morin, kaempferol, and luteolin<sup>23</sup>. Quercitrin and hyperoside isolated from the methanol extract of *Z. schinifolium* also may be usable as an antioxidant component. The findings of the present study indicate that the methanolic extract of *Z. schnifolium* seed coats and its components (quercitrin and hyperoside) may be useful for antioxidant.

## Structure elucidation of active principles

The interphase materials were subjected to chromatography using SiO<sub>2</sub> and Sephadex LH-20 to yield compounds 1 and 2 in the order of increasing polarity.

Compound 1, mp 178–180° C and compound 2, mp 254–6° C, showed positive Mg + HCl and Molisch tests. Acid hydrolysis of each compound afforded as the aglycone, quercetin, mp 315–6 and as the sugar, L-rhamnose from compound 1 and D-galactose from compound 2. The <sup>1</sup>H-NMR and <sup>13</sup>C-NMR (Table 4) spectra showed only one anomeric proton signal, in-

Table 4. 13C-NMR chemical shift of compounds 1 and 2

Carbon No.	1	2	Carbon No.	1	2
2	157.2	156.2	3′	145.1	144.7
3	134.2	133.5	4′	148.3	148.3
4	1771.7	177.4	.5′	115.6	115.7
5	161.3	161.1	6′	121.0	121.0
6	98.6	98.5	1"	101.7	101.6
7	164.1	164.0	2"	70.0°	71.1
8	93.5	93.4	3"	70.34	73,0
9	156.4	156.2	4"	71.2	67.6
10	103.9	103.6	5"	70.5 <sup>a)</sup>	75.7
1′	120.7	121.6	6"	17.4	60.0
2′	115.4	115.1			

<sup>&</sup>quot;assignments may reversed

dicating the presence of one mole of sugar in each compound.

The UV spectrum of each compound, exhibiting band I peak at 355-360nm, was very similar to those reported for a number of 3-hydroxy substituted flavonols24. A bathochromic shift of band I in the presence of AlCl3 or AlCl3+HCl and of band II in the presence of NaOAc indicated the presence of free 5-hydroxyl and 7-hydroxyl groups. And also a bathochromic shift with NaOMe, without a decrease in intensity, showed the presence of a free 4'-hydroxyl group. It was thus, suggested that the sugar might be attached to 3-hydroxyl group. The 13C-NMR spectrum of each compound confirmed this suggestion. The configuration and conformation of sugar moiety was determined by the I value of the anomeric proton signal. Compounds 1 and 2 were, therefore, identified as guercetin 3-O-α-L-rhamnopyranoside (quercitrin) and quercetin  $3-O-\beta-D$ -galactopyranoside (hyperoside), respectively.

#### REFERENCES

- Branen, A. L.: Toxicology and biochemistry of butylated hydroxyanisole and butylated hydroxytoluene. J. Am. Oil. Chem. Soc., 52, 59 (1975)
- Wattenberg, L. W.: Inhibition of carcinogenic and toxic effects of polycyclic hydrocarbons by phenolic antioxidant and ethoxyquin. J. Nat. Cancer Inst., 48, 1425 (1972)
- Ito, N., Fukushima, S. and Tsuda, H.: Carcinogenicity and modification of the carcinogenic response by BHA, BHT, and other antioxidants. CRC Crit. Rev. Toxicol., 15, 109 (1985)
- Ponder, D. L. and Green, N. R.: Effects of dietary fats and butylated hydroxytoluene on mutagen activation in rats. Cancer Res., 45, 558(1985)
- Selvan, R. S. and Rao, A. R.: Influence of butylated hydroxyanisole on oocyte depletion induced by 7,12– dimethylbenz[a]anthracene in mice. *Indian J. Exp. Biol.*, 23, 320 (1985)
- Takahashi, O., Sakamoto, Y. and Hiraga, K.: Lung hemorrhagic toxicity of butylated hydroxyanisole in the rat. *Toxicol. Lett.*, 27, 15 (1985)
- Yu, J. H., Cho, C. M., Oh, D. H. and Pyun, Y. R.: Antioxidant properties of red-pepper peel extracts on margarine. Korean J. Appl. Microbiol. Bioeng., 9, 21(1981)
- Kasuga, A., Aoyagi, Y. and Sugahara, T.: Antioxidant activities of edible plants. Nippon Shokuhin Kogyo Gakkashi, 35, 22 (1988)
- 9. Oh, M. J., San, H. Y., Kang, J. C. and Lee, K. S.:

- Antioxidative effect of Pueraria root extract on edible oils and fats. J. Korean Soc. Food Nutr., 19, 448 (1990)
- Hemeda, H. M. and Klein, B. P.: Effects of naturally occurring antioxidants on peroxidase activity of vegetable extracts. J. Food Sci., 55, 184 (1990)
- Maeng, Y. S. and Park, H. K.: Antioxidant activity of ethanol from Dodok (Codonopsis lanceolata). Korean J. Technol., 23, 311 (1991)
- Lee, J. S. and Lee, S. W.: The studies of fatty acids and antioxidant activities in parts of Omija (Schizandra chinensis Baillon). Korean J. Dietary Culture, 6, 143 (1991)
- Economu, K. D., Oreopoulou, V. and Thomopoulos, C. D.: Antioxidant activity of some plant extracts of the family Labiate. J. Am. Oil. Chem. Soc., 68, 109 (1991)
- Choi, Y., Shin, D. H., Chang, Y. S. and Shin, J. I.: Screening of natural antioxidant from plant and their antioxidative effect. Korean J. Food Sci. Technol., 24, 142 (1992)
- Jitoe, A., Maxuda, T., Tengah, I. G. P., Suprapta, D. N., Gara, I. W. and Nakatani, N.: Antioxidant activity of tropical ginger extracts and analysis of the contained curcuminoids. J. Agric. Food Chem., 40, 1337(1992)
- Han, Y. B., Kim, M. R., Han, B. H. and Ham, Y. N.: Studies on antioxidant component of mustard leaf and seed. Kor. J. Pharmacogn., 18, 41 (1987)
- Osawa, T. and Namiki, M.: Natural antioxidants from Eucalyptus leaf waxes. J. Agric. Food Chem., 33, 777 (1985)
- Lorenzo Jose De Rosenzweig Pasqrel and Babbit, J. K.: Isolation and characterization of natural antioxidant from shrimp (Pandalus joddani). J. Food Sci., 56, 143 (1991)
- Park, J. H., Kang, K. C., Baek, S. B., Lee, Y. H. and Rhee, K. S.: Separation of antioxidant compounds from edible marine algae. Korean J. Food Sci. Technol., 23, 256 (1991)
- Choi, J. S., Lee, J. H., Park, H. J., Kim, H. G., Young, H. S. and Mun, S. I.: Screening for antioxidant activity of plants and marine algae and its active principles from Prunus davidiana. Kor. J. Phrmacogn., 24, 299 (1993)
- Blos, M. S.: Antioxidant determinations by the use of a stable free radical. *Nature*, 181, 1199 (1958)
- Pratt, D. E.: Role of flavones and related compounds in retarding lipid-oxidative flavor changes in food. In "Phenolic, sulfur, and nitrogen compounds in food flavors" Charalambous, G. and Katz, I. (eds.), American Chemical Society, Washington, D.C., p.1 (1976)
- 23. Torel, J., Cillard, J. and Cillard, P.: Antioxidant activity of flavonoids and reactivity with peroxy radical. *Phytochemistry*, **25**, 383 (1986)
- Marby, T. J., Markham, K. R. and Thomas, M. B.: "The Systematic Identification of Flavonoids", Springer, N.Y., p.41 (1970)

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# 식용식물의 항산화 효과 검색과 산초의 항산화 성분

문숙임\*·류홍수\*·이회정\*·최재수\*

동주여자전문대학 식품영양과\*부산수산대학교 식품영양학과

## 요 약

30여종의 식용식물 메탄을 엑스에 대한 항산화 효과를 1,1-diphenyl-2-picrylhydrazyl (DPPH)을 사용하여 검색하였다. 생강, 산초, 후추, 고추 메탄을 엑스에서 DPPH radical을 소거하는 효과가 가장 강하게 나타 났으며 기타, 들깨, 돌나물, 머위, 쑥갓, 방아, 돌미나리, 배추씨에서는 그 효과가 다소 미약하였다. 산초 종 피의 여러 용매 추출 분획물 중에서 interphase 분획물이 free radical 소거 효과가 가장 현저하였으며 interphase 분획물을 silica gel 및 Sephadex column chromatography 하여 분리한 quercitrin과 hyperoside는 산초 종피의 항산화 활성성분들로 밝혀졌으며 이들은 L-ascorbic acid 보다 그 효과가 다소 높았다.