

Larvicidal and Antifeeding Activities of Oriental Medicinal Plant Extracts against *Plutella xylostella* and *Spodoptera litura*

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Abstract : Methanol extracts from 45 species of oriental medicinal plants belonging to 17 families were subjected to a screening test for their larvicidal and antifeeding activities against two species of lepidopteran larvae, diamondback moth (*Plutella xylostella* L.) and tobacco cutworm (*Spodoptera litura* F.) by a leaf-dipping method. At a concentration of 8,000 ppm, methanol extract of *Corydalis turrschaninovii* roots only was found to have potent larvicidal activity against *S. litura*. At 5,000 ppm, strong antifeeding activity against both larvae was observed from the extracts of *Clerodendron trichotomum*, *C. trichotomum* var. *esculentum*, *Inura helenium*, *Arctium lappa*, *Artemisia messerschmidiana*, *Anthriscus sylvestris*, *Angelica dahurica*, and *C. turrschaninovii* (Received October 22, 1994; accepted November 11, 1994).

Introduction

Among 296 species of the insect and mite pests of vegetables in the Republic of Korea, the most important of which are diamondback moth, *Plutella xylostella* (L.) and tobacco cutworm, *Spodoptera litura* (F.).^{1,2} If not managed properly, these species cause serious yield losses when larvae excessively feed on the developing vegetables. Control is primarily dependent upon continued or repeated applications of insecticides. Although they have effectively controlled these insect pests, their extensive use for the past decades has disrupted control of these insect populations by natural enemies and has led to outbreaks of these insect pests, and the development of widespread resistance to various types of insecticides.³⁻⁵ Decreased efficacy and increasing concern over adverse effects of the earlier types

of insecticide have brought about the need for the development of new types of selective alternatives or of methods of crop protection without, or with reduced use of organic insecticides.

Plants may be an alternative to currently used insect control agents, because they virtually are rich source of bioactive organic chemicals. Recently, much concern has been focused on the distribution, nature, and practical use of chemical substances having the antifeeding activity for insects in plants. Antifeedants from plants have no or little harmful effects on non-target organisms and environment,⁶⁻⁸ suggesting that they could be developed into products suitable for integrated pest management (IPM) in crops. Although oriental medicinal plants are rich source of organic chemicals,⁹ little work has been done to manage insect populations or their damage by using them.

Key words : Larvicidal activity, antifeeding activity, *Plutella xylostella*, *Spodoptera litura*, plant extract
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In the laboratory study described herein, we assessed the larvicidal and antifeeding activities against both *P. xylostella* and *S. litura* of methanol extract of 45 species of oriental medicinal plants belonging to 17 families. Among the plants tested, the antifeeding components of *Clerodendron trichotomum* var. *esculentum*, the only *Clerodendron* species in southern coastal region of the Republic of Korea, has been investigated and will be reported elsewhere.

Materials and Methods

Insects

Laboratory strains of two insect species were used in this study: diamondback moth (*Plutella xylostella* L.) and tobacco cutworm (*Spodoptera litura* F.). They have been reared for several years without exposure to any insecticide in our laboratory by the same procedures described previously.¹⁰⁻¹²⁾

Plants and sample preparation

The 45 oriental medicinal plant species belonging to 17 families are anecdotally selected and listed in Table 1. Fully developed leaves from *Clerodendron trichotomum*, *C. trichotomum* var. *esculentum*, *Symplocos chinensis* and *S. coreana* were collected during July to August 1992 at the Forestry Research Institute, Seoul, Republic of Korea. The other plant species were purchased from an oriental medicinal plant market in Seoul. These plant materials were dried in a blower at 60°C for 3 days, finely powdered using a blender, extracted twice with methanol at room temperature, and filtered (Toyo filter paper No. 2). The combined filtrate was concentrated *in vacuo* at 35°C, using rotary vacuum evaporator. The yield of each extraction is shown in Table 1.

Bioassay

The most important factor in primary screening for bioactive substances may be the starting concentration. In a preliminary test, a concentration of 5,000~8,000 ppm of a plant extract did not cause any problem such as solubility and detection of mi-

Table 1. Oriental medicinal plant species tested

Plant species	Family name	Part	Yield (%) ¹
<i>Morus bombycis</i>	Moraceae	leaf	15
<i>Broussonetia kazinoki</i>	Moraceae	leaf	19
<i>Acer barbinerve</i>	Aceraceae	leaf	23
<i>Chionanthus retusa</i>	Oleaceae	leaf	23
<i>Abeliophyllum distichum</i>	Oleaceae	leaf	25
<i>Fraxinus rhynchophylla</i>	Oleaceae	root	35
<i>Clerodendron trichotomum</i>	Verbenaceae	leaf	20
<i>Clerodendron trichotomum</i> var. <i>esculentum</i>	Verbenaceae	leaf seed	18 4
<i>Symplocos chinensis</i>	Symplocaceae	leaf	23
<i>Symplocos coreana</i>	Symplocaceae	leaf	19
<i>Eucommia ulmoides</i>	Eucommiaceae	leaf	11
<i>Inura helenium</i>	Compositae	root	11
<i>Arctium lappa</i>	Compositae	root	15
<i>Artemisia</i> <i>messerschmidtiana</i>	Compositae	stem	10
<i>Atractylodes japonica</i>	Compositae	root	8
<i>Carthamus tinctorius</i>	Compositae	flower	19
<i>Leonurus sibiricus</i>	Labiatae	leaf	20
<i>Schizonepeta tenuifolia</i> var. <i>japonica</i>	Labiatae	leaf	6
<i>Acanthopanax sessiliflorum</i>	Araliaceae	root	23
<i>Aralia continentalis</i>	Araliaceae	root	12
<i>Panax ginseng</i>	Araliaceae	root	10
<i>Kalopanax pictum</i>	Araliaceae	cortex	10
<i>Aconitum carmichaeli</i>	Ranunculaceae	root	16
<i>Clematis florida</i>	Ranunculaceae	root	19
<i>Paeonia japonica</i>	Ranunculaceae	root	10
<i>Aconitum pseudo-laeve</i> var. <i>erectum</i>	Ranunculaceae	root	13
<i>Anthriscus sylvestris</i>	Umbelliferae	root	21
<i>Angelica dahurica</i>	Umbelliferae	root	12
<i>Bupleurum falcatum</i>	Umbelliferae	root	15
<i>Ledebouriella seseloides</i>	Umbelliferae	root	17
<i>Angelica gigas</i>	Umbelliferae	root	12
<i>Torilis japonica</i>	Umbelliferae	seed	3
<i>Citrus aurantium</i>	Rutaceae	fruit	15
<i>Poncirus trifoliata</i>	Rutaceae	fruit	32
<i>Phellodendron amurense</i>	Rutaceae	cortex	15
<i>Gleditsia sinensis</i>	Leguminosae	fruit	10
<i>Pueraria thunbergiana</i>	Leguminosae	root	21
<i>Astragalus membranaceus</i>	Leguminosae	root	10
<i>Corydalis turrschaninovii</i>	Fumariaceae	root	1
<i>Chaenomeles sinensis</i>	Rosaceae	fruit	37
<i>Crataegus maximowiczii</i>	Rosaceae	fruit	39
<i>Prunus persica</i>	Rosaceae	seed	8
<i>Cyperus rotundus</i>	Cyperaceae	fruit	5
<i>Scirpus fluviatilis</i>	Cyperaceae	root	9
<i>Belamacanda chinensis</i>	Iridaceae	root	20

¹(Weight/100 g of dry weight of test material)×100

nor active components.

Leaf-dipping method was used for larvicidal and antifeeding activities of test materials against *P. xylostella* and *S. litura*. The plant extracts were tested at a concentration of 8,000 ppm for larvicidal activity and 5,000 ppm for antifeeding activity. Test samples suspended in distilled water with Triton X-100 added at the rate of 0.1 ml/liter were used. Leaves of chinese cabbage (*Brassica oleracea* var. *capitata* L.) grown in green house were collected and disks (2 cm diameter) were punctured. Three leaf disks were dipped in test solution for 30 sec. After evaporation in a hood for 2 hr, 10 second instars of each *S. litura* and *P. xylostella* were placed onto the trea-

ted and control leaf disks in Petri dishes. All treated materials were held in a room at $25 \pm 1^\circ\text{C}$, 50~60% RH, and a photoperiod of 16:8 (light/dark) hr. All treatments were triplicated.

Larvicidal activity was evaluated after 48 hr, and classified as follow: the strong activity + + +, mortality >80%; moderate + +, mortality 80~61%; weak +, mortality 60~40%; and no response -, mortality <40%. Antifeeding activity was calculated according to the method of Isman *et al.*,¹³⁾ evaluated after 24 hr, and classified as follows: the strong antifeeding activity + +, >80%; moderate +, 80~50%; and no activity -, <50%.

Table 2. Antifeeding activity of methanol extracts against larvae of *Plutella xylostella* (DBM) and *Spodoptera litura* (TCW) by a leaf-dipping method¹

Plant species	Antifeeding activity ²		Plant species	Antifeeding activity ²	
	DBM	TCW		DBM	TCW
<i>M. bombycis</i>	-	-	<i>A. carmichaeli</i>	+	+
<i>B. kazinoki</i>	-	-	<i>C. florida</i>	+	-
<i>A. barbinerve</i>	+	+	<i>P. japonica</i>	-	++
<i>C. retusa</i>	-	-	<i>A. pseudo-laeva</i>	++	-
<i>A. distichum</i>	+	-	var. <i>erectum</i>		
<i>F. rhynchophylla</i>	-	-	<i>A. sylvestris</i>	++	++
<i>C. trichotomum</i>	++	++	<i>A. dahurica</i>	++	++
<i>C. trichotomum</i>	++ ³	++	<i>B. falcatum</i>	++	-
var. <i>esculentum</i>	++ ⁴	++	<i>L. seseloides</i>	-	-
<i>S. chinensis</i>	+	+	<i>A. gigas</i>	++	-
<i>S. coreana</i>	-	+	<i>T. japonica</i>	++	-
<i>E. ulmoides</i>	+	+	<i>C. aurantium</i>	++	-
<i>I. helenium</i>	++	++	<i>P. trifoliata</i>	++	-
<i>A. lappa</i>	++	++	<i>P. amurense</i>	-	-
<i>A. messerschmidiana</i>	++	++	<i>G. sinensis</i>	++	+
<i>A. japonica</i>	++	+	<i>P. thunbergiana</i>	++	+
<i>C. tinctorius</i>	+	+	<i>A. membranaceus</i>	-	-
<i>L. sibiricus</i>	++	+	<i>C. turrschaninovii</i>	++	++
<i>S. tenuifolia</i>	++	-	<i>C. sinensis</i>	-	-
var. <i>japonica</i>			<i>C. maximowiczii</i>	-	-
<i>A. sessiliflorus</i>	++	+	<i>P. persica</i>	-	-
<i>A. continentalis</i>	-	-	<i>C. rotundus</i>	-	-
<i>P. ginseng</i>	++	+	<i>S. fluviatilis</i>	-	-
<i>K. pictus</i>	-	-	<i>B. chinensis</i>	++	-

¹5,000 ppm treatment.

² + +, >80%; +, 80~50%; -, <50%.

³Leaves.

⁴Seeds.

Results

Larvicidal activity

Among the plant species tested, the methanol extract of *C. turrschaninovii* (Family Fumariaceae) roots only showed potent larvicidal activity (+++) against *S. litura*. However, the yield of methanol extraction of this plant species was very low (1%). The other 44 plant extracts were nontoxic to the larvae of both *P. xylostella* and *S. litura*.

Antifeeding activity

The activity of test samples investigated against larvae of *P. xylostella* and *S. litura* was both insect and plant species dependent (Table 2). Of the 45 species of plants, the following eight samples exhibited a strong antifeeding activity against both lepidopteran larvae: *C. trichotomum* and *C. trichotomum* var. *esculentum* (Family Verbenaceae), *I. helenium*, *A. lappa* and *A. messerschmidtiana* (Family Compositae), *A. sylvestris* and *A. dahurica* (Umbelliferae), and *C. turrschaninovii* (Family Fumariaceae).

Discussion

In the laboratory study with larvae of *P. xylostella* and *S. litura*, antifeeding activity was both plant and insect species dependent. The plants belonging to the families Verbenaceae, Compositae, and Umbelliferae showed a strong antifeeding activity against both insect species, whereas the methanol extract of roots of *C. turrschaninovii* belonging to the family Fumariaceae showed both insecticidal and antifeeding activities against *S. litura* larvae. Jacobson¹⁴ pointed out that the most promising botanicals as sources of novel plant-based insecticides for use at the present time and in the future are species of the families Meliaceae, Rutaceae, Asteraceae, Annonaceae, Labiatae, and Canellaceae.

Certain plant-derived extracts and phytochemicals can not only be useful as insecticides, but also reduce plant damage below the economic injury level. They are being considered as potential alterna-

tives for organic insecticides.^{7,15} Derivatives of neem (*Azadirachta indica* A. Juss) belonging to the family Meliaceae are found to have a variety of biological activities against nearly 200 species of insects without any adverse effects on non-target organisms.¹⁶ It is well recognized that the use of antifeedants is of considerable potential value in crop protection, based upon the fact that they have selectivity towards the natural enemies of pests, and may be applied to the plant in the same way as other agricultural chemicals.^{6,17} In addition, most *P. xylostella* larvae were killed in immature stages,¹⁸ and the natural enemies were an important factor in larval mortality¹⁹ so that antifeedants may play a large role to increase mortality of survived larvae without any adverse effect on natural enemies.

Extracts from neem seed/neem seed kernels have shown great potential for IPM in controlling *P. xylostella* in cabbage and related vegetables owing to their selectivity, although their application in cabbage can have a change of plant color and reduction of headsize.⁸ Plant-derived extracts are found to be only effective against insects for a relatively short period of time. However, extracts of neem seeds gave good protection of crops against *P. xylostella* for 6 days,⁸ and against swarms of *Schistocerca gregaria* for up to 2 weeks if it was not washed off by rain.¹⁹ However, little information is available for antifeeding activity of oriental medicinal plants, although these plants have long been considered to have natural properties.⁹ In our study, the methanol extracts of *C. trichotomum*, *C. trichotomum* var. *esculentum*, *I. helenium*, *A. lappa*, *A. messerschmidtiana*, *A. sylvestris*, *A. dahurica* and *C. turrschaninovii* showed a strong antifeeding activity against larvae of both *P. xylostella* and *S. litura*.

Based upon our results and these earlier findings, antifeedants from oriental medicinal plants might be useful product for developing new types of biorational management agents for controlling both *P. xylostella* and *S. litura* populations, although their effects on natural enemies, vegetable qualities, or environment has not been fully investigated.

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References

1. Korean Society of Plant Protection (1986) In 'A List of Plant Diseases, Insect Pests, and Weeds in Korea,' The Korean Society of Plant Protection, Suwon, Republic of Korea
2. Agricultural Sciences Institute (1990) In 'Ecology and Control of Insect Pests of Vegetables,' Rural Development Administration, Suwon, Republic of Korea
3. Kim, G. H., Y. S. Suh, J. H. Lee and K. Y. Cho (1990) Development of fenvalerate resistance in the diamondback moth, *Plutella xylostella* Linne (Lepidoptera: Yponomeutidae). *Korean J. Appl. Entomol.*, 29, 194-200
4. Song, S. S. (1992) Resistance of diamondback moth (*Plutella xylostella* L.) against the pyrethroids. *Korean J. Appl. Entomol.*, 31, 338-344
5. Lee, S. C., Y. S. Cho and D. I. Kim (1993) Comparative study of toxicological methods and field resistance to insecticides in diamondback moth (Lepidoptera: Plutellidae). *Korean J. Appl. Entomol.*, 32, 323-329
6. Chapman, R. F. (1974) The chemical inhibition of feeding by phytophagous insects: a review. *Bull. Ent. Res.*, 64, 339-363
7. Arnason, J. T., B. J. R. Philogene and P. Morand (1989) In 'Insecticides of Plant Origin.' ACS Symposium 387, Amer. Chem. Soc., Washington, D. C.
8. Schmutterer, H. (1992) In 'Diamondback Moth and Other Crucifer Pests,' pp. 325-332, N. S. Talekar (ed.), Proceedings of the second international workshop, 10-14 December 1990, Tainan, Taiwan
9. Namba, T. (1986) In 'Colored Illustrations of Wakan-Yaku (the Crude Drugs in Japan, China and the Neighbouring Countries),' Hoikusha Publishing Co., Osaka, Japan
10. Cho, K. Y., J. W. Ahn and Y. J. Ahn (1988) In 'The Establishment of Screening System for Developing New Agrochemicals,' The Ministry of Science, Seoul, Republic of Korea
11. Ahn, Y. J. and K. Y. Cho (1992) Establishment of bioassay system for developing new insecticides I. Effects of organic solvents on the toxicity against insects, phytotoxicity and solubility of new compounds. *Korean J. Appl. Entomol.*, 31, 182-189
12. Ahn, Y. J., K. H. Kim, N. J. Park and K. Y. Cho (1992) Establishment of bioassay system for developing new insecticides II. Differences in susceptibilities of the insect species to insecticides according to different application methods. *Korean J. Appl. Entomol.*, 31, 452-460
13. Isman, M., Koul, B. O., Luczynski, A. and J. Kaminiski (1990) Insecticidal and antifeedant bioactivities of neem oils and their relationship to azadirachtin content. *J. Agric. Food Chem.*, 38, 1406-1411
14. Jacobson, M. (1989) In 'Insecticides of Plant Origin,' pp. 1-10, J. T. Arnason, B. J. R. Philogene and P. Morand (eds), ACS Symposium 387, Amer. Chem. Soc., Washington, D.C.
15. Jacobson, M. and D. G. Crosby (1971) In 'Naturally Occurring Insecticides,' Marcel Decker, New York
16. Saxena, R. C. (1989) In 'Insecticides of Plant Origin,' pp. 110-135, J. T. Arnason, B. J. R. Philogene and P. Morand (eds), ACS Symposium 387, Amer. Chem. Soc., Washington, D.C.
17. Munakata, K. (1970) In 'Control of Insect Behaviour by Natural Products,' pp. 179-187, D. L. Wood, R. M. Silverstein and M. Nakajima (eds), Academic Press, New York
18. Sivapragasam, A., Y. Ito and T. Saito (1988) Population fluctuations of the diamondback moth, *Plutella xylostella* (L.) on cabbages in *Bacillus thuringiensis* sprayed and non-sprayed plots and factors affecting within-generation survival of immatures. *Res. Popul. Ecol.*, 30, 329-342.
19. Wakisaka, S., R. Tsukuda and F. Nakasuji (1992) In Diamondback Moth and Other Crucifer Pests, pp. 15-26, N. S. Talekar (ed), Proceedings of the second international workshop, 10-14 December 1990, Tainan, Taiwan
20. Pradhan, S. and M. G. Jotwani (1968) Neem as an insect deterrent. *Chem. Age India*, 19, 756-760.

배추즙나방과 담배거세미나방 유충에 대한 한방식물체 추출물의 살충 및 섭식저해활성
권형욱 · 안용준* · 권정현 · 이상길¹ · 변병호¹(서울대학교 농생물학과, ¹임업연구원 산림곤충과)

초록 : 17과 45종 한방 식물체 메탄올 추출물의 배추즙나방과 담배거세미나방 유충에 대한 살충활성 및 섭식저해활성을 잎침지법으로 조사하였다. 살충활성은 8,000 ppm에서 조사하였는바, 현호색 추출물만이 담배거세미나방 유충에 대해 강한 살충활성을 보였다. 섭식저해활성은 5,000 ppm에서 조사한 결과, 누리장나무, 거문누리장나무, 목향, 우방자, 인진, 전호, 백지 및 현호색 추출물이 강한 섭식저해활성을 나타내었다.