

Oxalate Biosynthesis of lettuce grown in salt accumulated soil

Institute of Ecological Phytochemistry, Hankyong National University, Ansong 456-749, Korea : Sei-Joon Park,
Myung Yong Shim

Department of Plant Resources Science, Hankyong National University, Ansong 456-749, Korea : Sung-Yung
Yoo, Hyun-Hee Kim, Jung-Ah Jung, Seung-Gil Yun, Sang-Eun Lee, Tae Wan Kim*

Objectives

It has been well known that Ca fertilization has increased the efflux of NO_3^- or decreased an absorption of NO_3^- . A lot of reports had implied that calcium oxalate crystal may represent storage forms of calcium and oxalic acid, and there has been evidence of calcium oxalate resorption in times of calcium depletion. However, there are a few reports on a change in Ca-oxalate biosynthesis under surplus N-supplement. In this study, we had tried to illuminate the effect of lime application on and oxalate-biosynthesis under high salt stress.

Materials and Methods

- Soil preparation : Electrical conductivity (EC) of upland clay loamy soil with low fertility was adjusted to (low salt) and 5 dSm^{-1} (high salt) using compost and chemical fertilizer. The physico-chemical properties of soils were shown in table. Lime was applied in the level of $200 \text{ Kg } 10 \text{ a}^{-1}$ 30 days before transplanting.
- Proline and betaine analysis : Free proline was determined in 95% ethanol extracts from lettuce leaves. Samples of 0.5g tissues freshly harvested were crushed in 5mL 95% (v/v) ethanol. The insoluble fraction of the extract was washed twice with 5mL of 70% ethanol. All soluble fractions were centrifuged at $3,500 \times g$ for 10 min. The supernatants were collected and stored at 4°C for proline determination. Betaine was determined in 70% ethanol extracts from lettuce leaves. The betaine content was accounted according to the method by Comer et al (2000). Total Kjeldahl N in a sulfuric acid digestion was determined with a Lachat Autoanalyzer (Diamond, 1992) on leaf material that had been dried at 70°C for 48 h and ground through a 40 mesh screen. N concentrations were determined for individual leaves and expressed as Mg Kg^{-1} tissue. Other inorganic ions were analyzed using ICP. Sampled leaves were divided to two portions. One portion was directly extracted in de-ionized water (leaf extract), other was digested in $\text{HClO}_4 + \text{H}_2\text{SO}_4$.

Address : Tae Wan Kim E-mail: Taewkim@hknu.ac.kr Tel: 31-670-5081

RESULTS AND DISCUSSION

To investigate the oxalate biosynthesis lettuce (*Lactuca sativa*) were grown in the soil with electrical conductivity (EC) 1 and 5 dSm⁻¹, respectively. Betaine biosynthesis was also enhanced with nitrogen application. However, proline biosynthesis was not activated. Furthermore, it was observed that lime fertilization has exhibited an effect on the increase in chlorophyll biosynthesis, Chl a/b ratios and photosynthesis efficiency. But biomass productivity was rather reduced. It may paradoxically thicken by lime treatment. It was illuminated that pheophytin biosynthesis were not related with a lime treatment. Interestingly, soluble oxalate synthesis was extremely activated in low salt soil (EC 1dSm⁻¹) whereas insoluble oxalate was actively synthesized in high salt soil (EC 5dSm⁻¹). Ca application to lettuce rhizosphere has stimulated P and K assimilation.

Table 1. The biosynthesis of proline, betaine, anthocyanin, pheophytin and oxalate induced by salt and nitrogen stress

Treatments	Proline (mg kg ⁻¹)	Betaine (mg kg ⁻¹)	Anthocyanin (g .g ⁻¹ fw)	Pheophytin (g.g ⁻¹ fw)	Oxalate(mmol L ⁻¹ fw)	
					Soluble	Insoluble
Soil EC						
1 dSm ⁻¹	299	3.1	4.58	106.4	0.56	0.43
5 dSm ⁻¹	268	45.9	5.87	135.7	0.30	1.06
N fertilization						
1/2 ST*	355	21.3	4.41	69.3	0.56	0.65
Soil testing	297	12.5	5.26	155.3	0.30	0.90
2 fold ST	227	36.0	5.22	111.1	0.53	0.67
2 fold ST+lime	256	28.3	5.98	148.6	0.45	0.77

*ST : Total fertilization of N-P₂O₅-K₂O=20.0-5.9-12.8 Kg 10a⁻¹ subtracting the contents included in experimental soils.

ACKNOWLEDGEMENTS

This work was financially supported by GRRC project (No.3-1) from Hankyong National University and Bio-Green 21 from RDA.

REFERENCES

- Colmer, T. D., Coradini, F., Cawthray, G. R., Otte, M. L. (2000) Analysis of dimethyl- sulphonioacetate (DMSP) betaines and other organic solutes in plant tissue extracts using HPLC. *Phytochem. Anal.* 11, 163-168.
- Ota, K., Kagawa, A. (1996) Effect of nitrogen nutrients on the oxalate content in spinach (*Spinacea oleraceae*) plant. *J. Japan. Soc. Hort. Sci.* 65, 327-332