

antithrombotic agents *in vitro* and *ex vivo*, and its antithrombotic effect *in vivo*. Chungpesagan-tang, Rhei Rhizoma and Scutellariae Radix potently inhibited ADP-, collagen-induced rat platelet aggregation in a dose-dependent manner *in vitro*. Chungpesagan-tang and most of its ingredients did not affect such coagulation parameters as APTT, PT and TT in human plasma. However, Rhei Rhizoma potently protected plasma clotting. Chungpesagan-tang treated group was significantly inhibited platelet aggregation in orally administered rats *ex vivo*. Puerariae Radix and Rhei Rhizoma also showed significant prolongation of tail bleeding time in conscious mice. Chungpesagan-tang showed significantly protection from death due to pulmonary thrombosis in mice.

[PC2-5] [10/20/2000 (Fri) 15:30 – 16:30 / [Hall B]]

Purification and Characterization of novel salt-inducible heparinase II from *Bacteroides stercoris* HJ-15

Hong SW^{0,1}, Kim BT¹, Kim DH¹, Kim YS²

^{0,1}College of Pharmacy, Kyung Hee University, ²Natural Products Research Institute, Seoul National University

Bacteroides stercoris HJ-15, which is human colon gram-negative rod cell, has been known to degrade heparin, acharan sulfate and chondroitin sulfate. Recently we found to produce new five heparinases: one heparinase I, three heparinase II, one heparinase III. Among three heparinase II, one is a salt-inducible enzyme. In the present study, we tried to purify the enzyme. *Bacteroides stercoris* HJ-15 was cultured in 100L of tryptic soy broth containing hepain, collected, sonicated and centrifuged at 18,000rpm for 70min at 4°C. From the supernatant, the salt-inducible heparinase II was purified to homogeneity by QAE-cellulose, DEAE cellulose, CM Sephadex C-50, hydroxyapatite and phosphocellulose. The specific activity of the purified heparinase II was 81.3 $\mu\text{mole}/\text{mg}$. Its molecular weight was 94KDa. The optimal activity was observed to be pH 7.3 containing 50mM KCl. Cu^{+2} , Ni^{+2} , Co^{+2} , Fe^{+2} and Zn^{+2} inhibited the enzyme activity. These findings suggest that the biochemical properties of the purified enzyme should be different from the previously purified heparinase II.

[PC2-6] [10/20/2000 (Fri) 15:30 – 16:30 / [Hall B]]

Purification and Characterization of *Staphylococcus epidermidis* Urease

Min SH⁰ and Lee MH

Microbiology Lab., College of Pharmacy, Catholic University of Taegu

The multi-subunit urease from *Staphylococcus epidermidis* ATCC12228 was purified 1,127-fold by ion exchange and gel filtration chromatography. The enzyme activity was highest when the cells were grown up to late exponential phase in LB supplemented with 50 μM nickel chloride.

The specific activity of the purified urease was 993.8 μmol urea degraded min^{-1} . The enzyme was shown to have a M_r of 409,000 and consist of three different subunits of 73.3, 22.7, and 13.8 kDa. The Michaelis constant (K_m) was 8.9 mM urea. Activity gel-staining of the native urease with urea and cresol red revealed two major peptide bands.

[PC2-7] [10/20/2000 (Fri) 15:30 – 16:30 / [Hall B]]

Expression control of the *dnak* operon in *Streptococcus pneumoniae*