

Engineering the deoxysugar moiety of polyketides in *Streptomyces venezuelae*

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Several strategies using combinatorial biosynthesis were employed to produce polyketides with altered glycosylation pattern in *Streptomyces venezuelae* ATCC 15439. This wild type strain was manipulated by deleting different portions of the pikromycin genetic cluster to produce mutants that produce only the aglycon (10-deoxymethynolide and narbonolide), the deoxysugar (TDP-D-desosamine) or a completely deleted strain that is a non-producer, which were named YJ003, DHS2001 and YJ028, respectively. Firstly, a two plasmid system expressing the genes for the biosynthesis and transfer of exogenous deoxysugars, TDP-D-quinovose or TDP-D-olivose was incorporated into the strain YJ003 to produce novel quinovose or olivose glycosylated 10-deoxymethynolide and narbolide. Secondly, expression of the tylactone PKS from another two plasmid system in the strain DHS2001 led to the production of desosamine glycosylated tylactone. Thirdly, feeding of tylactone to the strain YJ028 that was transformed by plasmids expressing the TDP-quinovose or TDP-olivose biosynthetic and transfer genes led to the production of quinovose glycosylated or olivose glycosylated tylactone. In all these systems, an auxiliary protein besides the glycosyltransferase was found to be essential for glycosylation to proceed efficiently. The pairing of glycosyltransferase and auxiliary protein in glycosylation was further examined by deleting the gene encoding the auxiliary protein (DesVIII) from the wildtype *S. venezuelae* and complementation by exogenous auxiliary proteins from other polyketide gene clusters. These results demonstrate a successful attempt of genetically engineering the *S. venezuelae* for the generation of novel hybrid macrolide antibiotics.

Recent publication

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