Epigenetic Status of Genes on Imprinted Regions Jae Hoon Chung

Department of Biological Sciences, Korea Advanced Institute of Sciences and Technology, Taejon, 305-701 Korea

Genomic imprinting is a mammalian epigenetic mechanism marking gametic or zygotic genome leading to parent-of-origin-specific differential expression of the two alleles of a gene in somatic cells of the offspring. Such an epigenetic modification makes an animal functionally hemizygous for the imprinted genes and requires strict contribution of both parental genomes for the normal development of the progeny. The first convincing evidence for genomic imprinting came from nuclear transplantation experiments, which showed that development did not proceed properly in embryos derived from two maternal or two paternal nuclei, suggesting that the expression patterns of each parental genome are different and complementary (McGrath and Solter, 1984). Analysis of mice carrying partial uniparental disomies (UPDs) further demonstrated that there are unique chromosomal regions that can generate a parent-of-origin-specific phenotypes (Cattanach and Kirk, 1985). A certain part of chromosomes gave rise to embryonic lethality or abnormal development in the progeny when it was inherited exclusively from paternally or maternally. Partial UDPs were created by mating mice which carry an appropriate set of Robertsonian or reciprocal translocations in the chromosomes. Studies of UPDs also identified chromosomal regions which required both parental contributions in the offspring for the normal development. The regions identified by this manner were used as landmarks of candidate loci where an individual imprinted gene could be localized.

Imprinted genes were identified serendipitously (Mash2; Guillemot, 1995) or depending on the information including the map position of genes and their relevance to chromosomal region implicated in UPD studies (Igf2r, Snrpn, Ipw, Znf127 etc.) in the early 1990s. Recently, several imprinted genes have been identified through genome-wide systematic searches for monoallelically expressed genes. Differential display and differential cDNA screens of RNAs expressed by androgenetic or parthenogenetic embryos have successfully resulted in the identification of new imprinted genes on chromosomes 6 and 7, Peg1/Mest and Peg3, respectively (Kaneko-Ishino et al., 1995; Kuroiwa et al., 1996). Alternative systematic search for the imprinted genes was devised to take advantage of the fact that imprinted genes have differentially methylated region in their CpG clusters. This approach exploiting restriction landmark genomic scanning, or RLGS, has identified at least two imprinted genes, U2afbp-rs and Grf1 (Hayashizaki et al., 1994; Plass et al., 1996).

To date, about 30 imprinted genes have been identified in human and mouse. It is estimated that there are hundreds of imprinted genes to be identified. One of the major characteristics of imprinted genes is that they are clustered within small regions of chromosomes. Of the 27 imprinted

genes of mice, 11 exist in close proximity to imprinted genes. The largest cluster of imprinted genes is found at the distal end of mouse chromosome 7 (Caspary, 1998) and at the proximal end of human chromosome 11p15.5. Both regions show a clear synteny to each other. In the region at the distal end of mouse chromosome 7 spanning 1.5 Mb, 8 imprinted genes have been identified. It includes *Ipl, Impt1*, p57^{KIP2}, Kvlqt1, Mash2, Ins2, Igf2 and H19 (Dao et al., 1998). It is also well known that cluster of imprinted genes are located at human chromosome 15, where both the Prader-Willi and Angelman syndromes have been mapped (LaSalle and Lalande, 1995). This region contains 3 genes expressed paternally.

These features of clustering suggest the possibility that the mechanism of imprinting is not local or gene specific, but rather functions over long distances. Based on this tendency of clustering, it is plausible that more imprinted genes can be identified in the region which was known to contain few imprinted gene to date, not a cluster of imprinted ones. An imprinted gene, Peg1/Mest is selected for this study because it was mapped to a locus which is known to be implicated in UPD-related embryonic lethality and information of many genes around the locus in mice and humans is available through public databases.

Peg1/Mest, a maternally imprinted gene isolated by the subtractive hybridization between parthenogenetic and normal embryos, is the first imprinted gene identified on mouse chromosome 6. It functions as the mouse mesoderm-specific transcript that shares 70% sequence homology between mouse and human (Nishita et al., 1996). Peg1/Mest is mapped to the sub-proximal end of mouse chromosome 6 and its human homolog is localized in human chromosome 7q32, which is analogous to the mouse chromosomal region of Peg1/Mest. The sub-proximal region of mouse chromosome 6 has been known to be involved in UPD-related embryonic lethality in mice.

Thus, the region may have other imprinted genes close to Peg1/Mest and a search for new imprinted genes on this locus was tried in this study. Investigation of allele-specific expression was performed within the mouse chromosomal region homologous to human locus covering PEG1. Mouse genes and ESTs of which human homologs were mapped near the human PEG1 locus were subject to the investigation of parent-of-origin specific expression. In addition, Peg3 region on chromosome 7, Meg1/Grb10 region on chromosome 11, and Peg5/Nnat region on chromosome 2 were also investigated for the possible presence of novel imprinted genes. Polymorphisms of the genes and ESTs mapped to this regions in mouse and human were identified and used to physically discriminate one of parental alleles from the other in the F1 hybrids between two inbred mouse strains, Mus musculus molossinus and Mus musculus domesticus (C57BL/6J), Restriction fragment length polymorphism analyses of RT-PCR products were carried out to investigate the allele-specific expression of genes and ESTs in the F1 hybrids. Calu, Ube2h, Cappa2 genes, and a cDNA homologous to human STS Cda15a02 within Peg1/Mest region showed biallelic expression patterns without allelic expression bias. Other candidates, A2a/Emap2, A2b, A2e, P3a and P3b in the Peg3 region, Rbl1 within Peg5/Nnat region, M1a and M1b within Meg1/Grb10 region, were shown to be expressed biallelically. However, a cDNA derived from one of the mouse ESTs, tentatively named CopG2, was obviously shown to be expressed only from the paternal allele in the brain of F1 hybrids. Thus, CopG2 is maternally imprinted in the adult mouse brain. Expression pattern in the reciprocal cross unambiguously confirmed the maternal imprinting of CopG2 gene. Southern hybridization using the 3' sequence of CopG2 cDNA as a probe indicated that the CopG2 gene is present as a single copy in the mouse genome. Imprinting of CopG2 seems to be delicately regulated in a tissue-specific manner. In adult heart and lung in which CopG2 is less transcribed than in brain as monitored by Northern hybridization, imprinting status of CopG2 was relaxed. The extent of relaxation was quantitatively analyzed. It was estimated that the percentage of C57BL/6J allele was 93% for F1 hybrid of $(M \times C)$ in heart, 29% for hybrid of $(C \times M)$ in heart, 86% for hybrid of $(M \times C)$ in lung, and 50% for hybrid of $(C \times M)$ in lung. CopG2 provides an example of endogenous gene that shows relaxation of imprinting in several tissues. Such a relaxed imprinting of CopG2, along with Northern data, indicates that the function of this gene is regulated in a tissue-specific imprinting manner.

REFERENCES

Caspary T, Cleary MA, Baker CC, Guan XJ and Tilghman SM: Mol Cell Biol 1998, 18, 3466-3474.

Cattanach BM and Kirk M: Nature 1985, 315, 496-498.

Dao D, Frank D, Qian N, O'Keefe D, Vosatka RJ, Walsh CP and Tycko B: Hum Mol Genet 19987, 597-608.

Guillemot F, Caspary T, Tilghman SM, Copeland NG, Gilbert DJ, Jenkins NA, Anderson DJ, Joyner AL, Rossant J and Nagy A: *Nat Genet* 1995, 9, 235-242.

Hayashizaki Y, Shibata H, Hirotsune S, Sugino H, Okazaki Y, et al: Nat Genet 1994, 6, 33-40.

Kaneko-Ishino T, Kuroiwa Y, Miyoshi N, Kohda T, Suzuki R, Yokoyama M, Viville S, Barton SC, Ishino F and Surani MA: *Nat Genet* 1995, 11, 52-59.

Kuroiwa Y, Kaneko-Ishino T, Kagitani F, Kohda T and Li L-L: Nat Genet 1996, 12, 186-190.

LaSalle MJ and Lalande M: Nat Genet 1995, 9, 386-394.

McGrath J and Solter D: Cell 1984, 37, 179-183.

Nishita Y, Yoshida I, Sado T and Takagi N: Genomics 1996, 36, 539-542.

Plass C, Shibata H, Kalcheva I, Mullins L, Kotelevtseva N, et al: Nat Genet 1996, 14, 106-112.