BIOGEOCHEMICAL ACTIVITIES OF MICROORGANISMS IN
MINERAL TRANSFORMATIONS: CONSEQUENCES FOR METAL
AND NUTRIENT MOBILITY

GEOFFREY M. GADD

Division of Environmental and Applied Biology, Biological Sciences Institute, School of Life Sciences, University of Dundee, Dundee, DD1 4HN, Scotland, UK

Abstract

Bacteria and fungi are fundamental biotic components of natural biogeochemical cycles for metals and metalloids and play important roles in dissolution, precipitation, oxidation and reduction processes. Some processes catalyzed by microorganisms also have important applications in environmental biotechnology in the areas of ore leaching and bioremediation.

Introduction

Fungi and bacteria from all major taxonomic groups have been found in metal-rich habitats, such as mineral ores, acidic soils or polluted environments. Survival may depend on a number of biochemical, structural and physiological properties that modify

metal bioavailability and toxicity. Microorganisms have essential requirements for metals such as K, Na, Mg, Ca, Mn, Fe, Cu, Zn, Co and Ni, while some metals, e.g. Rb, Cs, Al, Cd, Ag, Au, Hg and Pb, and metalloids, e.g. As and Te, have no apparent essential function (14). However, all metals and metalloids, can be toxic in excess which has led organisms to develop diverse mechanisms of resistance (30,31). Both genetically-coded mechanisms of resistance and non-specific mechanisms, such as oxidation or reduction of the metals to produce less toxic chemical species, or sorption and precipitation on cell surfaces, can protect cells (14).

Microbial roles in alteration of metal speciation

The ability of microorganisms to affect metal speciation stems from two opposing phenomena: mobilization and immobilization, which influence the balance of metal species between soluble and insoluble phases. Mobilization of metals can be achieved by, e.g. protonation, chelation, and chemical transformation. Metal immobilization can occur by precipitation or crystallization of insoluble organic or inorganic compounds or by sorption, uptake and intracellular sequestration. In addition, redox reactions can mobilize or immobilize metals depending on the metal species involved. The extent to which microorganisms can affect metal contaminants is dependent on the

identity and chemical form of the metal and the physical and chemical nature of the contaminated site or substance. In general terms, microbial processes which solubilize metals increase bioavailability and toxicity, whereas those that immobilize them reduce bioavailability. In bioremediation, solubilization provides a route for removal from solid matrices such as soils, sediments, dumps and industrial wastes (4,7). Alternatively, immobilization processes may enable metals to be transformed *in situ* into insoluble and chemically inert forms and are also particularly applicable to removing metals from mobile aqueous phases (12,13,15,17,23,24).

METAL MOBILIZATION

Fungi and bacteria can facilitate the movement of metals into soluble forms via the dissolution of metal compounds including oxides, phosphates, sulfides and more complex mineral ores, or by desorption from exchange sites on clay minerals or organic matter. A number of mechanisms may be involved in metal mobilization.

Acidification

Fungi and bacteria can acidify their environment by proton efflux via plasma membrane H⁺-ATPases, maintenance of charge balance or as a result of respiratory carbon dioxide

accumulation. Acidification can lead to metal release via a number of obvious routes, e.g. competition between protons and the metal (in a metal-anion complex) leads to protonation of the anion, and results in the release of free metal cations. Organic acids can supply both protons and metal complexing anions (see below) (11,18).

Heterotrophic (chemoorganotrophic) leaching

Heterotrophic metabolism can also lead to leaching as a result of the efflux of protons, organic acids and siderophores. Citrate and oxalate anions can form stable complexes with a large number of metals. Many metal citrates are highly mobile and not readily degraded and the presence of citric acid in soil may enhance metal solubility (9). Oxalic acid can also act as a leaching agent for those metals that form soluble oxalate complexes, including Al and Fe (35). Organic acid production is an important agent of mineral deterioration, playing a role in both biogenic chemical weathering and soil formation (11). Heterotrophic solubilization can have consequences for other remedial treatments for contaminated soils. Pyromorphite (Pb₅(PO₄)₃Cl) is a stable lead mineral and can form in urban and industrially-contaminated soils. Such insolubility reduces lead bioavailability and the formation of pyromorphite has been suggested as a remediation technique for lead-contaminated land, if necessary by means of phosphate addition. However, pyromorphite can be solubilized by phosphate-solubilizing fungi, e.g. A. niger, and plants grown with pyromorphite as a sole phosphorus source accumulated both P and Pb (29). During fungal transformation of pyromorphite, biogenic production of lead oxalate dihydrate can occur (29). This study emphasises the importance of considering microbial processes in developing remediation techniques for metal-contaminated soils.

Autotrophic (chemolithotrophic) leaching

Metals can be leached from solid matrices as a result of autotrophic metabolism. Most autotrophic leaching is carried out by chemolithotrophic, acidophilic bacteria which fix carbon dioxide and obtain energy from the oxidation of ferrous iron or reduced sulphur compounds (3,8). These metabolic processes yield Fe(III) or H₂SO₄ as the respective end-products. The microorganisms involved in autotrotrophic leaching include sulphur-oxidizing bacteria, e.g. *Thiobacillus thiooxidans*, iron- and sulphur-oxidizing bacteria, e.g. *Thiobacillus ferrooxidans* and iron-oxidizing bacteria, e.g. *Leptospirillum ferrooxidans*. As a result of sulphur- and iron-oxidation by these bacteria, metal sulphides are solubilized and the pH of their immediate environment is decreased, which enhances the solubilization of other metal compounds (3,8,45).

Complexation by ligands

Microorganisms produce extracellular metabolites that can complex metals in solution, including polysaccharides, pigments, organic acids (see above) and siderophores (1,2,5,28). The bioavailability of a solubilized metal-ligand complex will depend ultimately on its long-term fate in the environment, including sorption onto soil components or potential utilization by biota (9).

Siderophores

Siderophores are highly specific Fe(III) ligands (formation constants often > 10³⁰). These low-molecular weight coordination molecules are excreted by a wide variety of fungi and bacteria to aid iron assimilation (14). Organisms may have evolved mechanisms for production of species-specific siderophores, or attachment to solid Fe minerals, e.g., Fe oxides, to facilitate contact with the Fe substrate.

Biomethylation

A range of bacteria and fungi under aerobic and anaerobic conditions can mediate methylation of Hg, As, Se, Sn, Te and Pb (22). Methyl groups are enzymatically transferred to the metal, and a given species may transform a number of different

metal(loid)s. Methylated metal compounds formed by these processes differ in their solubility, volatility and toxicity. Volatile methylated species, e.g. (CH₃)₂Se and (CH₃)₂Se₂, can be lost to the atmosphere (10).

Redox Transformations

Microorganisms can mobilize metals, metalloids and organometallic compounds by reduction and oxidation processes (10,20). For example, oxidation of metal-complexing dimethylsulfide, dimethylsulfoxide or thiosulfate increases metal availability if metal sulfates are formed, and the solubilities of Fe and Mn increase on reduction of Fe(III) to Fe(II) and Mn(IV) to Mn(II) (23). Most iron reduction is carried out by specialized anaerobic bacteria that use iron(III) as a terminal electron acceptor. Dissimilatory metalreducing bacteria can use a variety of metal(loid)s with an appropriate redox couple, including Fe(III), Mn(IV), Se(IV), Cr(VI) and U(VI) (25,34). While Fe and Mn increase their solubility upon reduction, the solubility of other metals such as U(VI) to U(IV) and Cr(VI) to Cr(III) decreases, resulting in immobilization (26,32). Reduction of Hg(II) to Hg(0) by bacteria and fungi results in diffusion of elemental Hg out of cells (30,31). Bacteria can also use redox-active organic compounds to shuttle electrons between cells and the metal being respired under anaerobic conditions (23). Many quinoid compounds, common components of humic substances, can function as electron shuttles. These compounds move electrons from the cell membrane to the target metal, such as iron contained in ferric oxides. The shuttle molecule regenerates when the metal is reduced (23).

IMMOBILIZATION

There are a number of processes whereby microorganisms immobilize metals. Although immobilization reduces the external free metal activity, it may also shift the equilibrium to release more metal into the soil solution.

Biosorption

The strong interaction of metals with cell surfaces is a result of cell wall characteristics, including component pigments and polysaccharides (15,16,17). Gram-positive bacterial walls consist of a thick matrix of peptidoglycan and secondary polymers. The phosphate and carboxyl groups within the wall fabric and exposed at the surface confer pH-dependent negative charge to the cell in environments where microbes generally exist; the point of zero charge is approximately pH 2-3. This charge provides the basis for metal sorption (21). Gram-negative cells are also negatively charged, although structurally more complex. In this case, a thin layer of peptidoglycan is located in a gel-like periplasm, sandwiched between the plasma membrane and the outer membrane.

The outer membrane has characteristic surface lipopolysaccharides (LPS) containing ionizable phosphate, carboxyl, hydroxyl and amine groups; the phosphate groups are the most metal reactive. In addition to these structures, surface capsules, sheaths and S-layers provide additional sites for metal interactions and the heterogeneous nucleation of minerals (1,21).

Fungal cell walls are predominantly composed of polysaccharides (80-90 %) with glycoproteins and some lipids. The polysaccharide component includes microcrystalline fibrils of β -linked polysaccharides (chitin, chitosan and β -glucans). As for bacteria, the chemical properties of the functional groups associated with fungal walls, including carboxyl, amine, hydroxyl, phosphate and sulfhydryl groups, provide the basis for the attraction of metals to cell walls. In addition, pigments like melanin have significant metal-binding properties, preventing metal entry into melanized structures (18).

Precipitation of metals and minerals

Cellular precipitation

At sufficient concentrations, metal precipitates nucleate on or within cells, or in the immediate extracellular environment (21). The formation of extracellular and sorbed

precipitates depends on the chemical composition of the extracellular environment and is often mediated by cellular processes. Anions, such as OH, SO₄²⁻, PO₄³⁻ and CO₃²⁻, provide counterions for mineral hydroxides, sulfate and sulfides, phosphates, and carbonates. It is possible that cellular excretion of such anions contributes to these minerals. Metal oxide and hydroxide minerals formed by abiotic reactions also attach to cells. Inside cells, metals may be effluxed, form intracellular precipitates, bind to metal-complexing molecules, and, in eukaryotes, partition into vacuoles (18).

Oxalates

Calcium oxalate is the most common form of oxalate associated with soils and leaf litter, occurring as dihydrate (weddellite) or the more stable monohydrate (whewellite) (11). Calcium oxalate crystals are commonly associated with free-living, pathogenic and plant symbiotic fungi and are formed by the precipitation of solubilized calcium as the oxalate (11,19). The formation of calcium oxalate by fungi has an important influence on biological and geochemical processes in soils, acting as a reservoir for calcium in the ecosystem, and also influencing phosphate availability. Fungi can also produce other metal oxalates with a variety of different metals and metal-bearing minerals, e.g. Cd, Co, Cu, Mn, Sr and Zn (11,29,45).

Carbonates

In many arid and semi-arid regions, calcareous soils and near surface limestones (calcretes) are often secondarily cemented with calcite (CaCO₃). This phenomenon has been partly attributed to physicochemical processes; however, the abundance of calcified fungal filaments in weathered profiles of chalky limestone and Quaternary calcretes indicates fungal activity (11). Mineralized carbonate precipitates are also found in association with bacterial biofilms (21).

Bacterial and fungal oxidation

The range of bacterial species with the capability to oxidize Fe and Mn is large. Bacterial Fe oxidation is ubiquitous in environments with sufficient Fe²⁺ and conditions to support bacterial growth such as drainage waters and tailing piles in mined areas, pyritic and hydric soils (bogs and sediments), drain pipes and irrigation ditches, and plant rhizosphere zones. Iron-oxidizers found in acidic soil environments are acidophilic chemolithotrophs, such as *Thiobacillus ferro oxidans*, significant for its role in generating acid mine drainage (8). Facultative chemolithotrophic microaerophiles such as *Leptothrix ochracea*, *Sphaerotilus natans* and *Gallionella ferruginea* are common in mildly acidic to neutral environments. Fungi also oxidize metals in their

environment. Desert varnish is an oxidized metal layer (patina) a few mm thick found on rocks and in soils of arid and semi-arid regions, and is believed to be of fungal and bacterial origin.

Dissimilatory metal-reduction by bacteria

When dissimilatory metal-reducing bacteria utilize metals such as Cr(VI) and U(VI) as electron acceptors, less soluble Cr(III) and U(IV) species result (23,32,34). Once reduced, metals sorb to the cells or partition to the bulk extracellular solution, where new minerals can form. Fe^{2+} and Mn^{2+} , generated by dissimilatory reduction, can form extracellular precipitates and nodules. Fungi and other microorganisms also precipitate reduced forms of metal species, e.g. Ag(I) to Ag(0); selenate [Se(VI)] and selenite [Se(IV)] to Se(0); tellurite [Te(IV)] to Te(0), by a variety of mechanisms (10,18,20).

Metal precipitation by sulphate-reducing and other bacteria

Sulphate-reducing bacteria (SRB) oxidize organic compounds or hydrogen coupled with the reduction of sulfate, producing sulphide (39,40,41). The solubility products of most heavy metal sulphides are very low, in the range of 4.65×10^{-14} (Mn) to 6.44×10^{-53} (Hg) so that even a moderate output of sulphide can remove metals to levels permitted

in the environment with metal removal being directly related to sulphide production (39,43). Sulphate-reducing bacteria can also create extremely reducing conditions which can chemically reduce metals such as uranium(VI) (43). In addition, sulphate reduction partially eliminates acidity from the system as a result of the shift in equilibrium when sulphate (dissociated) is converted to sulphide (largely protonated). This can result in the further precipitation of metals as hydroxides as well as increasing the efficiency of sulphide precipitation (41). Another form of bacterial bioprecipitation is mediated by a phosphatase enzyme, which liberates inorganic phosphate from a supplied organic phosphate donor molecule, e.g. glycerol 2-phosphate. Metals/radionuclides are precipitated as phosphates on the biomass (24,47).

ENVIRONMENTAL BIOTECHNOLOGY

Ore Leaching

Metals in ores that are not worth recovering by smelting can be effectively leached by bacteria and fungi (3,4,6,8,27). Important species of bacteria are Fe and S oxidizers such as *Thiobacillus ferrooxidans*, *T. thiooxidans* and *Leptospirillum ferrooxidans*. Their activities solubilize Fe and S contained in metal sulfides, with the concomitant release of associated metals such as Cu and Zn. Large amounts of Fe²⁺ and sulfuric acid are generated in the process (8). Industry can take advantage of the acidic leachate by re-circulating it through the ore body to dissolve more metal: soluble metals are then separated by sedimentation, solvents or electrodialysis. When uncontrolled, such as at abandoned mining sites, the same process can have disastrous environmental consequences resulting in acid mine drainage.

Extracellular ligands produced by fungi, including *Aspergillus* and *Penicillium* spp., have also been used to leach metals such as Zn, Cu, Ni and Co from a variety of solid materials, including low-grade mineral ore, solid waste and electronic scrap materials (4,6,11,18,28,35,37,38).

Bioremediation

Some of the processes outlined above have potential for application for treating contaminated land. In a bioremediation context, production of sulphuric acid by *Thiobacillus* species has been used to solubilize metals from sewage sludge, thus enabling separation from the sludge which can then be used as a fertiliser (33). Autotrophic leaching has been used to remediate other metal-contaminated solid materials including soil and red mud, the main waste product of Al extraction from bauxite (38). Although some processes could be used *in situ* (e.g. leaching using Soxidizing bacteria), many are probably most suitable for *ex situ* use in bioreactors, where the mobilized or immobilized metal can be separated from soil components (46). Living or dead fungal and bacterial biomass and metabolites have been used to remove metals and metalloids from solution by biosorption or chelation (14,24).

The increased interest in microbial activities in anaerobic, subsurface environments has opened up new possibilities for the bioremediation of metal contaminants. Metal(loid)s that form insoluble precipitates when reduced may be of particular interest for *in situ* treatment, such as Se(0), Cr(III), Tc(IV) and U(IV) (23,34,36). Natural processes in subsurface soil may immobilize contaminants otherwise predicted to leach. The sulphide produced from sulphate reduction plays a major role in metal sulphide

immobilization in sediments but has also been applied to bioremediation of metals in waters and leachates. Large-scale bioreactors have been developed using bacterial sulphate-reduction for treating metal-contaminated waters. A process integrating bacterial sulphate-reduction with bioleaching by sulphur-oxidizing bacteria has also been developed to remove contaminating toxic metals from soils. Sulphur- and iron-oxidizing bacteria liberated metals from soils in the form of an acid sulphate solution that enabled almost all the metals to be removed by bacterial sulphate reduction (46). SRB biofilm reactors may offer a means of process intensification and entrap or precipitate metals, e.g. Cu and Cd, at the biofilm surface (42,44).

Conclusions

Microorganisms play important roles in the environmental fate of metals, radionuclides, and metalloids with a multiplicity of physico-chemical and biological mechanisms effecting transformations between soluble and insoluble phases. Such mechanisms are important components of natural biogeochemical cycles, with some processes being of potential application to the treatment of contaminated materials

Acknowledgements

The author gratefully acknowledges financial support for his own work from the Biotechnology and Biological Sciences Research Council, the Natural Environment Research Council, and the Royal Societies of London and Edinburgh.

References

- 1. Beech, I.B., and C.W.S. Cheung. 1995. Interactions of exopolymers produced by sulphate-reducing bacteria with metal ions. *Int. Biodeter. Biodeg.* **35**: 59-72.
- 2. Birch, L., and R. Bachofen. 1990. Complexing agents from microorganisms. Experientia 46:827-834.
- 3. Bosecker, K. 1997. Bioleaching: metal solubilization by microorganisms. *FEMS Microbiol. Rev.* **20:**591-604.
- Bosshard, P.P., R. Bachofen, and H. Brandl. 1996. Metal leaching of fly-ash from municipal waste incineration by Aspergillus niger. Environ. Sci. Technol. 30: 3066-3070.
- 5. Bridge, T.A.M., C. White and G.M. Gadd. 1999. Extracellular metal-binding activity of the sulphate-reducing bacterium *Desulfococcus multivorans*. *Microbiol*. **145**: 2987-2995.

- 6. Burgstaller, W., and F. Schinner. 1993. Leaching of metals with fungi. *J. Biotechnol.* 27:91-116.
- 7. Diels L., M. DeSmet, L. Hooyberghs, and P. Corbisier. 1999. Heavy metals bioremediation of soil. *Mol. Biotechnol.* 12:149-158.
- 8. Ewart, D. K., and M. N. Hughes, M. N. 1991. The extraction of metals from ores using bacteria. *Adv. Inorg. Chem.* **36:**103-135.
- 9. Francis, A. J., C. J. Dodge, and J. B. Gillow. 1992. Biodegradation of metal citrate complexes and implications for toxic metal mobility. *Nature* **356:**140-142.
- 10. Gadd, G. M. 1993. Microbial formation and transformation of organometallic and organometalloid compounds. *FEMS Microbiol. Rev.* **11:**297-316.
- Gadd, G. M. 1999. Fungal production of citric and oxalic acid: importance in metal speciation, physiology and biogeochemical processes. *Adv. Microb. Physiol.* 41:47-92.
- 12. Gadd, G. M. 2000. Bioremedial potential of microbial mechanisms of metal mobilization and immobilization. *Curr. Opin. Biotechnol.* **11:**271-279.
- 13. Gadd, G. M. 2000. Heavy metal pollutants: environmental and biotechnological aspects, pp. 607-617. *In J. Lederberg (ed.)*, *The Encyclopedia of Microbiology, 2nd edition*, Academic Press, San Diego.

- 14. Gadd, G.M. 2001. Accumulation and transformation of metals by microorganisms, pp.
 225-264. In H.-J. Rehm, G. Reed, A. Puhler and P. Stadler (ed.), Biotechnology, a

 Multi-volume Comprehensive Treatise, Volume 10: Special Processes, Wiley-VCH

 Verlag GmbH, Weinheim, Germany.
- 15. Gadd, G.M. 2002. Interactions between microorganisms and metals/radionuclides: the basis of bioremediation, pp. 179-203. *In M. J. Keith-Roach and F.R. Livens* (ed.), *Interactions of Microorganisms with Radionuclides*, Elsevier, Amsterdam.
- 16. Gadd, G. M., and C. White. 1990. Biosorption of radionuclides by yeast and fungal biomass. *J. Chem. Technol. Biotechnol.* **49**:331-343.
- 17. Gadd, G. M., and C. White. 1993. Microbial treatment of metal pollution a working biotechnology? *Trends Biotechnol.* 11:353-359.
- 18. Gadd, G. M., and J. A. Sayer. 2000. Fungal transformations of metals and metalloids, pp. 237-256. In D.R. Lovley (ed.), Environmental Microbe-Metal Interactions, Am. Soc. Microbiol., Washington.
- 19. Gharieb, M. M., J. A. Sayer, and G. M. Gadd. 1998. Solubilization of natural gypsum (CaSO₄.2H₂O) and the formation of calcium oxalate by *Aspergillus niger* and *Serpula himantioides*. *Mycol. Res.* **102**:825-830.
- 20. Gharieb, M. M. Kierans, and G. M. Gadd. 1999. Transformation and tolerance

- of tellurite by filamentous fungi: accumulation, reduction and volatilization. *Mycol.*Res. 103:299-305.
- 21. Glasauer, S., E.P. Burford, F.A. Harper, G.M. Gadd, and T.J. Beveridge. 2002.
 Transformation of metals and metalloids by bacteria and fungi. *In D. Hillel, C. Rosenzweig, D. Powlson, K. Scow, M. Singer, and D. Sparks (ed.), Encyclopedia of Soils in the Environment, Academic Press, London (in press).*
- 22. Karlson, U., and W. T. Frankenberger. 1993. Biological alkylation of selenium and tellurium, pp. 185-227. In H. Sigel and A. Sigel, (ed.), Metal Ions in Biological Systems, Marcel Dekker, New York.
- 23. Lovley, D. R. and J. D. Coates. 1997. Bioremediation of metal contamination. *Curr. Opin. Biotechnol.* 8:285-289.
- 24. Macaskie, L. E. 1991. The application of biotechnology to the treatment of wastes produced by the nuclear fuel cycle biodegradation and bioaccumulation as a means of treating radionuclide-containing streams. Crit. Rev. Biotechnol. 11:41-112.
- 25. Oremland, R. S., N. A. Steinberg, T. S. Presser, and L. G. Miller. 1991. In situ bacterial selenate reduction in the agricultural drainage systems of Western Nevada.
 Appl. Environ. Microbiol. 57:615-617.
- 26. Phillips, E.J.P., E. R. Landa, and D. R. Lovley. 1995. Remediation of uranium

- contaminated soils with bicarbonate extraction and microbial U(VI) reduction. *J. Indust. Microbiol.* **14:**203-207.
- 27. Rawlings, D. E., and S. Silver. 1995. Mining with microbes. *Biotechnol*. 13:773-338.
- 28. Sayer, J.A. and G.M. Gadd. 2001. Binding of cobalt and zinc by organic acids and culture filtrates of *Aspergillus niger* grown in the absence or presence of insoluble cobalt or zinc phosphate. *Mycol. Res.* **105**: 1261-1267.
- 29. Sayer, J. A., J. D. Cotter-Howells, C. Watson, S. Hillier, and G. M. Gadd. 1999. Lead mineral transformation by fungi. *Curr. Biol.* **9:**691-694.
- 30. Silver, S. 1996. Bacterial resistances to toxic metal ions a review. Gene 179:9-19.
- 31. Silver, S. 1998. Genes for all metals a bacterial view of the periodic table. J.
 Indust. Microbiol. Biotechnol. 20:1-12.
- 32. Smith, W.L. and G.M. Gadd. 2000. Reduction and precipitation of chromate by mixed culture sulphate-reducing bacterial biofilms. *J. Appl. Microbiol.* **88**, 983-991.
- 33. Sreekrishnan, T. R., and R. D. Tyagi. 1994. Heavy metal leaching from sewage sludges: a techno-economic evaluation of the process options. *Environ. Technol.* 15:531-543.
- 34. Stolz, J. F., and R. S. Oremland. 1999. Bacterial respiration of arsenic and selenium.

- FEMS Microbiol. Rev. 23:615-627.
- 35. Strasser, H., W. Burgstaller, and F. Schinner. 1994. High yield production of oxalic acid for metal leaching purposes by *Aspergillus niger*. *FEMS Microbiol*. *Lett*. 119:365-370.
- 36. Thompson-Eagle, E. T., and W. T. Frankenberger. 1992. Bioremediation of soils contaminated with selenium, pp. 261-309. *In* R. Lal and B.A. Stewart (ed.), *Advances in Soil Science*, Springer-Verlag, New York
- 37. Tzeferis, P. G., S. Agatzini, and E. T. Nerantzis. 1994. Mineral leaching of non sulphide nickel ores using heterotrophic micro-organisms. Lett. Appl. Microbiol. 18:209-213.
- 38. Vachon, P. R. D., Tyagi, J. C. Auclair, and K. J. Wilkinson. 1994. Chemical and biological leaching of aluminium from red mud. *Environ. Sci. Technol.* 28:26-30.
- 39. White, C., and G. M. Gadd. 1996. A comparison of carbon/energy and complex nitrogen sources for bacterial sulphate-reduction: potential applications to bioprecipitation of toxic metals as sulphides. *J. Indust. Microbiol.* 17:116-123.
- 40. White, C., and G. M. Gadd. 1996. Mixed sulphate-reducing bacterial cultures for bioprecipitation of toxic metals: factorial and response-surface analysis of the effects of dilution rate, sulphate and substrate concentration. *Microbiol.* **142:**2197-2205.
- 41. White, C., and G. M. Gadd. 1997. An internal sedimentation bioreactor for

- laboratory-scale removal of toxic metals from soil leachates using biogenic sulphide precipitation. *J. Indust. Microbiol.* **18:**414-421.
- 42. White, C., and G. M. Gadd. 1998. Accumulation and effects of cadmium on sulphate-reducing bacterial biofilms *Microbiol*. **144:**1407-1415.
- 43. White, C., and G. M. Gadd. 1998. Reduction of metal cations and oxyanions by anaerobic and metal-resistant organisms: chemistry, physiology and potential for the control and bioremediation of toxic metal pollution, pp. 233-254. *In* W.D. Grant and T. Horikoshi (ed.), *Extremophiles: Physiology and Biotechnology*, John Wiley and Sons, New York.
- 44. White, C., and G. M. Gadd. 2000. Copper accumulation by sulphate-reducing bacterial biofilms and effects on growth. *FEMS Microbiol. Lett.* **183**: 313-318.
- 45. White, C., J.A. Sayer, and G. M. Gadd. 1997. Microbial solubilization and immobilization of toxic metals: key biogeochemical processes for treatment of contamination. FEMS Microbiol. Rev. 20:503-516.
- 46. White, C., A. K. Sharman, and G. M. Gadd. 1998. An integrated microbial process for the bioremediation of soil contaminated with toxic metals. *Nature Biotechnol.* **16:**572-575.
- 47. Yong, P., and L. E. Macaskie. 1995. Enhancement of uranium bioaccumulation by a Citrobacter sp. via enzymically-mediated growth of polycrystalline NH₄UO₂PO₄. J. Chem. Technol. Biotechnol. 63:101-108.