Ecological Distribution of Endomycorrhizal Fungi in Pogil-do in Tadohae-haesang National Park

SEO, HYUN CHANG¹, CHANG JIN KIM, SHIN DUK KIM, AND ICK DONG YOO*

Genetic Engineering Research Institute, Korea Institute of Science and Technology, P.O. Box 17, Taedok Science Town, Taejon 305-606, Korea

1Department of Food and Nutrition, Shingu Junior College, Sungnam 462-743, Kyongqi-do, Korea

Received 6 December 1991 / Accepted 13 February 1992

The ecological distribution of endomycorrhizas in evergreen woody species native to the evergreen forest ecosystem of Tadohae-haesang National Park in southern Korea in February, 1989 was studied. The abundance and diversity of vesicular-arbuscular (VA) mycorrhizal fungi were also determined. The spore densities ranged from 14 to 326 per 100 g of soil. Most of the spores of mycorrhizal fungi collected from 25 soil samples belonged to the genera Glomus and Gigaspora. The frequency and number of spores in Camellia japonica varied with location. Spores belonging to the genus Gigaspora were not found in Camellia japonica in Yesongri evergreen forests adjacent to the sea. Glomus sp. was the major constituent of the spore assemblage at this site. The most abundant species in Camellia japonica in the Yesongri evergreen forests in Pogildo was Glomus borealis. In the soil of a mountain at Buwhangri, in the central location of the island at an elevation of 250 m, Gigaspora sp. was present and Glomus sp. was a major constituent of the spore assemblage. In the urban area of Haenam spore densities were much higher than in the Pogildo area. The most abundant species in Camellia japonica in the urban area of Haenam was Gigaspora sp..

VA mycorrhizal associations in the rhizosphere are thought to be almost ubiquitous, having extremely wide host ranges, and are present in most existing species of vascular plants, including agricultural crops grown throughout the world (6). In addition to these wide host ranges there are many reports on improved plant growth due to the presence of mycorrhizal association (2, 5, 14, 16). These symbiotic associations with host plants have also been reported to promote plant health (7, 15, 19). Mycorrhizal plants are more resistant to plant stress induced by drought (15), high soil temperature (19) and root pathogens (7). These published reports of plant growth and health, and the real concern about pollution of the environment induced by chemical fertilizers and pesticides, have suggested the possible use of VA mycorrhizal fungi as "biofertilizer" (1, 13, 17).

*Corresponding author

Key words: VA mycorrhizal fungi, Glomus borealis, Camellia japonica, evergreen forests, native ecosystem, Gigaspora However, for the effective use of VA mycorrhizal fungi as biofertilizer, we need to understand their ecology and select suitable fungi. Only about 120 species of VA mycorrhizal fungi have been identified and there are few studies dealing with the ecology of VA mycorrhizal fungi in native ecosystems (3, 4, 18).

In agroecosystems of Korea a lot of pesticides and chemical fertilizers have been used. This type of agriculture may have killed many useful soil microorganisms, including VA mycorrhizal fungi, and may have damaged the ecosystems. In this respect, as a first step for the selection of VA mycorrhizal fungi for agricultural use, we studied the ecological distribution of VA mycorrhizal fungi from an unpolluted ecosystem. In this environment diverse VA mycorrhizal fungal species might be present (20) and possible candidates for VA mycorrhizal fungal biofertilizers might to be abundant. Reports on the taxonomic distribution of ecto- and endomycorrhizal fungi from Korean soil were surveyed by Lee and Koo (11).

The purpose of this study was to examine the ecological distribution of endomycorrhizal fungi in evergreen woody species native to the forest ecosystem of Tadohae-haesang National Park in southern Korea in February, 1989.

MATERIALS AND METHODS

Study Areas

Perennial plants were surveyed for colonization at 2 sites in the evergreen forests of Pogil-do and 1 site in the Haenam-up area at Chollanamdo (Fig. 1). A total of 25 soil samples were obtained from 19 woody species. Sampling site selection was based on general soil characteristics and floristic composition. The evergreen forests of Yesongri in Pogil-do were designated and preserved as a Natural Monument of Korea (No. 40). Camellia japonica was the major woody species at these sites. Site #1 was adjacent to a seashore of small pebbles and the soil was clay rich with abundant organic matter. Site #2 was in the Pogil-do forest ecosystem on the side of a mountain in Buwhangri, near the center of the island, at an elevation of 250 m. The sea is departed about 2.5 km from this site. In the urban area of Haenam, at site #3, samples were also collected for analysis of endomycorrhizal fungi. All the sites studied were characterized as temperate zone evergreen forests

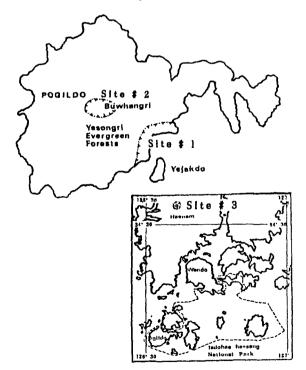


Fig. 1. The Pogil-do area and its location in the Tadohae-haesang National Park in Chollanamdo, Korea, and the locations of sampling sites.

where evergreen woody species could survive perennia-

Isolation and Identification of Endomycorrhizal Fungi

Evergreen woody species were selected for sampling of mycorrhizal fungi. Soil and root samples were collected from the top 10 to 20 cm soil. A representative sample of the entire root system was obtained from five or six different portions of the root system, according to the plant species. These portions were combined rather than obtaining one large sample from a single portion. Samples of 2 to 3 Kg were collected of each plant species. To measure the density of spores subsamples of 200 ml of soil and root fragments were wet sieved and soil sievings were then centrifuged in 1 M sucrose solution to separate detritus from the spores (9). Spores were examined under a dissecting microscope (20~40x) and then grouped by color, shape and size. A compound microscope (100~1,000x) was used for further examination and identification of the spores. Spores were preliminarily identified and then compared for structural details against information in the original species description. Identification of endomycorrhizal fungi was based on the "Manual for the identification of VA mycorrhizal fungi" by INVAM and the "Compilation of the Endogonaceae" by Shannon M. Berch (3, 8, 18). Sudan grass (Sorghum bicolor L.) was grown for 5 months in pot culture in a sterile sand-vermiculite medium (1:1, v/v)with spores from each plant species as VA mycorrhizal inoculum.

Observation of Infection Characteristics of VA Mycorrhiza

Root samples of fine terminal feeder roots attached to lower order roots, free of root systems of adjacent plants, were preserved in fixation solution (formalin- acetic acid-alcohol solution) before staining. Staining of the root samples was based on the method of Komanick (9).

RESULTS AND DISCUSSION

Distribution of Endomycorrhizal Fungi

Distribution of endomycorrhizal fungi was estimated as the number of spores per 100 g of soil collected in the rhizosphere of the evergreen forests. Most of the spores of mycorrhizal fungi collected from the 25 soil samples belonged to the genera Glomus and Gigaspora.

Among the sites surveyed, Sites #1 and #2 (Yesongri evergreen forests) have been well preserved as a Natural Monument. Glomus sp. was a major constituent of the spore assemblage at Site #1 (Fig. 2). The site was adjacent to the sea side and thought to be high in soil salinity. As VA mycorrhizal fungi have been shown to adapt to edaphic factors (10), salinity-resistant species or geno68 SEO ET AL. J. Microbiol. Biotechnol.

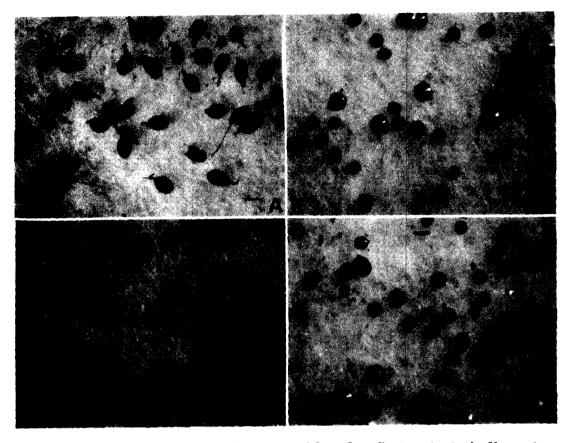


Fig. 2. Diversities of spores of endomycorrhizal fungi isolated from Camellia japonica in the Yesongri evergreen forests of Pogil-do (Site #1). Scale line=100 μm .

types may also exist in our isolates from coast areas. Spores belonging to the genus Gigaspora were not found at Site #1 and the wall layer of chramydospores found in Camellia japonica was generally thick. Site #2, a forest ecosystem in Pogil-do, was on a mountainside at an elevation of 250 m. The spore densities in evergreen woody species at the two sites in Pogil-do and one site in Haenam are compared in Table 1. The spore densities ranged from 14 to 326 per 100 g of soil at the 3 sites studied. Many environmental factors acting on the host plant may affect these differences in the production of VA mycorrhizal spores. However, the influence of host plants was thought to be minor among the factors, considering the spore densities in evergreen woody species common to the three sites (Table 1). The spore densities in the evergreen woody species Camellia japonica, common to the three sites were compared in Table 1. The spore densities were 58 in Site #1, 128 in Site #2, and 224 in Site #3. These differences are attributed to a difference in soil fertility, based upon the soil characteristics examined. The fact that the soil of Site #1 was clay rich with abundant organic matter, and the difference of spore density in Cinnamomum

Table 1. Density of spores of endomycorrhizal fungi isolated in the Pogil-do and Haenam areas in February, 1989.

Sampling site	Plant species	Spores/100 g soil
Site #1		
	Camellia japonica	58
Yesong-ri	llex integra	38
Evergreen	Cinnamomum japonicum	26
Forests	Kalopanax pictus	14
in	Cryptomeria japonica	60
Pogil-do	Neolitsea sericea	38
	Cinnamomum japonicum	16
Site #2		
	Chamaecyparis obtusa	28
Buwhang-ri	Camellia japonica	128
in	Machilus thunbergii	38
Pogil-do	Cinnamomum japonicum	48
	Elaeagnus umbellata	46
Site #3		
Haenam	Camellia japonica	224
	Ilex crenata	326

japonicum at Site #1 and Site #2 (Table 1) also support this suggestion. Changes in spore densities in agroecosystems have also been associated with changes in soil fertility (12). Schenck's result regarding the comparison of the density of spores between agroecosystems and native ecosystems (20) also supports our result that the differences are the result of soil fertility. Therefore, mycorrhizal fungal infestations of evergreen woody plants are thought to be dependent on the extent of organic matter in the soil. Thus, it can be said that Camellia japonica at Site #3 was more dependent on endomy-corrhizal fungi than at Site #1.

Comparison of the Types of Endomycorrhizal Fungi in Camellia japonica at Three Different Sites

The types of spores in the evergreen woody species Camellia japonica common to the three sites were compared. The differences in spores, grouped by color, shape and size, are shown in Fig. 2 (Site #1) and Fig. 3 (Sites #2 and #3). The types of spores found in the two sites in Pogil-do were different from each other. As

B

Fig. 3. Comparison of spores of endomycorrhizal fungi isolated from Camellia japonica in Buwhang-ri (Site #2, A) and the Haenam area (Site #3, B). Scale line=100 µm.

shown in Fig. 2, diverse mycorrhizal fungal species were found in Camellia japonica in the Yesongri evergreen forests at Site #1. Glomus sp. was a major spore type and Gigaspora sp. was not found at this site. The abundant species found in the site are shown in Fig. 2-(A) and spores of this type were not found in any site studied. In Site #2, Gigaspora sp. was found, but Glomus sp. was still the major spore type, while in Site #3 in the urban area of Haenam, Gigasporas sp. was the major spore type (Fig. 3-(B)). It is well known that the hyphae of VA mycorrhizal fungi serve as extensions of the root systems and are both physiologically and geometrically more effective organs of absorption than the roots themselves (2). Therefore, Camellia japonica in the urban area of Haenam studied is thought to be more dependent on VA mycorrhizal fungi than in the Pogil-do area.

Identification of Endomycorrhizal Fungi

Dominant spore types of endomycorrhizal fungi from Camellia japonica forests at Site #1 are shown in Fig. 2-(A). The spores have not been reported in Korea as yet so further analysis was performed. The spores were

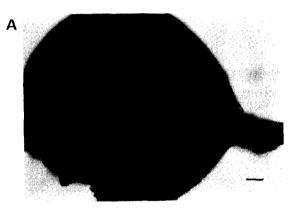




Fig. 4. A spore of endomycorrhizal fungi, Glomus borealis, predominantly abundant in Camellia japonica in the Yesongri evergreen forests of Pogil-do. Scale line=10 μm.

70 SEO ET AL. J. Microbiol. Biotechnol.

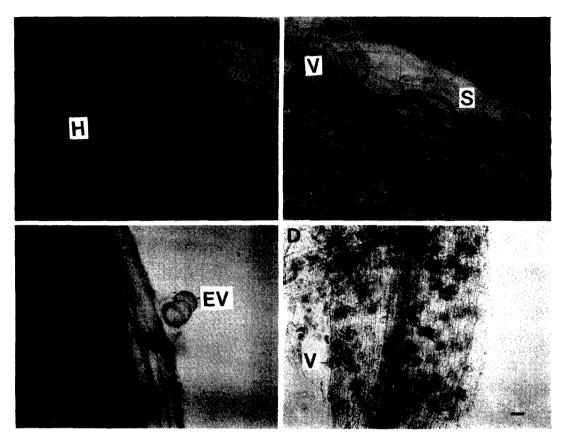


Fig. 5. Infection characteristics of hyphae of Glomus borealis in Camellia japonica.

(A, B) Intercellular hypha (H) and vesicles (V) found in Camellia japonica, (C) Intercellular hypha and extracellular vesicles (EV) found in Camellia japonica, (D) Abundant vesicles found intercellularly in Camellia japonica in the Yesongri evergreen forests of Pogil-do. Scale line = 10 μm.

reddish-brown about 100×140 µm in size, broadly and rather symmetrically ellipsoid, borne on a rather slender hyphae and frequently subtended by a septum (Fig. 4-(A)). The spores have a single layer spore wall. Spore contents were finely granular and more or less fibrous (Fig. 4-(B)). They were compared with published data and identified as Glomus borealis (3, 8, 18). The infection characteristics shown in Fig. 5 showed that the type of mycorrhiza was VA mycorrhiza. Fossil records of VA mycorrhiza-like structures have been dated at about 370 million years ago (6). Therefore, the widespread distribution of VA mycorrhizal fungi may have required millions of years for the dispersal and adaptation of mycorrhizal fungi. Glomus borealis is known only from Quebec, Canada and pot cultured mycorrhizal associations are not known (3). In this experiment sudan grass (Sorghum bicolor L) was grown for 5 months in pot culture in a sterile sand-vermiculite medium (1:1, v/v) with spores of Glomus borealis as VA mycorrhizal inoculum. The result showed that sudan grass was a good pot cultured

host plant for the mycorrhizal fungi. The importance of searching for endomycorrhizal fungi lies in the fact that most of the endomycorrhiza are a common feature of many woody and herbaceous plants, including agronomic crops. Further studies will be focused on the agricultural applications of these mycorrhizal fungi.

REFERENCES

- Abbott, L.K. and A.D. Robson. 1982. The role of vesicular and arbuscular mycorrhizal fungi in agriculture and the selection of fungi for inoculation. *Aust. J. Agric. Res.* 33: 389-408.
- Abbott, L.K. and A.D. Robson. 1984. The effect of VA mycorrhizae on plant growth, p.113-130. In C.L. Powell (ed.), VA Mycorrhiza, CRC Press, Inc., Boca Raton, Florida.
- Berch, S.M. 1988. Compilation of the Endogonaceae, p. 81-178. 1st ed. Mycologue Publications, Ontario.
- Bethlenfalvay, G.J., G.J. Dakessian, and R.S. Pakovsky.
 1984. Mycorrhizae in a southern California desert: ecological implications. Can. J. Bot. 62: 519-524.

131.

- 5. Capaccio, L.C.M. and J.A. Callow. 1982. The enzymes of polyphosphate metabolism in vesicular-arbuscular mucorrhizas. New Phytol. 91: 81-91.
- 6. Daniels, B.A. 1984. Ecology of VA mycorrhizal fungi, p. 35-55. In C.L. Powell (ed.), VA Mycorrhiza, CRC Press. Inc., Boca Raton, Florida.
- 7. Dehne, H.W. 1982. Interaction between vesicular-arbuscular mycorrhizal fungi and plant pathogens. Phytopathology **72**: 1115-1119.
- 8. Hall, I.R. and B.J. Fish. 1979. A key to the endogonaceae. Trans. Br. Mycol. Soc. 73: 261-270.
- 9. Kormanik, P.P. and H.D. Skipper. 1982. Quantification of VA mycorrhizae in plant roots, p. 37-45. In N.C. Schenck (ed.), Methods and Principles of Mycorrhizal Research, The American Phytopathological Society, St. Paul, Minnesota.
- 10. Lambert, D.H., D.E. Baker, and H. Cole, Jr. 1979. The role of mycorrhizae in the interactions of phosphorus with zinc, copper, and other elements. Soil Sci. Soc. Am. J. **43**: 976-980.
- 11. Lee, K.J. and C.D. Koo. 1983. Taxonomic distribution ecto- and endomycorrhizae among woody species in Korea. J. Korean For. Soc. 59: 37-45.
- 12. McGraw, A.C., and J.W. Hendrix. 1984. Host and soil fumigation effects on spore population densities of species of endogonaceus mycorrhizal fungi. Mycologia 76: 122-

- 13. Menge, J.A. 1983. Utilization of vesicular-arbuscular my-
- corrhizal fungi in agriculture. Can. J. Bot. 61: 1015-1024. 14. Mosse, B. 1973. Advances in the study of vesicular-arbus-
- cular mycorrhiza. Annu. Rev. Phytopathol. 11: 171-196. 15. Nelsen, C.E. and G.R. Safir. 1982. Increased drought tolerance of mycorrhizal onion plants caused by improved phosphorus nutrition. Planta 154: 407-413.
- 16. Powell, C.L. 1984. Field inoculation with VA mycorrhizal fungi, p. 205-222. In C.L. Powell (ed.), VA Mucorrhiza. CRC Press, Inc., Boca Raton, Florida.
- 17. Rhodes, L.H. 1980. The use of mycorrhizae in crop production systems. Outlook Agric. 10: 275-281.
- 18. Schenck, N.C. and Y. Perez. 1987. Manual for the Identification of VA Mycorrhizal Fungi, p. 96-195. 1st ed. IN-VAM, Gainesville, Florida.
- 19. Schenck, N.C. and V.N. Schroder. 1974. Temperature response of Endogone mycorrhiza on soybean roots. Mycologia 66: 600-605.
- 20. Schenck, N.C., J.O. Siequeira, and E. Oliveira. 1989. Changes in the incidence of VA mycorrhizal fungi with changes in ecosystems, p. 125-129. In V. Vancura (ed.), Interrelationships Between Microorganisms and Plant in Soil, Elsevier, New York.