

- Plant Pathol. J.* 19:143-147.
- Kim, D. S., Chun, S. J., Jeon, J. J., Lee, S. W. and Joe, G. H. 2004. Synthesis and fungicidal activity of ethaboxam against Oomycetes. *Pest Management Sci.* 60: 1007-1012.
- Kim, D.S., Lee, Y. S., Chun, S. J., Choi, W. B., Lee, S. W., Kim, G. T., Kang, K. G., Joe, G. H. and Cho J. H. 2002. Ethaboxam: a new oomycetes fungicide. *Proc. British Crop Protect Conf.-Pest and Disease BCPC*, Farnham, Surrey, UK, pp 377-382.
- Kim, D. S., Park, H. C., Chun, S. J., Yu, S. H., Choi, K. J., Oh, J. H., Shin, K. H., Koh, Y. J., Kim, B. S., Hahm, Y. I. and Chung, B. K. 1999. Field performance of a new fungicide ethaboxam against cucumber downy mildew, potato late blight and pepper phytophthora blight in Korea. *Plant Pathol. J.* 15:48-52.
- Uchida, M., Roberson, R. W., Chun, S. J. and Kim, D. S. 2004. *In vivo* effects of the fungicide ethaboxam on microtubules integrity in *Phytophthora infestans*. *Pest Management Sci.* (submitted).

SII-4

Plant growth promotion and induced systemic resistance by a selected PGPR strain, *Bacillus amyloliquefaciens* EXTN-1

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Various non-pathogenic Rhizobacteria have the ability to induce systemic resistance in plant, which provides protection against a broad spectrum of phytopathogenic microorganisms including fungi, bacteria, and viruses. Researches have demonstrated that induced systemic resistance (ISR) can be a potential mechanism by which plant growth-promoting rhizobacteria (PGPR) reduced diseases (Wei, 1991, Lie *et al.*, 1996). ISR acts through a different signaling pathway to that systemic acquired resistance(SAR), the ISR pathway is induced when the plant is challenged by pathogens. Bacterial determinants that are claimed to produce ISR including siderophore, O-antigen of lipopolysaccharide, pyoverdine and salicylic acid. The purpose of this research was application and elucidation of systemic resistance by *Bacillus amyloliquefaciens* EXTN-1.

Soil drenching or seed priming (10^6 cfu/ml) of *Bacillus amyloliquefaciens* strain EXTN-1 stimulated seed germination and growth of about 20 crops used without any harmful effect. Furthermore, treatment of *B. amyloliquefaciens* strain EXTN-1 showed a broad disease-controlling spectrum to the plant diseases caused by viral, bacterial, and fungal plant pathogens such as cucumber mosaic virus, tobacco mosaic virus, potato virus Y and X, *Pseudomonas syringae* pv. *lacrymans*, *Ralstonia solanacearum*, *Colletotrichum orbiculare*, *Magnaphorte grishia*, and *Fusarium oxysporum* (Park *et al.*, 2001).

Lettuce Plants with EXTN-1 showed great growth promotion compared to that of untreated control (Fig. 1). When *B. amyloliquefaciens* strain EXTN-1 was drenched to lettuce grown in hydroponic system, the population of *B. amyloliquefaciens* strain EXTN-1 was similar or increased in the rhizosphere compared to that of initially treated population, while the population was gradually decreased up to 10 folds in the hydroponic solution 4 weeks after treatment. In another experiment, we found that induced systemic resistance and plant growth promotion activity by EXTN-1 strongly showed in cool season than summer season. In case of cucumber, soil drenching after seed coating with EXTN-1 showed best disease protection and plant growth promotion in soil as well as in hydroponic

system as method of application for EXTN-1.

Treatment of EXTN-1 increased H_2O_2 amount in early stage and induced the expression of resistance genes, PR-1a, HMGR, PAL. In the previous reports (Park *et al* 2001, Jeun *et al* 2001, Park *et al.*, 2000, Ahn *et al.*, 2002), *B. amyloliquefaciens* strain EXTN-1 showed various beneficial effects on crops and mode of actions were also proposed for these phenomenon. Among the mechanisms, EXTN-1 provoked the expression of two representative markers, PR-1 and PDF 1.2, in the leaves of Arabidopsis ecotype Col-0 at the same time. This result implied that protection ability induced by EXTN-1 is dependent on salicylic acid and/or jasmonic acid-dependent pathways (Ahn *et al.*, 2002, Fig 2).

Six kinds of Cyclo dipeptide {cyclo(L-tyro-L-pro)} were identified as elicitors inducing systemic resistance, which were purified from butanol extract of EXTN-1 grown on TSA medium. Cyclo dipeptide isolated from EXTN-1 showed induced systemic resistance against cucumber anthracnose fungus as well as PR-1a promoter expression on tobacco plant (Park *et al.*, 2002, Table 1). This result suggests that a bacterial metabolite, cyclo (L-pro-L-tyr) involves in the activation of plant defense reactions, leading to systemic resistance against cucumber anthracnose fungi. Cyclic dipeptide, cyclo (L-pro-L-tyr) was isolated from *Pseudomonas putida* WCS358 (Degrassi, 2002) as a quorum sensing signal compound.

On the other hand, several of cyclic peptides are also produced by the fungal plant pathogen *Alternaria alternate*. Furthermore, cyclo (L-pro-L-tyr) and cyclo (L-pro-L-the) act as host-specific phytotoxins against spotted knapweed (*Centaurea maculosa*; Stierle *et al* 1988). The physiological or ecological role of these molecules in relation to quorum sensing remains to be investigated. For future work, it may need whether this compound acts as a quorum sensing signal molecule or has other biological functions in our system.

Mechanism involved in induced systemic resistance by EXTN-1 was revealed as simultaneous activation of SA and JA or

ethylene metabolic pathways and pre-treatment of EXTN-1 reduced germination and appressorium formation of conidia of *Colletotrichum orbiculare* on the leaf of cucumber with increase of callus formation. Furthermore, treatment of EXTN-1 promoted growth and quality of paprika grown in cool season with consistency of the effects and inhibited the bacterial wilt

on tomato caused by *Ralstonia solanacearum* for 4 weeks after treatment. Treatment of *B. amyloliquefaciens* strain EXTN-1 showed the increased plant height of the three barley varieties and shorten heading stage of two varieties compared with non-treated control.

Table 1. Induce systemic resistance and PR-1a promoter expression in cucumber plant by cyclo (L-tyr-L-pro) from EXTN-1

| Treatment | Plant Height (cm) | Stem diameter(mm) | Lesion No / Plant | PR-1aGUS activity (nM MU/10mg F.W/h) |
|---------------------------|-------------------|-------------------|-------------------|--------------------------------------|
| Water | 26.18 | 5.08 | 240.33 | 54.0 |
| BTH 0.1mM | 26.94 | 5.09 | 58.33* | 210,153* |
| Cyclo dipeptide 1.0ppm | 26.27 | 4.70 | 153.33* | 7,497 |
| Cyclo dipeptide 0.1 ppm | 26.68 | 5.05 | 157.11* | 34,967* |
| Cyclo dipeptide 0.001 ppm | 27.18 | 4.87 | 180.22* | 58,167* |
| LSD ($p=0.05$) | 1.07 | 0.20 | 48.31 | 33,267 |

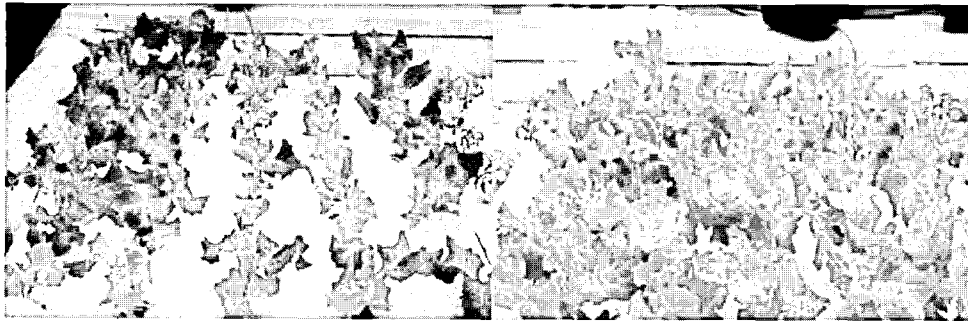


Fig 1. Plant growth promotion by treatment of *B. amyloliquefaciens* EXTN-1 in hydroponic.

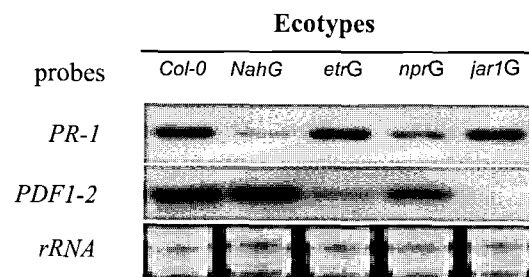


Fig. 2. Activation of Arabidopsis PR-1 and PDF1.2 in response to pretreatment with *B. amyloliquefaciens* EXTN-1.