

Gene-technological enhancement of stress tolerance of plants by biosynthesis of glycinebetaine

Norio Murata

National Institute for Basic Biology, Okazaki 444-8585, Japan

Many halotolerant plants, bacteria and algae accumulate the compatible solute glycinebetaine (betaine) in response to environmental stresses, such as high concentrations of salt, low temperature and drought. Betaine acts by stabilizing the cellular structure and maintaining the function of proteins and membranes. To generate plants capable of the biosynthesis of betaine, the *codA* gene encoding choline oxidase, the enzyme that synthesizes betaine, was isolated from the soil bacterium *Arthrobacter globiformis*. *Arabidopsis thaliana* was transformed with a modified *codA* gene containing an additional leader sequence for transport into chloroplasts. The resultant transgenic plants were more tolerant to salt, cold and heat stress than wild-type plants during both germination of seeds and the growth of young and mature plants. Seeds from the transgenic plants also tolerated cold and heat stress during imbibition. Moreover, transformation with the *codA* gene enhanced tolerance to salt, cold and high-light stress in the photosynthetic machinery and freezing tolerance of matured plants.

Recently, we examined the tolerance of transformed plants to salt stress at the reproductive stage, which is when plants are most sensitive to environmental stress. Salt-shock treatment of wild-type plants for 3 days resulted in abortion of flower buds and decreased the number of seeds per silique. These deleterious effects were clearly visible 6 days

after the termination of the salt-shock treatment. Microscopic examination of floral structures revealed that salt stress inhibited the development of anthers, pistils and petals. In particular, the production of pollen grains and ovules was dramatically inhibited. These effects of salt stress were significantly reduced by transformation with the *codA* gene, and our observations suggest that the enhanced tolerance of the transgenic plants is a result of the accumulation of betaine in the reproductive organs. Transformation with the *codA* gene has an additional unique characteristic. Such transformed plants grow faster and produce a more number of seeds than wild-type plants even under non-stress conditions, enhancing the productivity of the plant.

We also transformed the rice (*Oryza sativa* L.) japonica variety with the *codA* gene. Levels of betaine were as high as 1 and 5 mol per gram fresh weight of leaves in plants where choline oxidase was targeted to the chloroplast and the cytosol, respectively. Treatment of plants with 0.15 M NaCl inhibited growth of both wild-type and transgenic plants. However, after removal of the salt stress the transgenic plants began to grow again at the normal rate after a significantly shorter time than wild-type plants. Inactivation of photosynthesis indicated that plants exposed to salt stress and low temperature were more tolerant to photoinhibition if they contained a chloroplast-targeted choline oxidase than a

cytosol-targetedone. This indicates that the subcellular compartmentalization of the biosynthesis of betaine is a critical factor in the efficient enhancement of tolerance to stress in the engineered plants. We have also transformed the indica variety with the *codA* gene. This transgene also had enhanced tolerance of salt stress in young plants.

Very recently we transformed tomato (*Lycopersicon esculentum* Mill.) plants, which normally do not accumulate betaine and are susceptible to chilling stress, with the *codA* gene. Exposure to temperatures below 10 °C causes various injuries and greatly decreases fruit set in most cultivars. Transgenic plants expressing *codA* accumulate betaine in their leaves and reproductive organs at levels of upto 0.3 and 1.2 mol·g⁻¹ fresh weight, respectively. Over various developmental phases, from seed imbibition to fruit production, these betaine-accumulating plants are more tolerant to chilling stress than their wild-type counterparts. During reproduction, they yield, on average, 10-30% more fruits following chilling stress.

References

Reviews

1. G. C. Papageorgiou and N. Murata (1995) The unusually strong stabilizing effect of glycinebetaine on the structure and function of the oxygen-evolving photosystem II complex. *Photosynth. Res.* 44: 243-252.
2. H. Hayashi and N. Murata (1998) Genetically engineered enhancement of salt tolerance in higher plants. *Stress Responses of Photosynthetic Organisms* (eds., K. Satoh and N. Murata) p. 133-148. Elsevier Science, Amsterdam
3. A. Sakamoto and N. Murata (2000) Genetic engineering of glycinebetaine synthesis in plants: current status and implication for enhancement of stress tolerance. *J. Exp. Bot.* 51: 81-88.
4. A. Sakamoto and N. Murata (2001) The use of

choline oxidase, a glycinebetaine-synthesizing enzyme, to create stress resistant transgenic plants. *Plant Physiol. (Update)* 125: 180-188

5. A. Sakamoto and N. Murata (2002) The role of glycine betaine in the protection of plants from stress: clues from transgenic plants. *Plant Cell Environ.* 25: 163-171
6. T. H. H. Chen and N. Murata (2002) Enhancement of tolerance to abiotic stress by metabolic engineering of betaines and other compatible solutes. *Curr. Opin. Plant Biol.* 5: 250-257

Original articles

7. P. Deshniem, D.A. Los, H. Hayashi, L. Mustardy and N. Murata (1995) Transformation of *Synechococcus* with a gene for choline oxidase enhances tolerance to salt stress. *Plant Mol. Biol.*, 29:897-907
8. P. Deshniem, Z. Gombos, Y. Nishiyama and N. Murata (1997) The action *in vivo* of glycine betaine in enhancement of tolerance of *Synechococcus* sp. strain PCC 7942 to low temperatures. *J. Bacteriol.*, 179:339-344
9. H. Hayashi, Alia, L. Mustardy, P. Deshniem, M. Ida and N. Murata (1997) Transformation of *Arabidopsis thaliana* with the *codA* gene for choline oxidase; accumulation of glycinebetaine and enhanced tolerance to salt and cold stress. *Plant J.*, 12:133-142
10. Alia, H. Hayashi, T. H. H. Chen and N. Murata (1998) Transformation with a gene for choline oxidase enhances the cold tolerance of *Arabidopsis* during germination and early growth. *Plant Cell Environ.*, 21: 232-239
11. A. Sakamoto, Alia and N. Murata (1998) Metabolic engineering of rice leading to biosynthesis of glycinebetaine and tolerance to salt and cold. *Plant Mol. Biol.*, 38: 1011-1019
12. H. Hayashi, Alia, A. Sakamoto, H. Nonaka, T. H. H. Chen and N. Murata (1998) Enhanced germination under high-salt conditions of seeds of transgenic *Arabidopsis* with a bacterial gene (*codA*) for choline oxidase. *J.*

- Plant Res., 111: 357-362
13. Alia, H. Hayashi, A. Sakamoto and N. Murata (1998) Enhancement of the tolerance of *Arabidopsis* to high temperatures by genetic engineering of the synthesis of glycinebetaine. Plant J., 16: 155-161
 14. Alia, Y. Kondo, A. Sakamoto, H. Nonaka, H. Hayashi, P. P. Saradhi, T.H.H. Chen and N. Murata (1999) Enhanced tolerance to light stress of transgenic *Arabidopsis* plants that express the *codA* gene for a bacterial choline oxidase. Plant Mol. Biol., 40: 279-288
 15. A. Sakamoto, R. Valverde, Alia, T. H. H. Chen and N. Murata (2000) Transformation of *Arabidopsis* with the *codA* gene for choline oxidase enhances freezing tolerance of plants. Plant J., 22: 449-453
 16. M. Gao, A. Sakamoto, K. Miura, N. Murata, A. Sugiura and R. Tao (2000) Transformation of Japanese persimmon (*Diospyros kaki* Thunb.) with a bacterial gene for choline oxidase. Mol. Breeding, 6: 501-510
 17. A. Mohanty, H. Kathuria, A. Ferjani, A. Sakamoto, P. Mohanty, N. Murata and A. K. Tyagi (2002) Transgenics of an elite *indica* rice variety Pusa Basmati 1 harbouring the *codA* gene are highly tolerant to salt stress. Theoret. Appl. Genetics, 106: 51-57
 18. R. Sulpice, H. Tsukaya, H. Nonaka, T. H. H. Chen and N. Murata (2003) Enhanced formation of flowers and seeds in salt-stressed *Arabidopsis* after genetic engineering of the accumulation of glycinebetaine. Plant J. 36: 165-176
 19. E.-J. Park, Z. Jekni, A. Sakamoto, J. DeNoma, R. Yuwansiri, N. Murata and T. H. H. Chen (2004) Genetic engineering of glycinebetaine synthesis in tomato protects seeds, plants and flowers from chilling damage. Plant J. in press

Norio Murata

Nationality : Japan

Position : National Institute for Basic Biology
Myodaiji, Okazaki 444-8585, Japan

Phone : +81-564-55-7600

Fax : +81-564-54-4866

E.mail : murata@nibb.ac.jp

FINAL EDUCATION(Ph.D.)

1969 D. Sci. The University of Tokyo, Photosynthesis

EMPLOYMENT

1966 - 1978 Instructor, the University of Tokyo, Faculty of Science
1969 - 1970 Post-doctoral fellow, Carnegie Institution of Washington,
Department of Plant Biology
1978 - 1985 Associate Professor, the University of Tokyo,
College of Arts and Sciences
1985 - Present Professor, National Institute for Basic Biology (NIBB)
1989 - 1991; Chairman, Department of Regulation Biology (NIBB)
1993 - 1998
1999 2000 Dean, School of Life Science, Graduated University for
Advanced Studies

RESEARCH INTERESTS

- (1) Molecular biology of stress physiology
- (2) Biophysics and molecular biology of membrane lipids
- (3) Regulatory aspects of photosynthesis
- (4) Gene engineering of stress tolerance of plants
- (5) Signal transduction pathway of environmental stress