의공학회지: 제15권, 제1호, 1994 J. of KOSOMBE: Vol. 15, No. 1

연 구 논 문 94-15-1-02

Targets of Antithrombotic and Fibrinolytic Therapy

Sun-Ok Kim*

INTRODUCTION

The delicate balance between thrombotic and anti-thrombotic activities in maintaining hemostasis while preventing thrombosis is regulated by a variety of cells and by specific cell-mediated mechanisms. The endothelial cell plays an active role in the negative control of thrombus formation. For example, the endothelial cell synthesizes both prostacyclin¹⁾ and endothelial cell-derived relaxing factor²⁾ which inhibit platelet activation and vasoconstriction. Plasminogen activator, which initiates blood clot lysis, is also synthesized and released from endothelium³⁾.

However, when the vascular endothelium is damaged platelets become activated and undergo a series of changes: adhesion, shape change, secretion of constituents of their storage granule such as ADP, serotonin and arachidonate metabolites, thromboxane A₂(TXA₂) and prostaglandin H₂ (PGH₂), further aggregation to form a hemostatic platelet plug⁴). With the changes in platelet membrane during secretion and aggregation, blood coagulation pathway is also activated⁵). The generated thrombin is a powerful platelet stimulant as well as having the ability to catalyze fibrin formation, which consolidates the plug with strands of fibrin finishing the hemostasis process.

On the other hand, the same process governing hemostasis is apparently involved in the progression of atherosclerosis, acute coronary artery syndromes such as unstable angina, myocardial infarction, and sudden ischemic death and also in cerebrovascular disease causing stroke^{6, 7)}.

Thus, any damage to the vessel wall from atherosclerosis, thermal injury, or infectious agents, etc. is interpreted by this process in the same way as when a vessel is damaged. The resulting platelet activation may result in formation of mural or occlusive thrombi. When the location of the thrombus occurs in the heart or brain, it may be immediately fatal or seriously dehabilitating.

In order to prevent platelet activation in this pathological process, several approaches have been used as antithrombotic therapy, namely, inhibition of von Willibrand factor (vWF)-Glycoprotein (GP)Ib⁸, fibrinogen-GPIIbIIIa⁹, thrombin activation¹⁰, and thromboxane A2/prostaglandin H2(TXA2/PGH2) activation¹¹. Also, for the treatment of thrombosis, several plasminogen activators (PAs) such as tissue-type PA (t-PA) and urokinase-type PA (u-PA, two-chain and single-chain) and streptokinase (SK) have been applied to dissolve the fibrin clot as fibrinolytic therapy¹².

TARGETS OF ANTITHROMBOTIC THERAPY

Several receptors on platelet membrane mediate platelet responses upon stimulation. These include GPIb for vWF, GPIIbIIIa for fibrinogen, TXA2/PGH2 receptor and thrombin receptor. Because of the importance of these receptors in platelet responses, inhibitors of these receptors can be developed as effective antithrombotic agents. Understanding of the molecular mechanism of the ligand-receptor interaction has greatly aided development of the new antit-

^{*} Genetic Engineering Research Institute, KIST

hrombotic agents.

vWF-GPI_b

The first event related to the participation of vWF in thrombogenesis is the recognition of surface-bound vWF by platelets through the GPI_b receptor. This not only promotes the initial platelet contact with the thrombogenic surface but also mediates activation of GPII_bIII_a, which supports platelet aggregation⁽³⁾.

Therefore, it is apparent that the selective inhibition of vWF binding to GPI_b may be an appropriate early intervention likely to result in effective reduction of the thrombi formed on exposure of subendothelial vWF to flowing blood.

Progress has been made in identifying the binding domains of vWF and GPIb, which can be used for the rational design of the selective inhibitors of vWF-GPI_b interaction. A tryptic fragment of 52/48 kD comprising residues Val⁴⁴⁹-Lys⁷²⁸ has been demonstrated to interact with GP Ib14). Within the GP Ib binding domain of vWF, two discontinuous sequences corresponding to Cys474-Pro488, Leu⁶⁹⁴-Pro⁷⁰⁸ are thought to be involved in GP Ib binding¹⁵⁾. These peptides which are separated in the linear sequence by 205 residues and maintained in close spacial proximity in the folded molecule by disulfide bonding appear to be functionally optimal in combination (Fig. 1). In this regards, recombinant GPIb-binding domain of vWF (rvWF⁴⁴⁵⁻⁷³³), synthetic peptides representing linear sequence of vWF involved in the GPI_b binding (Cys⁴⁷⁴-Pro⁴⁸ 8, Leu⁶⁹⁴-Pro⁷⁰⁸) and monoclonal antibody against binding domains of vWF and GPIba can be used to inhibit vWF-GPIb interaction. Further studies on the structurefunction relations using site-directed mutagenesis is likely to provide the information necessary for the design of peptides and small organic molecules aimed at specifically interfering with platelet adhesion and aggregation.

Fibrinogen-GPIIIIIIa

Fibrinogen binding to platelet GPII_bIII_a is activation dependent, which includes the conversion of the platelet glycoprotein GPII_bIII_a from latent state to active conformation¹⁶⁾. As a consequence of fibrinogen binding to its receptor, platelet aggregation can ensue, which is a final com-

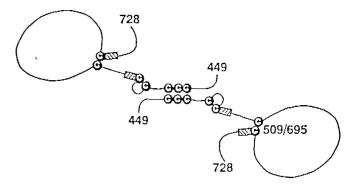


Fig. 1. Schematic representation of the glycoprotein Ib-binding domain of yon Willibrand Factor

mon pathway shared by a variety of stimuli, such as ADP, thrombin or epinephrine¹⁷⁾. Platelet aggregation is a key event in thrombus formation and under certain pathologic conditions, platelet aggregation can result in thrombus formation and ultimately stenosis of the vessel. Thus, antagonists of fibrinogen binding to GPII_bIII_a are potential therapeutic agents.

The first fibrinogen receptor antagonists were monoclonal antibodies, e.g., PAC-1¹⁸⁾ and 10E5¹⁹⁾ or F(ab')₂ which bind to GPII_bIII_a with high affinity and inhibit platelet aggregation²⁰⁾. In view of the physiologic and pathophysiologic significance of ligand binding to GPII_bIII_a, major emphasis has been placed on identifying fibrinogen-GPII_bIII_a binding sites. Two sets of peptides, corresponding to specific amino acid sequences within fibrinogen referred as "recognition peptides", RGD²¹⁾ and gamma chain peptides²²⁾, inhibit ligand binding to GPII_bIII_a and platelet aggregation.

Besides, potent platelet aggregation inhibitors termed "disintegrin" have been isolated from a wide variety of viper venom²³. These peptides all contain RGD recognition sequence with one exception (KGD sequence in case of barbourin with apparent GPIIbIIIa specificity²⁴) and high cysteine content conserved in primary sequence (Fig. 2). However, these are 500~1,000 times more potent inhibitors of platelet aggregation than linear RGDX peptides, which indicates that other structural determinants than recognition sequence are important for binding. The solution structure of kistrin²⁵ which is a member of disintegrin superfamily has been demonstrated a series of tightly packed

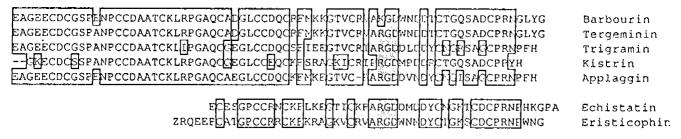


Fig. 2. Sequence alignment of viper venom GPII_bIII_a antagonists (disintegrin)

loops held together by disulfide bonds and RGD recognition sequence located at the apex of a long loop across the surface of the protein. These results indicate favorable conformational restraints of the RGD recognition sequence is required for enhanced binding to GPIIbIIIa and aided in the design of small synthetic peptide GPIIbIIIa antagonists.

Modification of linear recognition sequences has yielded several peptide antagonists and their structure-activity relationship has been studied^{26, 27)}. Only modest improvements in potency were achieved with linear analogues of pentapeptide, Ac-Gly-Arg-Gly-Asp-Val-OH, while dramatic results were achieved with constraining the conformation of peptides by cyclization, culminating in the cyclic disulfide Ac-Cyclo-S,S-(Cys-(N-Me)Arg-Gly-Asp-Pen)-NH₂ and Cyclo-S-Ac-(D-Tyr)-Arg-Gly-Asp-Cys-OH(sulfoxide bridge) which yielded equipotent antagonists as kistrin (Fig. 3).

Therefore, these "recognition peptides" and potent mimetics of these peptides have a potential as antithrombotic drugs.

TXA₂/PGH₂ activation

The arachidonic acid metabolites PGH_2 and TXA_2 released by platelets in response to a variety stimuli are potent vasoconstrictor and platelet activating agents, which serve as an amplification mechanism in the platelet activation sequence²⁸.

The profound biological actions of TXA₂/PGH₂ suggest its putative role as a pathological mediator of cardiovascular disease²⁹. Several clinical studies have suggested that TXA₂ is involved in a number of cardiovascular diseases as demonstrated by measuring the metabolite, TXB₂. Clearly,

Ac-Arg-Gly-Asp-Val-NH₂ Ac-Cyclo-S, S-(Cys-Arg-Gly-Asp-Cys)-NH₂ Ac-Cyclo-S, S-(Cys-(N-Me)Arg-Gly-Asp-Pen)-NH₂

Fig. 3. Structure of small peptide GPII_bIII_a antagonists

the regulation of TXA₂/PGH₂ synthesis or its interaction at a specific receptor has potential clinical implications.

There have been several approaches to the control of PGH₂/TXA₂ activation: cyclooxygenase inhibition, thromboxane synthase inhibition, and PGH₂/TXA₂ receptor antagonism³⁰⁾. However, cyclooxygenase inhibitors such as

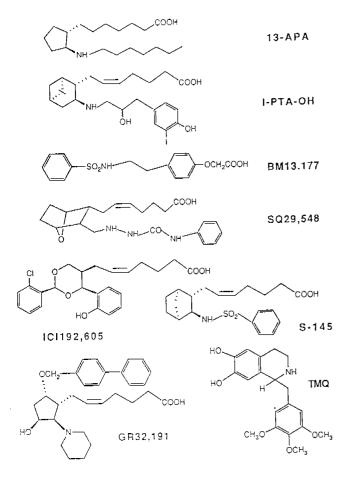


Fig. 4. Structure of TXA₂/PGH₂ receptor antagonists

aspirin can have the dual effect of reducing proaggregatory PGH₂/TXA₂ and antiaggregatory PGI₂. Also, in clinical trials thromboxane synthase inhibitors have been disappointing when compared to their efficacy *in vitro*. An explanation for the weak activity for thromboxane synthase inhibitors *in vivo* is that prostaglandin endoperoxide may substitute for the proaggegatory effect of TXA₂.

In this regard, TXA₂/PGH₂ receptor antagonism is the most plausible approach to selectively inhibit platelet activation induced by TXA₂/PGH₂. A number of TXA₂/PGH₂ receptor antagonists of diverse structure and potency have been reported (Fig. 4). The majority of TXA₂/PGH₂ receptor antagonists may be considered a typical alpha-side chain, but often with substantial modification of the omega-side chain. Of the many TXA₂/PGH₂ receptor antagonists, a few have been studied in humans, which include AH 23, 848, GR 32,191, BM 13,177, BM 13,505, SQ 31,491 and

ICI 192,605³¹⁾. Another approach to inhibit TXA₂/PGH₂ activation is dual inhibitors of thromboxane synthase and TXA₂/PGH₂ receptor by combining thromboxane synthase inhibition with thromboxane receptor antagonism in one drug³²⁾. These may provide benefits from the redirection of the prostaglandin metabolism to prostacyclin and PGD₂, which further increase the inhibitory potential on platelet and vasospasm.

Thrombin activation

In addition to its well-known role in cleaving fibrinogen to fibrin, thrombin is able to evoke biological responses from a variety of cells, including platelets, endothelial cells, and fibroblasts³³⁾. These multiple cell activating functions of thrombin may lead to orchestrate the response to vascular injury, mediating not only hemostatic but inflammatory and reparative responses. Platelet activation by thrombin at the sites of vascular injury, which is the most potent stimulator of platelet aggregation, is believed to be a critical event in arterial thrombosis34). Identification of the thrombin receptor revealed a unique proteolytic mechanism of receptor activation (Fig. 5)35, 36) and led to the development of a novel agonist peptide capable of activating the thrombin receptor independent of thrombin and thrombin's protease activity500 and will allow the development of thrombin receptor antagonists.

The anticoagulant effect of unfractionated heparin and most low molecular weight heparin is mediated primarily by enhancing the inactivation of thrombin by antithrombin III (AT III)³⁷⁾. However, thrombin which is bound to fibrin clot remains fully active and is resistant to inhibition by the AT III-dependent thrombin inhibitors³⁸⁾. In this regards, the AT III-independent thrombin inhibitors³⁹⁾ have been developed. These include: hirudin, hirulog, synthetic thrombin inhibitors, activated protein C and thrombin receptor antagonists.

Hirudin, the most potent natural inhibitor of thrombin, was isolated from the European medicinal leech *Hirudo medicinalis*⁴⁰, which forms a stable noncovalent 1:1 stoichiometric complex with thrombin (Ki 10⁻¹¹ to 10⁻¹⁴ M). Hirudin consists of a compact NH₂-terminal head and a long polypeptide COOH-terminal tail, which has been shown to interact with active site and anion binding exos-

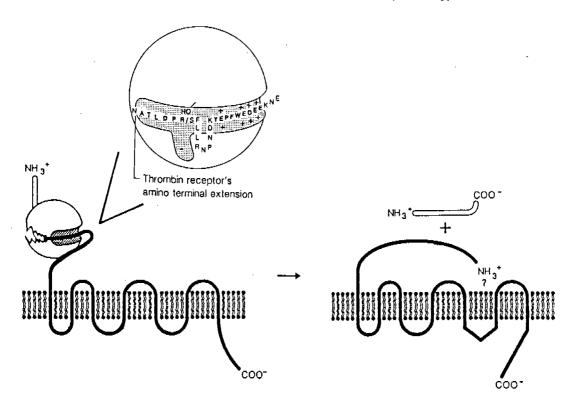


Fig. 5. Thrombin's binding interaction with its receptor and model of thrombin receptor activation

ite of thrombin, respectively (Fig. 6)^{41, 42)}. Based on the structure of hirudin, bivalent synthetic peptide called hirulogs have been designed⁴³⁾. Hirulogs consist of a catalytic-site inhibitor moiety ((D-Phe)-Pro-Arg-Pro), a flexible spacing segment ((Gly)n) and an anion binding exosite recognition moiety (COOH-terminal hirudin peptides) (Fig. 7). Studies on the optimal length of the oligoglycyl spacer showed that at least 3-4 glycines were necessary for optimal inhibitory activity. The anticoagulant activity of Hirulog-1 ((gly)4) has shown to be comparable to that of hirudin. Besides the sequence ((D-Phe)-Pro-Arg-Pro), other structures such as nonpeptide groups or other peptide sequences could be employed for a catalytic-site binding moiety.

The synthetic low molecular weight thrombin inhibitors act by blocking the active site of the thrombin to eliminate its enzymatic activity. The catalytic mechanism of the serine protease, thrombin, and its substrate specificity allow the design of several types of inhibitors⁴⁴. These can be classified as substrate-derived competitive inhibitors and mechanism-based inhibitors according to the kinetics of in-

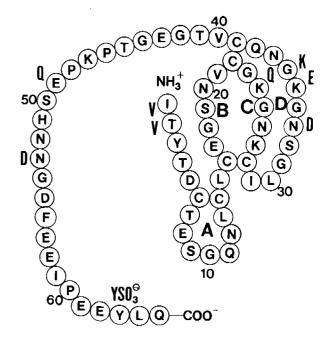
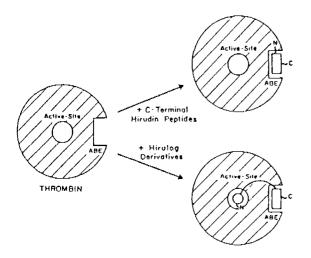


Fig. 6. The sequence of recombinant hiruding varient 2-Lys⁴⁷ $(rHV2-Lys^{47})$

hibition and the types of binding. A key structural feature of these inhibitors is arginine or a group mimicking this



(D-Phe)-Pro-Arg-Pro-(Gly)n-NGDFEEIPEEYL

Fig. 7. The sequence of hirulog and illustrative scheme of thrombin inhibition

amino acid, which governs the primary affinity of thrombin substrates. Figure 8 illustrates the structures of the most potent compounds of arginine- and benzamidine-derived inhibitors, namely, argipidine⁴⁵⁾, αNAPAP⁴⁶⁾ and PPACK⁴⁷⁾.

Thrombin also triggers anticoagulant pathway by binding to thrombomodulin⁴⁸⁾. This initiate rapid protein C activation (Fig. 9). Activated protein C then complexes with protein S on the surface of either platelets or the endothelium. These complexes catalyzed the proteolytic inactivation of factors Va and VIIIa, which results in inhibition of thrombin formation. Besides, thrombomodulin accelerate the inhibition of thrombin by antithrombin III⁴⁹⁾. Therefore, activated protein C or regulation of thrombomodulin activity would prevent the coagulation responses.

Thrombin receptor blockers would be the most useful to prevent cell activation initiated by thrombin. Monoclonal antibodies against receptor's hirudin-like sequence, thrombin cleavage site or yet unidentified agonist peptide binding site can be obtained to block receptor activation⁵¹. In addition, uncleavable peptides mimicking thrombin receptor domain or competitive antagonists of the agonist peptide and their structure-activity relationship should be explored⁵²). Further studies are required to elucidate arterial wall properties leading to thrombin generation and characteristics of thrombin inhibition for the development

Fig. 8. Structure of prototype compounds of the arginine derivative, benzamidine derivative, and tripeptide-type synthetic thrombin inhibitors

of a new class of antithrombotic drugs.

Targets of Fibrinolytic Therapy

The blood fibrinolytic system essentially consists of proteolytic enzyme activation processes leading to the formation of the fibrin degrading enzyme plasmin, and thus is the counterpart of the blood coagulation system. The proenzyme plasminogen can be converted to the active enzyme plasmin by plasminogen activators (PAs), which both are counter balanced by inhibitors in order to protect the body from systemic proteolysis. The fibrinolytic and antifibrinolytic activity are determined primarily by the balance between PAs and plasminogen activator inhibitor (PAI), which are under control of hormones and growth factors ⁵³⁾.

One approach to the treatment of thrombosis is the pharmacological dissolution of the blood clot via the intravenous infusion of PAs. Two physiologic PAs, t-PA and u-PA have been identified^{54, 55)} and recombinant t-PA (rt-PA), recombinant single-chain u-PA (rscu-PA), two-chain u-PA (tcu-PA), streptokinase (SK) and anisoylated plasminogen streptokinase activator complex (APSAC) are either approved for clinical use or under clinical investigation⁵⁶⁾.

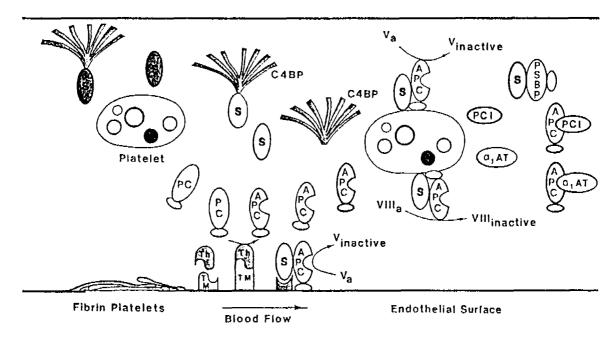


Fig. 9. Formation and function of activated Protein C

However, all available fibrinolytic agents still suffer significant limitations including large therapeutic doses, a limited fibrin specificity, a very short plasma half-life as a result of rapid hepatic clearance, and bleeding complication.

New developments towards further improved efficacy and fibrin specificity of fibrinolytic agents and, on the other hand, thrombolytic strategies to overcome resistance to clot lysis, to prevent reocclusion and to reduce bleeding tendency are being explored⁵⁷⁾. These include mutants and variants of rt-PA or rscu-PA, chimeric t-PA/u-PA plasminogen activators, and conjugates of plasminogen activators and either antifibrin or antiplatelet monoclonal antibodies. Several mutants of rt-PA obtained by deletion/substitution of functional domains of single amino acids have markedly reduced plasma clearance, but usually also reduced specific thrombolytic activity, resulting in unchanged or only marginally improved thrombolytic potency⁵⁷⁾. The t-PA of saliva from the vampire bat is highly homologous with human t-PA, but lacks K2 and the plasmin cleavage site for conversion to a two-chain form⁵⁸⁾. It was found to be a potent and fibrin-specific thrombolytic agent, the activity of which was stimulated 45,000-fold in the presence of fibrin compared to 200-fold for human t-PA. The structural characteristics of bat-PA might serve to improve specificity of thrombolytic therapy.

A low molecular derivative of human scu-PA, scu-PA-32k, which lacks the amino terminal 143 residues, was found to be less pronounced systemic fibrinogen breakdown with comparable clot lysis⁵⁹⁾. Several chimeric PAs consisting of various portions of t-PA and u-PA have been constructed and characterized. These chimeras have maintained the enzymatic properties of u-PA or of t-PA but their fibrin specificity was very similar to those of scu-PA⁶⁰. PAs conjugated with antifibrin or antiplatelet antibodies such as monoclonal antibodies against NH2-terminal of the Bbeta-chain of fibrin⁶¹⁾ or against glycoprotein IIbIIIa 62) are targeted to blood clot, since thrombus contains both fibrin-rich and platelet-rich material. These conjugates resulted in a significantly increased thrombolytic potency. The use of conjugates of PAs and specific monoclonal antibodies may allow a significant reduction of the total dose of PAs and systemic side effects.

Concluding Remarks

Thrombosis plays a major role in acute ischemic syndromes of cardiovascular and cerebrovascular disease. Several approaches can be used to treat these disease, which might be antiplatelet, antifibrin, anticoagulant or combi-

nation of these. However, the choice of therapy should be based on the type of thrombus and its pathogenesis. Several lines of research towards improvement of antithrombotic and fibrinolytic agents should be explored based on the molecular mechanism of target molecules.

References

- Moncada S, Gryglewski MW, Bunting S, and Vane JR Nature 263: 663-665, 1976
- Moncada S, Radomski, M.W., and Palmer, R.M. Biochem. Pharmacol. 37: 2495-2501, 1988
- Loskutoff, D.J. in Vascular Endothelium in Haemostasis and Thrombosis (Gimbrone, M.A., ed) pp. 120-141, Churchill Livingstone, Edinbergh, Scotland, 1986
- Packham, M.A. and Mustard, F.J. in Blood Platelet Function and Medicinal Chemistry (Lasslo, A. ed) pp. 61-128, New York. Amsterdame. Oxford, Elsevier Biomedical, 1984
- Tracy, P.B. and Mann, K.G. in CRC Platelet Responses and Metabolism 1 (Holmsen, H. ed) pp. 297-324. Boca Raton, CRC Press, 1986
- Mustard, J.F., Packham, M.A., and Kinlough-Rathbone, R.L. Circulation 81 (Suppl I): 124-127, 1990
- 7. Badimon, L., Chesebro, J.H., and Badimon, J.J. Circulation 86 (Suppl III): III 74-III 85, 1992
- 8. Ruggeri, Z.M. Circulation 86(Suppl III): III 26-III 29, 1992
- Plow, E.F., Marguerie, G., Ginsberg, M. Biochem. Pharmacol. 36: 4035-4040, 1987
- Badimon, L., Badimon, J.J., Lassila, R., Heras, M., Chesebro, J.H., and Fuster, V. Blood 78: 423-434, 1991
- 11. Patrono, C. Circulation 81 (Suppl I): I12-I15; 1990
- 12. Collen, D. and Lijnen, H.R. Blood 78: 3114-3124, 1991
- Ikeda, Y., Handa, M., Kawano, K., Kamata, T., Murata, M.,
 Araki, Y., Anbo, H., Kawai, Y., watanabe, K., Itagaki, I.,
 Sakai, K., Ruggeri, Z.M. J. Clin. Invest. 87: 1234-1240, 1991
- 14. Fujimura, Y., Holland, L.Z., Ruggeri, Z.M., and Zimmerman, T.S. Blood 70: 985-988, 1987
- Mohri, H., Fujimura, Y., Shima, M., Yoshioka, A., Houghten, R., Ruggeri, Z., and Zimmerman, T.S. J. Biol. Chem. 263: 17901-17904, 1988
- 16. Plow. E.F., D'souza, S., and Ginsberg, M.H. Sem. Thrombosis and Hemostasis 18: 324-332, 1992
- Kieffer, N., Phillips, D.R. Ann. Rev. Cell Biol. 6: 329-357, 1990
- Shattil, S.J., Hoxie, J.A., Cunningham, M., and Brass, L.F. J. Biol. Chem. 260: 11107-11114, 1985
- Coller, B.S., Peerschke, E.I., Scudder, L.E., and Sullivan, C.A.
 J. Clin. Invest. 72: 325-338, 1983
- Colle, B.S., Folts, J.D., Scudder, L.E., and Smith, S.R. Blood 68:783-786, 1986
- 21. Gatner, T.K. and Bennett, J.S. J. Biol. Chem. 260: 11891-11894, 1985

- Kloczewiak, M., Timmons, S., Lukas, T.J., and Hawiger, J. Biochemistry 23: 1767-1774, 1984
- Dennis, M., Henzel, W.J., Pitti, R.M., Lipari, M.T., Napier, M.A., Deisher, T.A., Bunting. S., and Lazarus, R.A. Proc. Natl. Acad. Sci. USA 87: 2471-2475, 1989
- Scarborough, R.M., Rose, J.W., Hsu, M.A., Phillips, D., Fried, V.A., Cambell, A.M., Nannizzi, L., and Charo, I.F. J. Biol. Chem. 266: 9359-9362, 1991
- Adler, M., Lazarus, R.A., Dennis, M.S., and Wagner, G. Science 253:445-448, 1991
- Samanen, J., Ali, F., Romoff, T., Calvo, R., Sorenson, E., Vasko, J., Storer, B., Berry, D., Bennett, D., Strohsacker, M., Powers, D., Stadel, J., and Nichols, A. J. Med. Chem. 34: 3114-3125, 1991
- 27. Barker, P.L., Bullens, S., Bunting, S., Burdick, D.J., Chan, S., Deisher, T., Eigenbrot, C., Gadek, T.R., Gantzos, R., Lipari, M.T., Muir, C.D., Napier, M.A., Pitti, R.M., Padua, A., Quan, C., Stanley, M., Struble, M., Tom, J,Y.K. and Burnier, J.P. J. Med. Chem. 35: 2040-2048, 1992
- Ambler, J., Birch, J., Maguire, E.D., and Wallis, R.B. Adv. Exp. Med. Biol. 192: 293-308, 1985
- Whittle, B.J.R. and Moncada, S. Brit. Med. Bulletin 39: 232-238, 1983
- Gresele, P., Deckmyn, H., Nenci, G.G., and Vermylen, J. Trends Pharmacol. Sci. 12: 158-163, 1991
- Humphrey, P.P.A., Hallet, P., Hornby, E.J., Wallis, C.J., Collington, E.W., and Lumley, P. Circulation 81 (Suppl I): 142-152, 1990
- 32. Gresele, P., Arnout, J., Dechmyn, H., Huybrechts, E., Pieters, G., and Vermylen, J. J. Clin. Invest. 87: 1435-1445, 1987
- 33. Shuman, M.A. Ann. NY. Acad. Sci. 485: 228-239, 1986
- Chesebro, J.H., Webster, M.W.I., Zoldhelyi, P., Roche, P.C., Badimon, L., and Badimon, J.J. Circulation 86 (Suppl III): III 100-III 110, 1992
- Vu, T-KH., Hung, D.T., Wheaton, V.I., and Coughlin, S.R. Cell 64: 1057-1068, 1991
- 36. Vu, T-KH., Wheaton, V.I., Hung, D.T., Chao, I., and Coughlin, S.R. Nature 353: 674-677, 1991
- 37. Beguin, S., Lindhout, T., and Hemker, H.C. Thromb. Haemostas. 60: 457-462, 1988
- 38. Weitz, J.I., Hudoba, M., Massel, D., Maraganore, J., and Hirsh, J. J. Clin. Invest. 86: 385-391, 1990
- 39. Rubens, F.D., Weitz, J.I., Brash, J.L., and Kinlough-Rathbone, R.L. Thromb. Haemostas. 69: 130-134, 1993
- 40. Markwardt, F. Method Enzymol. 19: 924, 1970
- Rydel, T.J., Ravichandran, K.G., Tulinsky, A., Bode, W., Huber, R., Roitsch, C., and Fenton, J.W. Science 249: 277-280, 1990
- Rydel, T., Tulinsky, A., Bode, W., and Huber, R. J. Mol. Biol. 221: 583-601, 1991
- Maraganore, J.M., Bourdon, P., Jablonski, J., Ramachandran, K.L., and Fenton, J.W II. Biochemistry 29: 7095-7101, 1990

- 44. Hauptmann, J. and Markwardt, F. Sem. Thrombosis and Hemostasis 18: 200-217, 1992
- Okamoto, S., Hihikata, A., Kikumoto, R., Tonomura, S., Hara, N., Ninomiya, K., Maruyama, A., Sugano, S., and Tamao, Y. Biochem. Biophys. Res. Commun. 101: 440-446, 1981
- 46. Sturzebecher, J., Markwardt, F., Voigt, B., Wagner, G., and Walsmann, P. Thromb. Res. 29: 635-642, 1983
- 47. Kettner, C. and Shaw, E. Thromb Res. 14: 969-973, 1979
- 48. Esmon, C.T. J. Biol. Chem. 264: 4743-4746, 1989
- Teitel, J.M. and Rosenberg R.D. J. Clin. Invest. 71: 1383-1391, 1983
- Vassallo, R.R.Jr., Kieber-Emmons, T., Cichowski, K., and Brass, L.F. J. Biol. Chem. 267: 6081-6085, 1992
- 51. Coughlin, S.R., Vu, T.-K.H., Hung, D., Wheaton, V.I Sem. Thrombosis and Hemostasis 18: 161-166, 1992
- Hung, D., Vu, T.-K.H., Wheaton, V.I Charo, I.F., Nelken,
 A., Esmon, N., Esmon, C.T., and Coughlin, S.R. J. Clin. Invest. 89: 444-450, 1992
- 53. Bevilacqua, M.P., Schleef, R.R., Gimbrone M.A., and Loskut-

- off, D.J. J. Clin. Invest. 78: 587-591, 1986
- Ny, T., Elgh, F., and Lund, B. Proc. Natl. Acad. Sci. USA 81: 5355-5359, 1984
- Holmes, W.E., Pennica, D., Blaber, M., Rey, M.W., Guenzler, W.A., Steffens, G.J., and Heyneker, H.L. Biotechnology 3: 923-929, 1985
- 56. Collen, D. ISI Atlas of Science. Pharmacology 3: 116, 1988
- 57. Lijnen, H.R. and Collen D. Thromb Haemost. 66: 88,1991
- Gardel, S.J., Duong, L.T., Dich, R.E., York, J.D., Hare, T. R., Register, R.B., Jacobs, J.W., Dixon, R.A.F., and Friedman, P.A. J. Biol. Chem. 264: 17947-17952, 1989
- Lijnen, H.R., Nelles, L., Holmes, W., and Collen, D. J. Biol. Chem. 263: 5594, 1988
- Collen, D., Stassen, J.M., Demarsin, E., Kieckens, L., Lijnen, H.R., and Nelles, L. J. Vasc. Med. Biol. 1: 234, 1989
- Bode, C., Matsueda, G., Hui, K.Y., Haber, E. Science 229: 765, 1985
- Bode, C., Meinhardt, G., Runge, M.S., Eberle, T., Schuler, G., ubler, W., and Haber, E. Throm. Haemost. 62: 483, 1989