

Liver Protective Activities of Korean Medicinal Plants

—Pharmacology of *Plantago Semen*—

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Plantago semen, the seeds of *plantago* species, have long been used for medicinal purposes for controlling various liver diseases. In addition, they have been used as anti-inflammatory, antitussive, obstipant and diuretic agents for treating various alimentary, respiratory and renal disorders.¹⁻⁵⁾

On the course of our preliminary screening tests for hepatotonic plant materials as shown on Table I, several of the plant materials including *plantago semen* were found to exert potent liver protective activities against carbon tetrachloride intoxication in mice,^{6,7)} The biochemical and pharmacological examinations on the anti-hepatotoxic effects of *plantago semen* have been undertaken.

Reports have been indicated that rat liver intoxication with halogenated hydrocarbons, especially, carbon tetrachloride caused structural and metabolic changes in hepatic tissues and cells, with distinctive changes of endoplasmic reticulum and marked depression in microsomal enzymatic activities and hepatic protein syntheses.^{8,10)} Following such disturbances, liver necrosis is usually accompanied.¹¹⁾ The chosen dose schedule of carbon tetrachloride in our experiments could induce histological changes in appearance of liver closely equivalent to diffused hepatitis in men.^{12,13)}

Table II shows the dose schedule of carbon tetrachloride and test samples against carbon

tetrachloride intoxication. Each mice was treated for four days with saline, CCl₄ and test samples

Table 1. List of medicinal plants and their liver protective activities.⁶⁾

Family name	Scientific name	Parts of plants	Activities ^(a)
Lardizabalaceae	<i>Akebia quinata</i>	vi	Toxic
Alismataceae	<i>Alisma orientale</i>	tu	#
Umbelliferae	<i>Angelica gigas</i>	ra	+
Compositae	<i>Artemisia Messer-Schmidtiana var. viridis f. typica</i>	ha	-
Aristolochiaceae	<i>Asiasarum sieboldii</i>	wp	Toxic
Liliaceae	<i>Asparagus cochinchinensis</i>	tu	#
Leguminosae	<i>Astragalus membranaceus</i>	ra	0
Compositae	<i>Atractylodes japonica</i>	rh	Toxic
Compositae	<i>Atractylodes japonica</i>	rh (alba)	#
Umbelliferae	<i>Bupleurum falcatum</i>	ra	-
Umbelliferae	<i>Bupleurum longerdium</i>	ra	Toxic
Compositae	<i>Carduus crispus</i>	ha	0
Leguminosae	<i>Cassia tora</i>	sm	#
Amarantaceae	<i>Celosia argentea</i>	sm	--
Papaveraceae	<i>Chelidonium majus</i>	ha	-
Cyatheaceae	<i>Cibotium barometz</i>	rh	#
Compositae	<i>Cirsium pendulum</i>	ha	-
Rutaceae	<i>Citrus aurantium</i>	fs	0
Cornaceae	<i>Cornus officinalis</i>	fr	0
Rosaceae	<i>Crataegus pinnatifida</i>	fr	+
Cyperaceae	<i>Cyperus rotundus</i>	tu	#
Equisetaceae	<i>Equisetum hyemale var. japonicum</i>	ha	#
Rutaceae	<i>Evodia rutaecarpa</i>	fr	#
Gentianaceae	<i>Gentiana scabra</i>	ra	#

Leguminosae	<i>Glycyrrhiza uralensis</i>	ra	††
Solanaceae	<i>Lycium chinense</i>	fr	+
Polyporaceae	<i>Pachyma hoelen</i>	sc	††
Ranunculaceae	<i>Paeonia albiflora var. trichocarpa</i>	ra	††
Araliaceae	<i>Panax ginseng</i>	ra	††
Scrophulariaceae	<i>Picrorrhiza kurroa</i>	rh	††
Araceae	<i>Pinellia ternata</i>	tu	0
Plantaginaceae	<i>Plantago asiatica</i>	sm	††
Liliaceae	<i>Polygonatum japonicum</i>	rh	††
Polygonaceae	<i>Polygonum multiflorum</i>	ra	--
Rutaceae	<i>Poncirus trifoliata</i>	fs	††
Labiatae	<i>Prunella vulgaris</i>	ha	-
Rosaceae	<i>Prunus persica</i>	sm	††
Scrophulariaceae	<i>Rehmania glutinosa</i>	ra	††
Rhamnaceae	<i>Rhamnus crenatus</i>	b	††
Polygonaceae	<i>Rheum undulatum</i>	rh	††
Labiatae	<i>Salvia multicorrhiza</i>	ra	+
Labiatae	<i>Scutellaria baicalensis</i>	ra	+
Menispermaceae	<i>Sinomenium acutum</i>	ra	+
Compositae	<i>Taraxacum platycarpum</i>	wp	-

a: activities counteracting carbon tetrachloride intoxication measured with the duration of sleeping time after injection of hexobarbital.

b: bark fr: fruit skin ha: herba ra: radix rh: rhizome sc: sclerotium sm: semen tu: tuber vi: vine wp: whole plant

as indicated and pharmacological and biochemical evaluations were undertaken on the fifth day.

Table 2. Schedule of treatment of mice with carbon tetrachloride and test samples.

	Days					
	1	2	3	4	5	
Control	Sal.	Sal.	Sal.	Sal.	Sal.	Biochemical or pharmacological measurements were undertaken
CCl ₄	Sal.	CCl ₄	CCl ₄	Sal.		
Test samples	T.S.	T.S. + CCl ₄	T.S. + CCl ₄	T.S.		

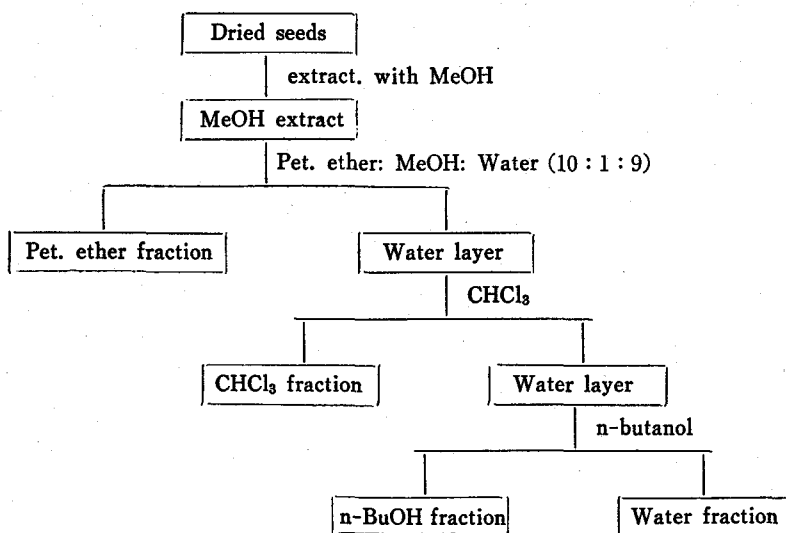
Sal.: 0.9% physiological saline, T.S.: Test sample in saline

Sample preparations were made with plantago semen for hepatotonic activity tests as described in chart I. Total methanol extract was first prepared from plantago semen and fractionated into four different solvent fractions; petroleum ether, chloroform, n-butanol and water fractions. Methanol extract and each fraction was subjected to pharmacological and biochemical evaluations.

1) Effects on the duration of sleeping time induced by hexobarbital:

The duration of sleeping time was measured

Chart I. Fractionation of methanol extract of Plantago semen.



following hexobarbital injection ¹⁴⁾ on the fifth day of experimental schedule as appeared in Table II. Carbon tetrachloride intoxication caused elongation of sleeping time comparing with that of control group which could be explained by the reduction of enzymatic metabolism of hexobarbital in liver. Table III shows the favorable effects of methanol extract, water and chloroform fractions of plantago semen counter-acting carbon tetrachloride intoxication.

Especially, the effect of water fraction restored the sleeping time to nearly normal state. However, the duration of sleeping became longer with butanol and petroleum fractions.

Table 3. Effects of plantago semen on duration of sleeping time induced by hexobarbital.

	duration of sleep
Control	13
CCl ₄	21
MeOH ext.+CCl ₄	16
H ₂ O fr.+CCl ₄	14
n-BuOH fr.+CCl ₄	27
CHCl ₃ fr.+CCl ₄	18
Pet, Ether fr.+CCl ₄	73

2) Effects on serum transaminase activities:

The elevation of serum glutamate-oxaloacetate transaminase (S-GOT, EC. 2, 6, 1, 1) and serum glutamate-pyruvate transaminase (S-GPT, EC. 2, 6, 1, 2) activities have very close relationships. ^{15,16)} On the fifth day of experiment following four-days of treatment of mice as on Table II, blood samples were collected and the enzymatic activities were assayed.¹⁴⁾ As the fig. I and II show, both S-GOT and S-GPT activities were markedly increased with CCl₄ intoxication comparing with the control. Methanol extract, water and chloroform fraction treated group of mice showed S-GOT and S-GPT acti-

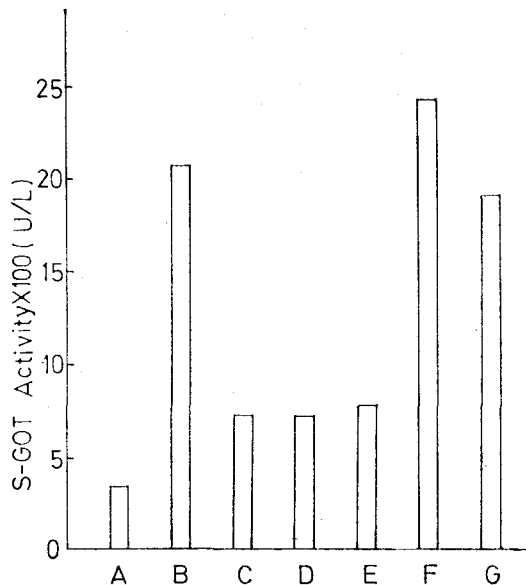


Fig. 1. S-GOT activities

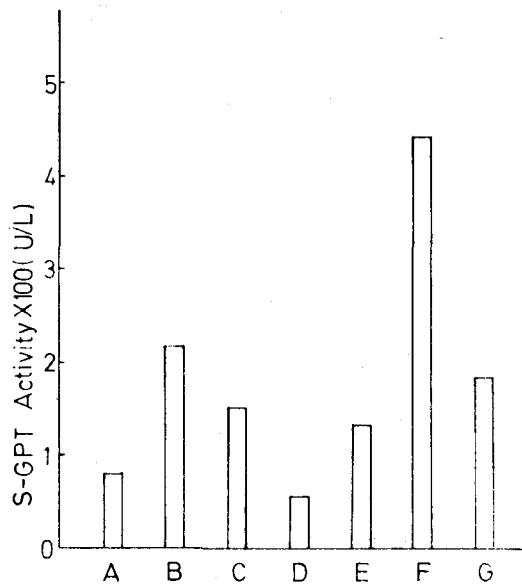


Fig. 2. S-GPT activities

A: Control, B: CCl₄, C: Methanol extract+CCl₄, D: H₂O fraction+CCl₄, E: CHCl₃ fraction+CCl₄, F: Pet. ether fraction+CCl₄, G: n-BuOH fraction+CCl₄

vities comparable with that of control group while petroleum and butanol fractions rather elevated S-GOT and S-GPT levels. These biochemical data appeared to be correlated with

the pharmacological evaluation that methanol extract, water and chloroform fractions of plantago semen have counteracting effects against CCl_4 intoxication.

3) Histological observation of liver tissue:

On the day of five, after treating mice as the schedule on Table II, liver slices were prepared with each mice for direct microscopic observations. The biopsy data confirmed the previous pharmacological and biochemical evaluations.¹⁷⁾ With the administration of CCl_4 , liver showed typical hepatic symptoms especially around portal area. Treatment with methanol extract, water or chloroform fraction of plantago semen together with CCl_4 reduced the hepatotoxic effects of CCl_4 restoring the overall condition near normal state. Butanol and petroleum ether fraction showed rather toxicity. The overall histological observation appeared to be coincide with the results obtained from measurement of the duration of sleeping time induced by hexobarbital and serum transaminase activities that the whole methanol extract, water and chloroform fraction had liver protective effects and that especially the hepatotonic effect of water fraction was significant.

The water fraction of plantago semen contain various mono-, di-, tri- and poly-saccharides and the presence of aucubin was confirmed. The chemical and biological mechanism of active constituents from plantago semen should next be investigated.

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References

1. Heh, Joon: Secrets of Oriental Medicines-Translations into Modern Language, Pyungyun Publishing Co., Seoul, Korea(1966)
2. Lee, S.J.: Korean Folk Medicines, Seoul National University Publishing Office (1966)
3. Yun, G. Y.: Oriental Prescriptions, Minersa, Seoul, Korea (1964)
4. Herbal pharmacology in the people's Republic of China -Trip Report of the the American Herbal Pharmacology Delegation: National Academy of Science, Washington, D.C. (1975)
5. Akamasu, E.: Modern Oriental Drugs, Yishiyakusha, Tokyo, Japan (1970)
6. Yun (Choi), H.S. and Chang I.M.: Plants with Liver Protective Activities(I), *Korean J. Pharmacog.* 8, 125 (1977)
7. Chang, I.M. and Yun (Choi), H.S.: Plants with Liver Protectivities (III), *Korean J. Pharmacog.* 10, 79 (1979)
8. Smuckler, E. A. and Benditt, E.A.: *Science* (N.Y.), 140, 308 (1963)
9. Recknagel, R.O. and Ghoshal, A.K.: *Exp. Mol. Pathol.* 5, 108 (1966)
10. Ress, K.R. and Sinha, K.P.: *J. Pathol. Bacteriol.*, 80297 (1990)
11. Wigglesworth, J.S.: *J.Pathol. Bacteriol.*, 87, 333 (1964)
12. Hahn, V.G., Lehmann, H.D., Kurten, M., Uebel, H. and Vogel,G.: *Arzneim. Forsch.*, 18, 698 (1968)
13. Marchand, C., McLean, S., Plaa, G.L. and Traiger G.: *Biochem. Pharmacol.*, 20, 869 (1971)
14. Chang, I.M. and Yun(Choi), H.S., *Kor. J. Pharmacog.*, 9, 139 (1978)
15. Balazs, T., Murray, T.K., McLaughlan, J.M. and Grice, H.C.: *Toxic. Appl. Pharmac.*, 3, 71 (1961)
16. Zimmerman, H.J., Kodera, Y. and West, M.: *J. Lab. Clin. Med.*, 66, 315 (1965)
17. Chang, I.M. and Yun (Choi), H. S., unpublished data