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Identification of the Genes Involved in Stationary-Phase Specific Acid Tolerance Response of *Salmonella typhimurium*

방 일수\*, John W. Foster<sup>1</sup>, 이 영록, 박 용근  
고려대학교 이과대학 생물학과

<sup>1</sup>Dept. of Microbiology & Immunology, Univ. of South Alabama

*Salmonella* can experience and survive at severe acid stress during its natural or pathogenic life cycle. Entering to stationary growth phase, that organism is much more resistant to acid(1000-fold more than log-phase cell), and has specific acid tolerant system different from log-phase cells'(Foster et al., J. Bacteriol. 176:1422-1426). As part of on going investigation of stationary-phase specific acid resistance, we have searched for *spatr* mutations in virulent *S. typhimurium* UK-1 using the MudJ fusion technique and two lethal selection procedures including DNP(dinitrophenol) selection media and microtiter-plate selection method. Five acid sensitive mutations have been identified and designated *spatr* k1, *spatr* k2, *spatr* k3, *spatr* k4, *spatr* k5. These mutations removed both stationary-phase acid tolerant effect and stationary-phase specific acid resistance. Most of stationary-phase regulatory genes were tested for participating on stationary-phase acid tolerant response. Non-specific histone like protein, H-NS and stationary-phase specific sigma factor, RpoS showed little contribute to that system at respective single mutation(5-10 fold decrease). But, when both mutations were combined together, no acid resistance was achieved. Futhermore, the acid resistance of *hns* mutant decreased in direct proportion to the loss of RpoS activity.

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Cloning and Expression of Anaerobiosis- and pH-regulatory Genes, *oxrG* in *Salmonella typhimurium*.

김정철\*, 방일수, 이영록, 박용근.  
고려대학교 이과대학 생물학과

*OxrG* eliminates log-phase specific acid tolerance response and acts as a positive regulator for three anaerobiosis and low-pH inducible genes(*anic*, *aniI* and *aciK*), and *oxrG* was mapped 88min in *Salmonella typhimurium* (Foster et al., Microbiology 140 (2):341-352 1994). The transcription of three operon fusions requires tyrosine as a coinducer. By using the library of *S. typhimurium* chromosomal DNA, we cloned *oxrG* in about 7.5kb. This recombinant plasmid, pFW53 complemented *oxrG*-Tn10 mutation on minimal-tyrosine and rich medium. We amplified and extracted plasmid coded proteins by chloramphenicol inhibition method. And the periplasmic portion and cytoplasmic portion of extracted proteins were subjected to SDS-PAGE. Finally, specific band for *oxrG* was identified in the periplasmic portion, and was strongly induced by tyrosine and anaerobiosis. These results suggest that *oxrG* acts as a transducer of signals including anaerobiosis and tyrosine in *S. typhimurium*.