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**Construction of Ethanol-inducible Expression Vector and its Expression Analysis in *Arabidopsis* Protoplast**  
Hee-Jung Yuck<sup>1</sup>, Sung-Jin Kim<sup>1</sup>, Youn-Hee Choi<sup>1</sup>, Ji-Seon Baek<sup>1</sup>, Kwang-Woong Lee<sup>C</sup>

*School of Biological Sciences, Seoul National University, Seoul 151-747*

In order to establish ethanol-inducible gene expression system in plant, alcoholic fermentable gene *ADH2* (Alcohol Dehydrogenase 2) promoter of yeast (*Saccharomyces cerevisiae*) and *ADR1* transcription factor were isolated. *ADH2* gene is expressed by the transcription factor, *ADR1* to use alcohol as carbon source. In this experiment, The transcription factor *ADR1* was introduced into pBI121 vector and *ADH2* promoter into pCAMBIA1303 vector. Finally, p35S-*ADR1*-tnos region of pBI121 vector transferred modified pCAMBIA1303 vector and ethanol-inducible expression vector was constructed. To examine the reaction of ethanol-inducible expression vector in *Arabidopsis* protoplast, expression level of  $\beta$ -glucuronidase (*GUS*) gene and green fluorescence protein (*GFP*) was monitored after 2% ethanol was treated to transgenic protoplast. All of the three ethanol-inducible plant expression vectors showed the *GFP* expression after 2% ethanol treatment. Among them, p*ADH2*:p35S fused plant expression vector of those showed the highest *GFP* expression level. The results suggest that the ethanol-inducible plant expression vector works under ethanol treatment.

G216

**Molecular Genetic Analysis of the Brassinosteroids Signaling Mutants**  
Indeok Hwang<sup>P</sup>, Hyeonsook Cheong<sup>C</sup>

*Department of the genetic engineering, Chosun university, Gwangju 501-759*

Brassinosteroids (BRs) are a class of steroid hormones essential for normal plant growth and development. In order to expand our knowledge of the molecular mechanisms of plant steroid signaling, we performed a genetic screening, in which activation-tagged Columbia lines were examined on the medium containing Brassinazole (Brz) in dark. Mutants resistant to a BRs biosynthetic inhibitor, Brassinazole, were shown to have longer hypocotyls than the ones with the wild type background. We identified 3 mutants that have longer hypocotyls and recovered T-DNA flanking sequences from the activation-tagged brassinazole insensitive (*abz*) mutants. They were designated as *abz126*, *abz314*, and *abz453*. These mutants were analyzed for the genetic mapping and characterization. *abz126* is a loss-of-function mutant of *Gigantea (GI)* gene by T-DNA insertion that delayed flowering under long days and displayed longer hypocotyls on the medium containing Brz. This phenotype was similar to the *gi-1* and *gi-2* in aspect of the late flowering and the longer hypocotyls on the medium containing brz in the dark. Genetic analysis of *abz314* indicated that the T-DNA was inserted in the centromere region. *abz314* mutant showed both the longer hypocotyls on the medium containing Brz in dark and the phenotype including longer petiole and accelerated senescence. Further functional repression of the centromere in plants might be elucidated by the mutants analysis. The *abz453* is knock-out mutant of the 24-sterol C-methyltransferase gene (*24-SMF*) in BAC F14O10. RT-PCR result also showed that *FCP* serine phosphatase gene (*FSP*), neighboring gene of *24-SMF*, was over-expressed by the four enhancers.

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**Molecular Genetic Analysis of the *Bril*-suppressor Mutants (*bsu*)**  
Lyounghmae Kim<sup>P</sup>, Hyeonsook Cheong<sup>C</sup>, Indeok Hwang<sup>1</sup>

*Department of the genetic engineering, Chosun University, Gwangju 501-759*

Genetic screens were performed for the identification of brassinosteroid (BRs) signal components using activation tagging *bril-5* in *Arabidopsis*. We selected based on the *bril* suppressed phenotype in brassinazole (Brz) which BRs biosynthesis inhibitor to block specifically at steps from campestanol to teasterone catalyzed cytochrome P450. *bril* suppressor mutants *bsu2* and *bsu7* have reduced sensitivity to Brz and restored dwarfism with longer hypocotyls and having elongated petioles such as a nearly wild type phenotype. *bsu2* and *bsu7* were semidominant and have the longer petioles and swirling leaves. It has been revealed that *BSU2* encodes a cell division related protein kinase *cdc2* and *BSU7* encode upstream region of cytochrome P450 CYP90. Further genetic analysis of these *bril* suppressor mutants and cloning of the tagged genes will elucidate their functions in BRs signaling

G601

**Control of *XIST* Expression by MeCP2 and HDAC**  
Jee-Hye Choi<sup>3</sup>, Yoon Sung Kang<sup>1</sup>, Jin-Sook Son<sup>1</sup>, Na-Young Min<sup>1</sup>, Young-Hoon Park<sup>1</sup>, Kwang-Ho Lee<sup>C</sup>

*Department of Life Science, College of Natural Science, Chung-Ang University, Seoul 156-756*

The *XIST* expression is regulated by differential methylation of CpG sites in the promoter region of the *XIST* gene. The proteins that bind to the differentially methylated *XIST* promoter and mediate transcriptional silencing have not been identified, although a family of proteins containing a methyl-CpG-binding domain is obvious candidate. Previously, we reported that MeCP2 (methyl-CpG-binding protein 2) is recruited to the methylated promoter of the *XIST* on Xa (active X) and can repress the expression of the *XIST* in a methylation-dependent manner. In this study, we observed re-expression of the *XIST* on Xa by treatment with TSA (trichostatin A), an HDAC (histone deacetylase complex) inhibitor. The result indicates that HDAC is required for transcriptional repression of the *XIST* and probably acts as a mediator, which links some regulating proteins including MeCP2 for transcriptional repression. In addition, to ascertain whether the *XIST* is re-expressed by dysfunction of MeCP2, we analysed the expression pattern of transcriptionally inert *XIST* on Xa through a functional blocking of MeCP2 using antisense RNA in male lymphoblastoid cells. These results imply that MeCP2 and HDAC participate in *XIST* silencing through chromatin remodeling by methylation of CpG sites and acetylation of histone.