

Electron Microscopic Study on Detoxication Effect of Red Ginseng Extract on Mouse Liver Injury induced by Methyl Mercury

Chung, Hee Won

메틸수은으로 損傷된 생쥐 肝臟에 대한 紅蔘 抽出物の
解毒效果에 관한 電子顯微鏡的 研究

鄭 喜 媛

(Received April 9, 1987)

抄 錄

생쥐 肝細胞의 微細構造에 미치는 메틸수은의 毒性的 影響과 이에 대한 紅蔘 抽出物の 解毒效果를 電子顯微鏡的으로 연구하였다.

對照群의 肝細胞에 비하여 메틸수은만을 投與한 群에서는 核의 일부가 崩壞되고, mitochondria의 膨大, cristae의 收縮 내지 損失 및 다수의 대형 腔胞의 出現이 일어나고 glycogen의 量이 감소되었다.

메틸수은과 紅蔘 抽出物を 併行 投與한 群에서는 메틸수은만을 投與한 群에서보다 mitochondria의 膨大 程度가 적고 腔胞의 크기와 수가 감소되었으며 正常 細胞와 비슷한 상태로 나타났다.

Introduction

Methyl mercury absorbed generally through respiratory organ, gastrointestinal tract, and skin (Tunncliffe and Wood, 1973; Vallee and Ulmer, 1972) is bound with hemoglobin in the red blood cells and circulates in this form for months (Arena, 1986). It passes easily into the cell membranes due to its lipid solubility (Fang and Fallin, 1976; Lau and Sarkar, 1979). It is distributed mainly liver, kidney, and central nervous system, then has toxic influences on

these organs (Donaldson and Gubler, 1978). The accumulation of a large amount of mercury after methyl mercury administration also produces a variety of morphological alterations in these organs (Valek, 1965; Ware *et al.*, 1974).

Meanwhile, ginseng is absorbed rapidly in the gastrointestinal tract and distributed to many organs, such as the liver, kidney, brain, stomach, and lung (Han, 1974). Brekhman and Darcymov (1969) reported adaptogenic theory that ginseng increases resistance of a living body under various undesirable conditions non-

specifically. Ginseng promotes synthesis of protein in the liver and is helpful to rapid hepatic regeneration when the liver is injured by various causes (Bae, 1978). Furthermore, ginseng prevents the accumulation of toxic materials by speeding up the metabolic rate of toxic materials (Hong *et al.*, 1979). However, the detoxifying mechanism of ginseng has not been clearly defined.

In this study, the detoxifying effect of red ginseng extract against toxicity of methyl mercury to fine structures of mouse liver was investigated by means of electron microscopy.

Materials and Methods

Experimental animal

Female ICR mice, 20~30 g, were used and divided into three groups (control, group I and group II). Ten animals per group were housed in cage at 20~25°C and permitted free access to food (Samyang Laboratory Chow) and water.

Treatment

Control group was administered orally with distilled water only for 23 days. Group I was treated with methyl mercury chloride of 15 mg/kg, b.w./day by oral administration with animal feeding needle for 23 days. Group II was administered with red ginseng extract of 200 mg/kg, b.w./day for 7 days before mercury treatment, and then treated with methyl mercury chloride of 15 mg/kg, b.w./day and red ginseng extract of 200 mg/kg, b.w./day as post-treatment. The red ginseng extract containing crude saponin of 150 mg/g was manufactured by the Korean office of monopoly.

Electron microscopic observation

The excised mouse liver was diced into one cubes, immediately fixed in 2% paraformaldehyde-2.5% glutaraldehyde mixture buffered with 0.1M phosphate (pH 7.4) for 2 hours,

postfixed in 1% osmium tetroxide in the same buffer for 1 hour (Palade, 1952), dehydrated in graded ethanol and acetone, and embedded in Epon 812 (Luft, 1961). Ultra-thin sections, made by LKB III Ultramicrotome, were stained with uranyl acetate and lead citrate (Reynolds, 1963), and examined under JEM-100B type electron microscope.

Results and Discussion

Hepatocyte of control group showed a large nucleus containing a prominent nucleolus. The chromatin was scattered throughout the nucleoplasm and the periphery of nucleus showed typical rough endoplasmic reticula (Figs. 1, 2). There were numerous microvilli on the base surface of the cell, which projected into the space of Disse and promote rapid metabolic material exchange by increase of surface area (Fig. 2). Numerous mitochondria contained normal cristae (Fig. 3). The glycogen granules were distributed widely in the cytoplasm. And there were lysosomes and numerous microvilli near bile canaliculus. The desmosomes, symmetrical plaques between two adjacent cells, were observed near bile canaliculus (Figs. 1, 2). Microbody showed a spherical body with dense core (Fig. 3), which is responsible for detoxifying action such as decomposition of hydrogen peroxide toxic to cellular life (Lentz, 1971).

Hepatocyte of group I showed appearance of many large vacuoles and partial rupture of nucleus (Fig. 4). Mitochondria were swelled to the whole volume increase considerably. Matrix with a low electron density and cristae shrunk near inner membrane of mitochondria were observed (Figs. 4, 5). Such mitochondrial change also was observed in case of the necrosised hepatic cells of rat treated with HgCl₂ (Gritzka and Trump, 1968; Trump *et al.*, 1965) or CCl₄,

(Ashworth *et al.*, 1963; Leduc and Wilson, 1958). Considering that normal mitochondria enable hepatic cell to use the energy for synthesis of metabolic material (Lentz, 1971), the degenerations of mitochondria would suggest the abnormal hepatic function. Glycogen granules were seldom and this phenomenon might be caused by the toxicity of methyl mercury. That is, the enzyme activity for synthesis and decomposition of glycogen in carbohydrate metabolism was inhibited by the methyl mercury toxicity. Trump *et al.* (1965) reported that toxic material administered to mouse induces the decrease of glycogen content during the hepatocyte necrosis.

In hepatocyte of group II, swelling level of mitochondria was diminished and the vacuoles were decreased in size and number compared with those of group I. From this event, it would be assumed that the red ginseng extract enabled damaged liver cells to be recovered around the control. This is supported by the report that the ginseng is helpful to rapid hepatic regeneration of injured liver due to toxic materials (Bae, 1978). Yu (1977) also reported that the morphological alterations of mouse liver treated with ginseng and carbon tetrachloride are milder than liver treated with carbon tetrachloride only and there is no cytoplasmic ballooning in the hepatic cells.

Summary

Detoxication effect of red ginseng extract against toxicity of methyl mercury on ultrastructure of mouse liver was studied by electron microscopy.

The hepatocyte of methyl mercury treatment group showed partial rupture of nucleus, mitochondrial swelling, decrease of glycogen content, and appearance of a great number of large va-

cuoles.

While, the hepatocyte of methyl mercury-red ginseng extract treatment group showed slight mitochondrial swelling and decrease of vacuoles in size and number than those of methyl mercury treatment group.

References

- Arena, J.M. 1986. Poisoning: Toxicology, symptoms, treatments. 5th ed. Thomas C.C. Publisher, Illinois, U.S.A., pp.206~209.
- Ashworth, C.T., F.J. Luibel, E. Sanders and N. Arnald. 1963. Hepatic cell degeneration. Arch. Pathol. 75, 212~215.
- Bae, H.W. 1978. Korean ginseng. 2nd ed. Korea Ginseng Research Institute Publication. pp. 148~151.
- Brekhman, I.I. and I.V. Dardymov. 1969. New substance of plant origin which increase non-specific resistance. Ann. Rev. Pharmacol. 9, 419~430.
- Donaldson, M.L. and C.J. Gubler. 1978. Biochemical effects of mercury poisoning in rats. Am. J. Clin. Nutr. 31, 859~864.
- Fang, S.C. and E. Fallin. 1976. The binding of various mercurial compounds to serum proteins. Bull. Environ. Contam. Toxicol. 15, 110~117.
- Gritzka, T.L. and B.F. Trump. 1968. Renal tubular lesions caused by mercuric chloride. Am. J. Pathol. 52, 1225~1278.
- Han, B.H. 1974. Metabolism of triterpene glycosides of Korean ginseng (I). Korean J. Ginseng Sci. 2, 17.
- Hong, S.A., J.K. Lim and C.W. Park. 1979. Pharmacological action of ginseng. J. Ginseng Sci. 3, 66~93.
- Lau, S. and B. Sarkar. 1979. Inorganic mercury (II)-binding components in normal human blood serum. J. Toxicol. Environ. Health

- 5, 907~916.
- Lentz, T.L. 1971. Cell fine structure: An atlas of drawings of whole cell structure. Saunders Co., Philadelphia, pp. 3~10.
- Leduc, E.M. and J.W. Wilson, 1958. Injury to liver cells in carbon tetrachloride poisoning. Am. J. Pathol. 65, 147.
- Luft, J.H. 1961. Improvements in epoxy resin embedding method. J. Biophys. Biochem. Cytol. 9, 409~414.
- Palade, G.E. 1952. A study of fixation for electron microscopy. J. Exp. Med. 95, 285.
- Reynolds, E.S. 1963. The use of lead citrate at high pH as an electron opaque stain in electron microscopy. J. Cell Biol. 17, 208~212.
- Trump, B.F., P.I. Glodbatt and R.E. Stowell. 1965. Studies of necrosis *in vitro* of mouse hepatic parenchymal cells. Lab. Invest. 14, 1946~1986.
- Tunncliff, G. and J.D. Wood. 1973. The inhibition of mouse brain neurotransmitter enzymes by mercury compounds and a comparison with the effects of hyperbaric oxygen. Comp. Gen. Pharmacol. 4, 101~105.
- Valek, A. 1965. Acute renal insufficiency in toxication with mercury compounds. I. Aetiology, clinical picture, renal function. Acta Med. Scand. 177, 63~67.
- Vallee, B.T. and D.D. Ulmer. 1972. Biochemical effects of mercury, cadmium and lead. Ann. Rev. Biochem. 41, 91~123.
- Ware, R.A., L.W. Chang and P.M. Burkholder. 1974. Ultrastructural evidence for foetal liver injury induced by in utero exposure to small doses of methylmercury. Nature 251, 236~237.
- Yu, C.H. 1977. Effect of ginseng extract on normal and damaged hepatic cells. T.M. Chungang Univ. p. 29.

Figure Legends

- Fig. 1.** Electron micrograph of hepatocyte from control mouse with normal nucleus(N), nucleolus(Nu). Glycogen granules(Gl), mitochondria(M) with cristae(Cr), rough endoplasmic reticulum(RER) and smooth endoplasmic reticulum(SER) are distributed throughout the cytoplasm. Lysosomes(Ly) are seen near the bile canaliculus(BC) and microvilli(Mv) are toward the bile canaliculus. Arrowhead indicates desmosome(D). $\times 9,000$.
- Fig. 2.** Electron micrograph of hepatocyte from control mouse. Microvilli(Mv) are projected into the space of Disse(*). $\times 9,500$.
- Fig. 3.** Electron micrograph of hepatocyte from control mouse. Note the mitochondria(M) with well developed cristae(Cr). Microbody(Mb) is typical structure. $\times 12,000$.
- Fig. 4.** Electron micrograph of hepatocyte from the mouse treated with methyl mercury. Note the swollen mitochondria(M) with matrix rarefaction and loss of cristae(Cr). The large vacuoles(V) appeared. Nucleus(N) is destructed partially(arrow) and glycogen granules(Gl) are decreased. $\times 12,000$.
- Fig. 5.** Electron micrograph of hepatocyte from the mouse treated with methyl mercury. There are numerous large vacuoles(V). $\times 12,000$.
- Fig. 6.** Electron micrograph of hepatocyte from the mouse treated with methyl mercury and red ginseng extract. Nucleus(N), nucleolus(Nu) and mitochondria(M) are similar to those of the control. Vacuoles(V) are decreased in size and number as compared with the methyl mercury treatment group. RER, Rough endoplasmic reticulum. $\times 9,000$.





