

## Dermatophyte and Cyclohexamide-Resistant Fungi Isolated from Patients with Tinea Capitis and from Air in Hospitals in Minia, Egypt

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**ABSTRACT:** Out of 210 tinea capitis cases studies, 16 were negative when examined with KOH, among the remaining 194 cases, 123 were males (63.4%) and 71 were females (36.6%) and the age of incidence ranged between 7 and 15 years. *Microsporum* was the main causal agent being identified in 82 cases (42.3%) and was represented by 5 species among which *M. canis* was common in Egypt (55 cases, 28.4%). *Trichophyton* constituted 32% and was represented by 8 species among which *T. violaceum* was the most common (24.2% of total cases). *Candida* were isolated from 3.6% of total cases. The 47 species and twenty-five genera from nondermatophyte-cyclohexamide resistant fungi were recovered from the diseased skin tissue. *Penicillium* and *Aspergillus* were the most abundant followed by *Scopulariopsis*, *Alternaria*, *Thermoascus*, *Chrysosporium* and *Cladosporium*. Studies of the air-borne fungi in-door the hospital wards revealed the occurrence of 57 species belonging to 28 genera, among which *Aspergillus flavus*, *Penicillium chrysogenum*, *P. corylophilum*, *A. niger*, *Tritirachium rosum* and *Alternaria alternata* were the most common. Results of the out-door experiments were basically similar to those of the in-door experiments.

**KEYWORDS:** Dermatophytes, cyclohexamide, tinea capitis, fungi.

Tinea capitis is a common and widely distributed disease throughout the world. It was reported from; Iran (Chadgani *et al.*, 1987); Argentina (Tello & Chappuis, 1984); West of Scotland (Gentles & Scott, 1981); India (Khosa *et al.*, 1981); Australia (McAlleer, 1981); and from Kuwait (Karaoui *et al.*, 1979). In Egypt, tinea capitis in Cairo (Verhagen, 1979); In Ismailia (Abdallah *et al.*, 1986) and in Assiut (Abdel-Hafez *et al.*, (1980).

Studies on fungal air spore in hospitals could provide information on dissimination and spread of fungal spores causing mycosis. Such studies are lacking in Egypt, particularly at hospitals in upper Egypt. Generally, studies on keratinolytic fungi of the air are rare (Alteras and Lehrer, 1977 and Della Franca and Caretta, 1984). In Egypt, Maghazy (1989) studied the keratinophilic fungi of the air and the floor dust in primary schools in Assiut.

The aim of this study was to study the dermatophytic causal agents and the non-dermatophytic combined infections of tinea capitis Minia Governorate. The fungal flora of the air in the hospitals where patients with mycosis are located is also reported.

### Materials and Methods

Two hundred and ten patients diagnosed clinically as tinea capitis in three hospitals at Minia city (Skin and Venereal clinic at Minia University hospital, Dermatology and Venereology disease clinic and Dermatology clinic at Minia chest hospital) were studied. Specimens for mycological study were obtained from the lesions after wiping the surfaces with 70% ethyl alcohol to remove surface adhering organisms and dust from the skin. These were then packed in steril filter paper envelopes, sealed and labelled. Care was taken to avoid contamination with tick scales which were

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mostly composed of debris and dirt but no fungal elements. Direct microscopic examination of KOH preparations was carried out. Fungal isolations were performed on Sabouroud's dextrose agar medium (Moss and McQuown, 1969) supplemented with 20 I.U./ml sodium salt of penicillin, 40 µg/ml dihydrostreptomycin and 0.05% cyclohexamide (actidione). Before adding to the agar, the first two antibiotics were separately dissolved in sterile distilled water while the third was dissolved in methanol. Plates were incubated at 28°C for 3-4 weeks and the developing colonies were examined and identified.

Determination of air-borne fungi was performed by exposing Petri plates containing Sabouraud's dextrose agar supplemented with the antibiotics mentioned previously to the air once every month for 2 hours between 9:00 AM - 1:00 PM during a year period. Exposure took place in the in-door and out-door in the hospital wards in Dermatology and Venereology disease clinic, Minia. Plates were incubated at 30°C for 2 weeks, colonies were counted and fungi were isolated and identified.

## Results and Discussion

**Dermatophytes from diseased skin tissues:** Out of 210 cases, 16 were negative for what ? when examined with KOH. Among the remaining 194 positive cases, 123 were males (63.4%) and 71 were females (36.6%), indicating more tinea capitis infected males than females. In this respect it could be mentioned that, Guinani & NjoKu-OBL (1986) also noticed more males infected with tinea capitis than females and similar results were also recorded in Eastren Nigera by Wolf *et al.*, (1986) and by Abdel-Hafez *et al.*, (1980) in Assiut. On the contrary, Abdallah *et al.*, (1986) in Ismailia city observed more females infected with tinea capitis than males.

The age incidence of tinea capitis ranged between one and 15 years (Table 1). Two age groups (1-5 years and 6-10 years) contributed 78 cases each (40.2%) and 38 cases belonged to the age group 11-15 years (19.6%). Comparing our data with those reported by other investigators, it could be mentioned that; in Jordan, Shtayeh & Arda

**Table 1.** Age distribution of tinea capitis (194 cases).

Age group by years	No. of cases	%
1-5	78	40.21
6-10	78	40.21
11-15	38	19.59
Total	194	100.00

(1985) recorded higher frequency of tinea capitis in children up to 14 years, and in Nigera, Ajao & Akintude (1985) recorded more tinea capitis in 1-13 years-old children. This means that infection by tinea capitis is generally more frequent among children up to 13-14 years old.

*Microsporum*, *Trichophyton* and *Candida* were the main dermatophytes associated with tinea capitis as indicated in Table (2) and Fig. (1). *Microsporum* was isolated more frequently and was represented by 5 species among which *M. canis* was the most dominant and was isolated from 55 cases. This fungus was reported to be the most common causal organism of tinea capitis in Jordan, (Shtayeh & Arda, 1985); in London, (Ridley, 1979); in Cairo, Egypt, (Verhagen, 1974) and in Assiut (Abdel-Hafez *et al.*, 1980). On the other hand, *M. audouinii* (which isolated less frequently) was reported to be the main causal organism of tinea capitis in Nigeria, (Ajao & Akintude, 1985) and in USA (Sinski & Kelley, 1987). Other *Microsporum* species were very rare and were isolated from 1-4 patients only.

*Trichophyton* was the second most prevalent causal agent of tinea capitis (recorded in 32% of total cases) with eight species identified among which, *T. violaceum* was the most common. This fungus was also the main etiological agent of tinea capitis in Sengal and Mouritani Valley (Marill *et al.*, 1976) and in Assiut, Egypt (Abdel-Hafez *et al.*, 1980). Other *Trichophyton* sp were infrequent and occurred only in 1-4 cases.

*Candida* ranked third among the dermatophytes isolated. It was isolated from 7 males only accounting for 3.6% of the total cases and was represented by 3 species: *C. albicans*, *C. tropicalis* and *Candida* sp. *Candida* species were recovered from

**Table 2.** The isolation and occurrence of fungal dermatophytes isolated from human patients infected with tinea capitis.

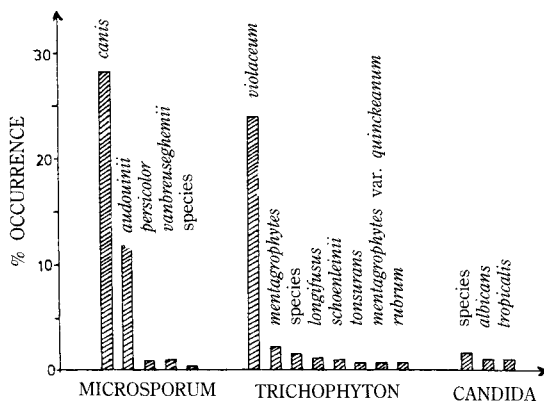
Genera & Species	No. of cases of isolation			% of occurrence			O. R.
	O+♀	O	♀	O+♀	O	♀	
<i>Microsporum</i>	82	52	30	42.3	26.8	15.5	H
<i>M. canis</i>	55	32	23	28.4	16.5	11.9	L
<i>M. audouinii</i>	23	16	7	11.9	8.3	3.6	R
<i>M. persicolor</i>	2	2	0	1.0	1.0	0.0	R
<i>M. vanbreuseghemii</i>	1	1	0	0.5	0.5	0.0	R
<i>Microsporum</i> sp.	1	1	0	0.5	0.5	0.0	R
<i>Trichophyton</i>	62	40	22	32.0	20.6	11.3	L
<i>T. violaceum</i>	47	31	16	24.2	16.0	8.2	L
<i>T. mentagrophytes</i>	4	3	1	2.1	1.6	0.5	R
<i>T. longifusus</i>	3	0	3	1.5	0.0	1.5	R
<i>T. schoenleinii</i>	2	2	0	1.0	1.0	0.0	R
<i>T. tonsurans</i>	1	0	1	0.5	0.0	0.5	R
<i>T. mentagrophytes</i> var. <i>quinckeanum</i>	1	1	0	0.5	0.5	0.0	R
<i>T. rubrum</i>	1	1	0	0.5	0.5	0.0	R
<i>Trichophyton</i> sp.	3	2	1	1.6	1.0	0.5	R
<i>Candida</i>	7	7	0	3.6	3.6	0.0	R
<i>C. albicans</i>	2	2	0	1.0	1.0	0.0	R
<i>C. tropicalis</i>	2	2	0	1.0	1.0	0.0	R
<i>Candida</i> sp.	3	3	0	1.5	1.5	0.0	R

O. R. = Occurrence remarks.

H = High occurrence: more than 65 cases (out of 194 cases).

L = Low occurrence: between 30-65 cases.

R = Rare occurrence: less than 30 cases.

**Fig. 1.** The occurrence of dermatophytes isolated from Tinea capitis.

tinea capitis lesions from USA (Cox, 1986), Argentina (Silvia *et al.*, 1986) and from Nigeria (Ogbonna *et al.*, 1986).

**Non-dermatophytes from diseased skin tissues:** Twenty-five genera and forty-seven species from other keratinophilic and cyclohexamide resistant fungi were recovered as associated fungi with tinea capitis infection as shown in Table (3). *Penicillium* and *Aspergillus* were the most predominant saprophytic genera isolated. These were recovered from 48 and 24 cases comprising 24.7% and 12.4% of the total cases studied and were represented by 8 and 6 species respectively. *Sco-*

**Table 3.** The isolation and occurrence of cyclohexamide-resistant fungi isolated from human patients infected with tinea capitis.

Genera & Species	No. of cases of isolation	% of occurrence	O.R.	Genera & Species	No. of cases of isolation	% of occurrence	O.R.
<i>Penicillium</i>	48	24.7	L	<i>Cladosporium</i>	5	2.6	R
<i>P. chrysogenum</i>	20	10.3	P	<i>C. herbarum</i>	2	1	R
<i>P. corylophilum</i>	11	5.7	R	<i>C. cladosporioides</i>	2	1	R
<i>P. lilacenum</i>	5	2.6	R	<i>C. sphaerospermum</i>	1	0.6	R
<i>P. citrinum</i>	4	2.0	R	<i>Acremonium</i>	4	2	R
<i>P. funiculosum</i> sp.	3	1.5	R	<i>A. strictum</i>	1	0.5	R
<i>P. brevi-compactum</i>	2	1	R	<i>A. kiliense</i>	1	0.5	R
<i>P. purpurogenum</i>	2	1	R	<i>A. roseum</i>	1	0.5	R
<i>P. surantio-vireus</i>	1	0.5	R	<i>Acremonium</i> sp.	1	0.5	R
<i>Aspergillus</i>	24	12.4	R	<i>Mucor</i>	2	1	R
<i>A. ochraceus</i>	8	4	R	<i>M. racemosus</i>	1	0.5	R
<i>A. flavus</i>	6	3	R	<i>M. pusillus</i>	1	0.5	R
<i>A. niger</i>	6	3	R	<i>Myrothecium verrucaria</i>	1	0.5	R
<i>A. oryzae</i>	2	1	R	<i>Chaetomium indicum</i>	1	0.5	R
<i>A. candidus</i>	1	0.5	R	<i>Sporodromium</i> sp.	1	0.5	R
<i>A. sydowii</i>	1	0.5	R	<i>Trichoderma harizianum</i>	1	0.5	R
<i>Scopulariopsis</i>	8	4	R	<i>Myceliophthora anamorph</i>	1	0.5	R
<i>S. brevicaulis</i>	6	3	R	of <i>Arthroderma tuberculatum</i>			
<i>S. prumbitii</i>	2	1	R	<i>Gymnoascus reesii</i>	1	0.5	R
<i>Alternaria alternata</i>	6	3	R	<i>Ulocladium atrum</i>	1	0.5	R
<i>Thermoascus aurantiacus</i>	5	2.6	R	<i>Trichothecium roseum</i>	1	0.5	R
<i>Chrysosporium</i>	5	2.6	R	<i>Geotrichum candidum</i>	1	0.5	R
<i>C. keratinophilum</i>	3	1.5	R	<i>Stachybotrys atra</i>	1	0.5	R
<i>C. tropicum</i>	2	1	R	<i>Phialophora spinifera</i>	1	0.5	R
<i>Malbranchea</i>	3	1.5	R	<i>Phialophora</i> sp.	1	0.5	R
<i>M. chrysosporioides</i>	2	1	R	<i>Scolecobasidium humicola</i>	1	0.5	R
<i>Malbranchea</i> sp.	1	0.5	R	<i>Hormiactis candida</i>	1	0.5	R
<i>Microascus trigonosporus</i>	2	1	R	<i>Paecilomyces lilacinus</i>	1	0.5	R

O. R. = Occurrence remarks.

H = High occurrence: more than 65 cases (out of 194 cases).

L = Low occurrence: between 30-65 cases.

R = Rare occurrence: less than 30 cases.

*pulariopsis* was isolated from 8 cases; *Alternaria*, 6 cases; *Thermoascus*, *Chrysosporium* and *Cladosporium* (5 cases for each). The remaining species were recovered from less than 5 cases. These non-dermatophytic cyclohexamide resistant fungi

are not responsible for causing the disease and did not originate as contaminants during preparation and isolation of specimens, but represent saprophytes or secondary invaders of the damaged skin substrate.

**Table 4.** In-door air-borne fungi in the hospital wards during one year period.

Genera & Species	Total count	Percentage of total count	Number of months of isolation	O.R.
Total count	654			
<i>Aspergillus</i>	152	23.24	11	R
<i>A. flavus</i>	86	13.15	6	M
<i>A. niger</i>	54	8.26	5	M
<i>A. ochraceus</i>	4	0.61	3	M
<i>A. sydowii</i>	6	0.92	2	L
<i>A. nidulans</i>	1	0.15	1	L
<i>A. melleus</i>	1	0.15	1	L
<i>Penicillium</i>	223	34.10	10	H
<i>P. chrysogenum</i>	67	10.24	8	H
<i>P. corylophilum</i>	56	8.56	7	H
<i>P. citrinum</i>	44	6.73	4	M
<i>P. luteum</i>	7	1.07	2	L
<i>P. funiculosum</i>	4	0.61	2	L
<i>P. thomii</i>	8	1.22	1	L
<i>P. miczynskii</i>	16	2.45	1	L
<i>P. cyclopium</i>	1	0.15	1	L
<i>P. rubrum</i>	1	0.15	1	L
<i>P. capsulatum</i>	1	0.15	1	L
<i>Acremonium</i>	33	5.05	10	H
<i>A. strictum</i>	15	2.29	6	M
<i>Acremonium</i> spp.	7	1.07	4	M
<i>A. kiliense</i>	4	0.61	3	M
<i>A. rutilum</i>	7	1.07	3	M
<i>Alternaria</i>	33	5.05	8	H
<i>A. alternata</i>	32	4.89	8	H
<i>A. tenuissima</i>	1	0.15	1	L
<i>Chrysosporium</i>	20	3.05	8	H
<i>C. keratinophilum</i>	9	1.38	4	M
<i>C. queenslandicum</i>	3	0.45	3	M
<i>Chrysosporium</i>	5	0.76	3	M
<i>C. xerophilum</i>	2	0.31	1	L
<i>C. tropicum</i>	2	0.31	1	L
<i>Cladosporium</i>	33	5.05	6	M
<i>C. cladosporioides</i>	27	4.13	5	M
<i>C. macrocarpum</i>	5	0.76	2	L
<i>C. herbarum</i>	1	0.15	1	L
<i>C. sphaerospermum</i>	2	0.31	1	L

Table 4. Continued

Genera & Species	Total count	Percentage of total count	Number of months of isolation	O.R.
<i>Candida</i>	8	1.22	4	M
<i>C. albicans</i>	6	0.92	3	M
<i>C. tropicalis</i>	1	0.15	1	L
<i>Candida</i> sp.	1	0.15	1	L
<i>Malbranchea</i>	8	1.22	4	M
<i>Malbranchea</i>	7	1.07	3	M
<i>M. chrysosporioidea</i>	1	0.15	1	L
<i>Tritirachium roseum</i>	40	6.12	2	L
<i>Scopulariopsis</i>	3	0.45	2	L
<i>S. brevicaulis</i>	2	0.31	1	L
<i>S. brumptii</i>	2	0.31	1	L
<i>Mucor racemosus</i>	3	0.46	2	L
<i>Geotrichum candidum</i>	2	0.31	2	L
<i>Botryotrichum piluliferum</i>	2	0.31	2	L
<i>Gliocladium roseum</i>	1	0.15	2	L
<i>Thermoascus aurantiacus</i>	2	0.31	2	L
<i>Monodictys levis</i>	3	0.45	1	L
<i>Trichurus spiralis</i>	1	0.15	1	L
<i>Emericellopsis</i> spp.	1	0.15	1	L
<i>Phialophora fastigiata</i>	1	0.15	1	L
<i>Auxarthron zuffianum</i>	1	0.15	1	L
<i>Chaetomiun jodphurense</i>	1	0.15	1	L
<i>Paecilomyces inflatus</i>	1	0.15	1	L
<i>Cunninghamella echinulata</i>	1	0.15	1	L
<i>Stachybotrys atra</i>	1	0.15	1	L
<i>Mortierella</i> sp.	1	0.15	1	L
<i>Arachniotus citrinus</i>	1	0.15	1	L
<i>Oidiodendron griseum</i>	1	0.15	1	L
<i>Beauveria</i> sp.	1	0.15	1	L
Sterile hyphae	1	0.15	1	L
Yeast cells	75	11.47	10	H

O. R. = Occurrence remarks.

H = High occurrence: more than 6 months (out of 12).

M = Moderate occurrence: between 3-6 months.

L = Low occurrence: less than 3 months.

**Air-borne fungi from hospital wards:** Fifty-seven species which belong to 28 genera were collected from in-door experiments (Table. 4). Five genera

were of high seasonal occurrence and these were, *Aspergillus*, *Penicillium*, *Acremonium*, *Alternaria* and *Chrysosporium*.

*Aspergillus* was the most frequent genus and was one of the basic components of air-borne fungi. It regularly appeared in eleven months and was represented by 6 species. *Penicillium* and *Acromonium* occurred in ten months each. *Penicillium* count irregularly fluctuated and was represented by eleven species, of which *P. chrysogenum* and *P. corylophilum* were the most common. *Alternaria* and *Chrysosporium* were encountered in 8 months. Among the group of chrysosporia isolated from the air in this investigation were; *C. keratinophilum*, *C. queenslandicum*, *C. xerophilum*, *C. tropicum* were *Chrysosporium* sp. *Cladosporium* emerged in 6 months and was represented by 4 species. *Candida* was isolated in 4 months and 3 species were identified namely; *C. tropicalis*, *C. albicans* and *Candida* sp. *Malbranchea* was reported in 4 months and two species were recovered which were *Malbranchea chrysosporioides* and *Malbranchea* sp. Twenty species were of low occurrence (1-2 months) as shown in Table, (4). Results of the out-door experiments were basically similar to those of the in-door experiments. Most of these species are common in the open-air in Egypt as reported by Moubasher & Moustafa (1974) in Assiut; by Mazen & Shaban (1983a) in Minia; by Moubasher *et al.*, (1981a) in Quena, and by Moubasher *et al.*, (1985b) in Sohag.

Della Franca and Caretta, (1984) studied keratinophilic fungi in the air at Pavia, Italy and recovered *A. flavus*, *Alternaria alternata*, *Cladosporium* (6 species), *Penicillium* (3 species) and many others which are resistant to antibiotics. Maghazy (1989) in Egypt studied the air keratinophilic fungi in primary schools in Assiut and isolated *Aspergillus* (6 spp.), *Penicillium* (6 spp), *Cladosporium* (2 spp), *Chrysosporium* (2 spp) and *Alternaria* (1 sp.). It is worth mentioning that through most of the non-dermatophytes isolated from the diseased skin tissues were isolated from the air no real dermatophytic fungus was caught from the air of the Dermatology hospital wards during this study indicating that the pathogenic dermatophytes on the contrary to the saprophytes are not transmitted by air but mainly by some other means.

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