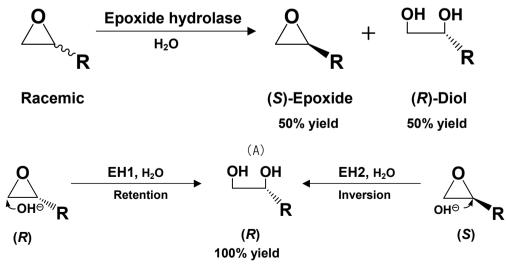
## Marine and Microbial Epoxide Hydrolases and Their Application to Stereospecific Biocatalysis

Eun Yeol Lee\* and Hee Sook Kim

Department of Food Science & Technology, Kyungsung University

Chiral epoxides and diols are valuable intermediates for the synthesis of high-value-added chiral pharmaceuticals (1). Chiral epoxide and diols can be prepared using epoxide hydrolase(EH)-catalyzed enantioselective hydrolysis of racemic epoxides. EH is an enzyme that catalyses a hydrolysis reaction of an epoxide to the corresponding vicinal diol (2). Enantioselective hydrolysis of one enantiomer of racemic epoxides by EHs yields the remaining enantiomer in an enantiopure form(Fig. 1A). EH is a cofactor-independent, relatively stable and easy-to-use biocatalyst (3). EHs are ubiquitous enzymes. Recently, marine bioresource-originated EHs have been characterized for the development of novel EHs (Fig. 2)(4-6). Some marine and microbial EHs have been shown to have enantio-complementary regioselectivity on racemic epoxide substrates, leading to an enantioconvergent process (Fig. 1B). In this presentation, the discovery of EHs from marine/microbial bioresources and its application to stereospecific biocatalysis will be presented.



(B)

Fig. 1. The kinetic resolution(A) and enantioconvergent hydrolysis(B) of racemic epoxides for preparation of chiral epoxides and diols using EHs(from Ref. 2).

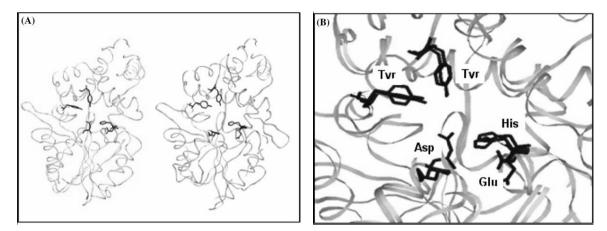


Fig. 2. Superimposition of *Mugil cephalus* mEH model (gray) on *Aspergillus niger* EH, 1qo7.pdb (yellow), shows similarity of the active site region and residues (blue wire frame). The representation was made using RASWIN. *M. cephalus* mEH model was constructed using 396 residues from 62<sup>nd</sup> to 467<sup>th</sup> amino acid sequence.
(A) Structural alignment of the putative mEH model and 1qo7.pdb. (B) Expanded view of active site region with the catalytic triad and two tyrosine residues (from Ref. 6).

## Acknowledgment

This work was supported by the Marine and Extreme Genome Research Center Program, Ministry of Marine Affairs and Fisheries, Republic of Korea.

## References

- Steinreiber A, Faber K. 2001. Microbial epoxide hydrolases for preparative biotransformations. Curr. Opin. Biotechnol. 12, 552-558.
- 2. Lee EY, Shuler ML. 2007. Molecular engineering of epoxide hydrolase and its application to asymmetric and enantioconvergent hydrolysis. Biotechnol. Bioeng. *Accepted for publication*.
- Kim HS, Lee SJ, Lee EY. 2006. Development and characterization of recombinant whole-cell biocatalysts expressing epoxide hydrolase from *Rhodotorula glutinis* for enantioselective resolution of racemic epoxides. J. Mol. Catal. B: Enzym. 43, 2-8.
- Kim HS, Lee SJ, Lee EJ, Hwang JW, Park S, Kim SJ, Lee EY. 2005. Cloning and characterization of a fish microsomal epoxide hydrolase of *Danio rerio* and application to kinetic resolution of racemic styrene oxide. J. Mol. Catal. B: Enzym. 37, 30-35.
- 5. Kim HS, Lee OK, Lee SJ, Hwang S, Kim SJ, Yang SH, Park S, Lee EY. 2006. Enantioselective epoxide hydrolase activity of a newly isolated microorganism, *Sphingomonas echinoides* EH-983, from seawater. J. Mol. Catal. B: Enzymatic 41, 130-135.
- 6. Soo Jung Lee, Hee Sook Kim Sang Jin, Sunghoon Park, Beum June Kim, Michael L. Shuler, Eun Yeol Lee (2007) Cloning, expression and enantioselective hydrolytic catalysis of a microsomal epoxide hydrolase from a marine fish, *Mugil cephalus*. Biotechnol. Lett., 29, 237-246.