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The role of Na⁺-Ca²⁺ exchange on calcium activated chloride current in single isolated cardiac myocyte in pulmonary vein of rabbit.

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We have shown the Ca²⁺-activated chloride current is present in cardiac myocyte in rabbit pulmonary vein (Kim et al., 2002). This current amplitude was increased as [Na⁺], was increased and we suggested this chloride current may be involve in the spontaneous action potential frequency change. Since this current is activated by the increase of intracellular Ca²⁺, we would like to test what is the inducer of the increase of [Ca²⁺]; between a L-type Ca²⁺-current or a reverse mode of Na⁺-Ca²⁺ exchange current. White rabbit (1.5 kg) was used and anesthetized with Ketamin (100 mg/kg). Pulmonary vein (PV) was isolated and sleeve area between left atrium and PV was dissected. Using collagenase (Worthington 0.7 mg/cc), single cardiac myocytes were isolated. In the presence of 15 mM of Na⁺, three steps of voltage pulses were applied (holding potential: -40 mV, -80 mV for 50 msec, 30 mV for 5 msec, 10 mV steps from -70 mV to 60 mV). The inward and outward tail current was activated after brief 5 msec prepulse. The outward tail current was blocked by the removal of extracellular chloride substituted by glucuronic acid or by a chloride channel blocker, 5 mM 9-AC. But the inward tail current was still remained even though the amplitude was decreased. The reversal potentials were changed to the direction of the change of chloride equilibrium potential (E_{Cl}) but the shift of equilibrium potential was not enough to match to the theoretical equilibrium potential shift. In the presence of L-type Ca²⁺ channel blocker, nifedipine 1 uM, inward tail currents were greatly reduced but the outward current tail currents were still remained. In the presence of Na⁺-Ca²⁺ exchange current blocker, 10 uM KB-R7943, the inward and outward tail currents were blocked almost completely. We tried to test the Ca²⁺ sensitivity of the chloride current with various [Ca²⁺]_i in pipette solution from 100 nM to 1 uM but we failed to activate Ca²⁺-activated chloride currents even though the cell became contracted in the presence of 1 uM Ca²⁺. From these results, we could conclude that the increase of [Ca²⁺]_i to activate the outward Ca²⁺-activated chloride current was mainly induced by the activation of the reverse mode of Na⁺-Ca²⁺ exchanger. But for the increase of [Ca²⁺]_i to activate the inward tail current, L-type Ca²⁺ current may be the major provoking current. Since the cytosolic increase of [Ca²⁺]_i through pipette solution have failed to activate Ca2+-activated chloride current, this chloride current may have very low Ca²⁺ sensitivity or a compartmental increase Ca²⁺ such as in subsarcolemmal space may activate the chloride current. Since there are several reports and models that the increase of Ca²⁺ in subsarcolemmal space would be over several to tens of uM, both possibility may be valid together.

Acknowledgement: This work was supported by grant No. (R01-2000-000-00172-0) from the Korea Science & Engineering Foundation and by the grant (No. IMT2000-C3-3) from the Ministry of Information and Communication.

References

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