Triterpenes from the Seeds of Phytolacca sp.

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Abstract—Direct comparison of the triterpenoids isolated from the seeds of *Phytolacca* sp. and synthetic compounds confirmed that the natural products were acetylaleuritolic acid and 3-acetylmyricadiol rather than epiacetylaleuritolic acid, acetyloleanolic acid and phytolaccanol.

Keywords—*Phytolacca esculenta* • *P. americana* • *P. acinosa* • Phytolaccaceae • triterpenoids • acetylaleuritolic acid • 3-acetylmyricadiol • epiacetylaleuritolic acid

In the course of phytochemical work on Phytolacca sp. $^{1\sim 11)}$, taraxeranes were encountered from the seeds.

The least polar, major compound (1) obtained from the chloroform soluble portion of methanol extract from the seeds of *Phytolacca americana* was a pentacyclic triterpene acetate, $C_{32}H_{50}O_4$, mp $303\sim4^\circ$. Its IR spectrum showed acetoxyl peaks at 1735 and 1243 cm⁻¹, carboxyl peak at 1692 cm⁻¹, and trisubstituted double bond peaks

at 825 and 810 cm⁻¹, and its NMR spectrum showed signals at $\delta 2.02$ and 5.49 ppm. The acetoxyl group was shown to be secondary equatorial judging from a signal for an axially oriented methine proton at $\delta 4.43$ (1H, dd, J=7 and 9Hz)¹²⁾. Seven tertiary methyl singlets between $\delta 0.85$ and 0.92 and a well-defined signal at $\delta 5.49$ (1H, dd, J=4 and 8Hz) for the olefinic proton of the trisubstituted double bond suggested the taraxer-14-ene skeleton¹²⁾. This suggestion

- (2) R = Ac
- (3) R = H

- AcO
 - (1) M^+ ; $m_{/Z}$ 498 (R=COOH)
 - (4) M+; m/z 484 (R=CH2OH)

- ii m/z 234(R=COOH)
 - m/z 220(R=CH2OH)

iii ^m/z 189

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was supported by strong peaks at m/z 344(i), 329(i-CH₃), 234(ii) and 189(iii) in its mass spectrum¹³⁾. The double bond in taraxer-14-ene derivatives was known to be isomerized readily under mild acidic conditions to give corresponding olean-12-enes¹⁴⁾. Thus, as expected, treatment of this compound with HCl-acetic acid afforded acetyloleanolic acid (2) which confirmed by direct comparison with an authentic sample.

On the basis of the above findings, this compound was identified as acetylaleuritolic acid. Its identity was further confirmed by direct comparison with an authentic sample of acetylaleuritolic acid¹⁵⁾ which was kindly supplied by Dr. Khastgir.

Although we were unable to detect acetyloleanolic acid (2) and/or oleanolic acid (3) in any extracts of our sample of P. species, Le Quesne and his coworker¹⁶ previously reported the isolation of acetyloleanolic acid from the seeds of P. americana. Direct comparison of their sample

with ours resulted in the identity of these two terpenoids.

Therefore, Le Quesne's sample was revised to be acetylaleuritolic acid¹⁷⁾.

The seeds of P. esculenta gave, besides the known acetylaleuritolic acid(1), a compound (4), $C_{32}H_{52}O_3$, mp 257~8°, which showed hydroxyl peaks at 3520 and 3470 cm⁻¹, acetoxyl peaks at 1710 and 1242 cm⁻¹, and trisubstituted double bond peak at 810 cm⁻¹ in its IR spectrum. Two well-defined double doublets centered at $\delta 5.48 \ (J=4 \text{ and } 8Hz)$ and at $\delta 4.43 \ (J=6.5)$ and 9Hz) together with seven tertiary methyl signals between 80.87 and 1.06 in its NMR spectrum suggested a taraxer-14-ene skeleton bearing a secondary equatorial acetoxyl group at C-3 as indicated in acetylaleuritolic acid¹². The presence of a primary hydroxyl group at C-17 was suggested by a well-defined AB quartet at δ 3. 24 (1H, d, J=11Hz) and 3. 10 (1H, d, J= 11Hz), which was further supported by peaks

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at m/z 453 (M⁺-CH₂OH), 344 (i), 329 (i-CH₃), 284 (i-HAc), 220 (ii) and 189 (iii) in its mass spectrum¹³.

This suggestion was confirmed by the fact that acid-catalized isomerization gave erythrodiol(6), mp 230°, which followed by treatment with Nbromosuccinimide in acetonitrile yielded a bromoepoxide $(7)^{18}$, mp $125\sim8^{\circ}$ and 195° . Therefore this compound was identified as myricadiol 3-acetate, which has not been reported previously¹⁹⁾. The structure of this compound was finally confirmed by partial synthesis from acetylaleuritolic acid. Acetylaleuritolic acid(1) was first reduced with LiAlH4 to give myricadiol (5), mp 260~1°. Acetylation of myricadiol (5) afforded myricadiol diacetate(8), mp 182~ which on mild hydrolysis with K₂CO₃ gave myricadiol 3-acetate(4), mp $258 \sim 9^{\circ}$. This synthetic compound was identical with the natural one in every respect. In the course of our work, Prof. Razdan group²⁰⁾ reported the isolation of epiacetylaleuritolic acid(11) and

phytolaccanol (16) from *Phytolacca acinosa*. However, direct comparisons of their samples (12 and 16) with our acetylaleuritolic acid methylester (15) and myricadiol 3-acetate (4), respectively, led to find that both were identical with ours. The β -configuration of the C-3 acetoxyl group of natural products (1 and 4) was confirmed by direct comparison with synthetic comp

Table I. Physical properties of epimeric pairs and their derivatives.

Compound		Mp	$(\alpha)_{\mathbf{D}}$	Rf
free	3α-OH (9) 3β-OH(13)		- 2.4° +49.4°	0.00
methylester	3α-OH(10) 3β-OH(14)		+ 1.2° +11.1°	
acetate	3α-OAc(11) 3β-OAc (1)		-14.7° +25.5°	
methyl acetate	3α-OAc(12) 3β-OAc(15)		-24.1° +23.5°	

^abenzene: diethyl ether=4:1

bCHCl₃: n-hexane=1:3

ounds.

Oxidation of aleuritolic acid (13) gave aleuritolonic acid (17), mp $286\sim8^{\circ}$, which followed by treatment with aluminum isopropoxide^{21,22)} for 5hr yielded two reaction products. Chromatographic separation of this mixture afforded the less polar compound (9), mp $306\sim7^{\circ}$, as a major reaction product (56%) and followed by the more polar compound (13), mp $304\sim4^{\circ}$.

As shown in Table I, the physical properties of both compounds and their derivatives are clearly different from each other. The mass spectrum of the less polar compound (9) and its derivatives (10, 11 and 12) are summerized in Fig. 1. As expected, the mass spectrum of 9 was very

similar to that of aleuritolic acid (13), with the base peak at m/z 189 and abundant fragments at m/z 302, 287, 269 and 234. As the fragmentations of its derivatives (10 \sim 12) were also similar to those of 3β -epimeric counterparts (14, 1 and 15) with only slight differences in intensities.

However, the NMR spectrum of the less polar compound (9) showed a triplet at $\delta 3.39$ ($J=2.8\,\mathrm{Hz}$), indicating an equatorially oriented C-3 methine proton. An axially oriented methine proton signal was upfield shifted to $\delta 3.19$ (dd, J=7.1 and $9.2\,\mathrm{Hz})^{23}$). As shown in Table II, the changes in chemical shifts and splitting patterns of the C-3 carbinol protons of other deriva-

Fig. 1. Mass fragments of compounds 9, 10, 11 and 12.

Table II. ¹H NMR chemical shifts of epimeric pairs in CDCl₃

compo	und	3-H	15-H	C-CH ₃	others
Free	(9)	3. 39(t, 2. 8)	5. 48(dd, 3. 5, 7. 6)	0.86, 0.93(x6)	
	(13)	3. 19(dd, 7. 1, 9. 2)	5. 49(dd, 3. 5, 7. 4)	0.80, 0.93(x4)	
				0.96(x2)	
Methylester	(10)	3.39(t, 2.8)	5. 49(dd, 3. 7, 7. 7)	0.85, 0.93(x6)	3.57(OCH ₃)
	(14)	3.17(m, W/2=20 Hz)	5.51(dd, 3.6, 7.8)	0.78, 0.93(x5)	3.56(OCH ₃)
				0.96	
Acetate	(11)	4.63(t, 2.4)	5. 51 (dd, 3. 4, 7. 5)	0.83, 0.91, 0.92,	2.04(OAc)
				0.95(x2),	
				0.97(x2)	
	(1)	4. 43(dd, 7, 9)	5. 49(dd, 3. 1, 7. 3)	0.85, 0.88(x2),	2.02(OAc)
				0.92(x4)	
Methyl acetate	(12)	4.63(t, 2.6)	5.50(dd, 3.6, 7.7)	0.83, 0.89,	2.04(OAc)
				0.94(x4), 0.97	3.57(OCH ₃)
	(15)	4. 45(dd, 7, 9)	5.51(dd, 3.1, 7.3)	0.85, 0.88(x2),	2.02(OAc)
				0.92(x4)	3.47(OCH ₃)

Coupling patterns and coupling constants are indicated in parentheses.

tives are in accordance with the above observations. From the above data, the less polar compound must be epialeuritolic acid. A comparison of the ¹³C NMR spectrum of the less polar compound methylester (10) with the spectrum of its epimer (14) further confirmed the above result.

As expected, conversion of the less polar com-

Table III. ¹⁸C NMR chemical shift data of methyl epialeuritolic acid(10) and methyl aleuritolic acid(14).

Carbon	10	14
1	32. 39	38.03(+5.64)
2	25. 17	27.21(+2.04)
3	76. 16	78.96(+2.80)
4	37. 46ª	38.76(+1.30)
5	49. 34 ^b	55.68(+6.34)
6	18. 17	18.76
7	35. 64	35.61
8	39. 21	39. 01
9	48.89 ^b	49. 22
10	38. 11	37.44
11	17. 24	17.32
12	31.80°	31.78a
13	37. 28 ²	37.91
14	160.78	160.62
15	116. 40	116. 51
16	31. 10°	31.07ª
17	51. 32	51.30
18	42.04	42.00
19	41.16	41.22
20	29. 25	29. 24
21	33.87 ^d	33.84 ^b
22	33. 58 ^d	33.51 ^b
23	28. 12	28.01
24	22. 15	15. 42(-6. 73)
25	15. 22	15. 42
26	28.68	28.69
27	26. 32	26. 19
28	178.39	178. 33
29	32. 13	32.14
30	22. 35	22. 41
OCH_3	51.50	51.54

a, b, c, dAssignments bearing the same superscript may be interchanged in each column.

pound methylester (10) to the more polar compound methylester (14) results in an upfield shift of about 6.7ppm for an axial C-4 methyl group, while the equatorial C-4 methyl group is essentially unaffected by this transformation²⁴,²⁵). Moreover, molecular rotation differences between the less polar compound derivatives (9 and 14) and their acetates (11 and 12) were negative values $(-63.2^{\circ} \text{ and } -129^{\circ})^{21},^{22},^{26}$ (Table IV). Therefore, the less polar compound was identified as epialeuritolic acid. Identity of the derivatives from natural products and those from the more polar compound confirmed that the triterpenoid from *Phytolacca* sp. was acetylaleuritolic acid.

All the findings are positive in denying the presence of epiacetylaleuritolic acid (11) and

Table IV. Molecular rotation differences between epihydroxyl compounds and their acetates.

(M) _D	△OAc	Reference
+126°	,,,	(21)
+281°	-155°	
$+105^{\circ}$		(22)
$\pm 297^{\circ}$	-192°	
+154°		(22)
$\pm 179^{\circ}$	-25°	
$+257^{\circ}$		(26)
$\pm 298^{\circ}$	- 41°	
-117°		(26)
- 76.7°	- 40.3°	
- 73.2°		
- 10.9°	-62.3°	•
-123.4°		
+ 5.6°	-129°	
	+126° +281° +105° +297° +154° +179° +257° +298° -117° - 76.7° - 73.2° - 10.9° -123.4°	$+126^{\circ}$ $+281^{\circ}$ -155° $+105^{\circ}$ $+297^{\circ}$ -192° $+154^{\circ}$ $+179^{\circ}$ -25° $+257^{\circ}$ $+298^{\circ}$ -41° -76.7° -40.3° -73.2° -10.9° -62.3°

phytolaccanol (16) in Phytolacca acinosa.

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Literature Cited

- Woo, W.S.: J. Pharm. Soc. Korea 15, 99(1971);
 18, 229(1974); Lloydia 36, 326(1973); Phytochemistry 13, 2887(1974); 14, 1885(1975); J. Natl. Acad. Sci. Korea 17, 271(1978); Proc. 5th Cong. Kor. Sci. Eng. 5, 85(1978).
- Woo, W.S. and Kang, S.S.: J. Pharm. Soc. Korea 17, 152, 161 (1973); 18, 203, 231(1974);
 19, 189(1975); 21, 159(1977); Kor. J. Pharmacogn. 5, 69, 125(1974); 7, 95(1976); 10, 83 (1979).
- 3. Yang, K.S., Woo, W.S. and Kang, S.S.: J. Pharm. Soc. Korea 19, 9(1975).
- Woo, W.S., Shin, K.H. and Kang, S.S.: Kor. J. Pharmacogn. 7, 47(1976).
- Woo, W.S., Chi, H.J. and Kang, S.S.: Kor. J. Pharmacogn. 7, 51(1976).
- Woo, W.S., Kang, S.S., Wagner, H., Seligmann,
 O. and Chari, V.M.: Planta Medica 34, 87 (1978).
- Woo, W.S., Kang, S.S., Wagner, H. and Chari,
 V.M.: Tetrahedron Lett. 1978, 3239.
- Woo, W. S., Kang, S.S., Yamasaki, K. and Tanaka, O.: Arch. Pharm. Res. 1, 21(1978).
- 9. Kang, S.S. and Woo, W.S.: J. Nat. Prod. 43, 510(1980).
- 10. Woo, W.S., Kang, S.S., Seligmann, O., Chari,

- V.M. and Wagner, H.: Tetrahedron Lett. 21, 4255(1980).
- 11. Woo, W.S., Kang, S.S., Seligmann, O. and Wagner, H.: *Arch. Pharm. Res.* 5, 1(1982).
- 12. Hui, W.-H. and Li, M.-M.: J. Chem. Soc. Perkin I, 23(1976); 897(1977).
- Budzikiewicz, H., Wilson, J.M. and Djerassi, C.: J. Am. Chem. Soc. 85, 3685(1963).
- Nakanishi, K., Goto, T., Ito, S., Natori, S. and Nozoe, S.: Natural Products Chemistry, Vol. 1, Academic press, New York, Chap. 5, pp. 347-348(1974).
- Misra, D.R. Khastgir, H.N.: Tetrahedron 26, 3017(1970).
- 16. Burke, D.E. and Le Quesne, P.W.: *Phytochemistry* 10, 3319(1971).
- 17. Woo, W.S. and Wagner, H.: *Phytochemistry* 16, 1845(1977).
- 18. Woo, W.S., Kang, S.S. and Jew, S.S.: *Planta Medica* 501 (1985).
- 19. Woo, W.S. and Kang, S.S.: *Phytochemistry* 24, 1116(1985).
- 20. Razdan, T.K., Harkar, S., Kachroo, V. and Koul, G.L.: Phytochemistry 21, 2339(1982).
- 21. Huneck, S.: Tetrahedron 19, 479(1963).
- 22. Huneck, S. and Snatzke, G.: Chem. Ber. 98, 120(1965).
- 23. Corbett, R.E., Cumming, S.D. and Whitehead, E.V.: J. Chem. Soc. Perkin I, 2827(1972).
- 24. Crews, P. and Kho-Wiseman, E.: Tetrahedron Lett. 2483(1978).
- Chen, T.K., Ales, D.C., Baenziger, N.C. and Wiemer, D.F.: J. Org. Chem. 48, 3525(1983).
- 26. Pradhan, B.P., Hassan, A. and Ray, T.: Tetrahedron 41, 2513(1985).